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Phytases (*myo*-inositol hexakisphosphate phosphohydrolases; EC 3.1.3.8) are  
5 enzymes that hydrolyze phytate (*myo*-inositol hexakisphosphate) to *myo*-inositol and  
inorganic phosphate and are known to be valuable feed additives.

A phytase was first described in rice bran in 1907 [Suzuki et al., Bull. Coll. Agr.  
Tokio Imp. Univ. 7, 495 (1907)] and phytases from *Aspergillus* species in 1911 [Dox and  
Golden, J. Biol. Chem. 10, 183-186 (1911)]. Phytases have also been found in wheat bran,  
10 plant seeds, animal intestines and in microorganisms [Howsen and Davis, Enzyme Microb.  
Technol. 5, 377-382 (1983), Lambrechts et al., Biotech. Lett. 14, 61-66 (1992), Shieh and  
Ware, Appl. Microbiol. 16, 1348-1351 (1968)].

The cloning and expression of the phytase from *Aspergillus niger* (ficcum) has been  
described by Van Hartingsveldt et al., in Gene, 127, 87-94 (1993) and in European Patent  
15 Application, Publication No. (EP) 420 358 and from *Aspergillus niger* var. *awamori* by  
Piddington et al., in Gene 133, 55-62 (1993).

Cloning, expression and purification of phytases with improved properties have been  
disclosed in EP 684 313. However, since there is a still ongoing need for further improved  
phytases, especially with respect to their thermostability, it is an object of the present  
20 invention to provide the following process which is, however, not only applicable to  
phytases.

A process for the preparation of a consensus protein, whereby such process is  
characterized by the following steps:

- 25 a) at least three, preferably four amino acid sequences of a defined protein family  
are aligned by any standard alignment program known in the art;
- b) amino acids at the same position according to such alignment are compared  
regarding their evolutionary similarity by any standard program known in the  
art, whereas the degree of similarity provided by such a program which defines  
the least similarity of the amino acids that is used for the determination of an  
30 amino acid of corresponding positions is set to a less stringent number and the  
parameters are set in such a way that it is possible for the program to determine

- 5 from only 2 identical amino acids at a corresponding position an amino acid for the consensus protein; however, if among the compared amino acid sequences are sequences that show a much higher degree of similarity to each other than to the residual sequences, these sequences are represented by their consensus sequence determined as defined in the same way as in the present process for the consensus sequence of the consensus protein or a vote weight of 1 divided by the number of such sequences is assigned to every of those sequences;
- 10 c) in case no common amino acid at a defined position can be identified by the program, any of the amino acids of all sequences used for the comparison, preferably the most frequent amino acid of all such sequences is selected or an amino acid is selected on the basis of the consideration given in Example 2;
- d) once the consensus sequence has been defined, such sequence is back-translated into a DNA sequence, preferably using a codon frequency table of the organism in which expression should take place;
- 15 e) the DNA sequence is synthesized by methods known in the art and used either integrated into a suitable expression vector or by itself to transform an appropriate host cell;
- f) the transformed host cell is grown under suitable culture conditions and the consensus protein is isolated from the host cell or its culture medium by
- 20 methods known in the art.

In a preferred embodiment of this process step b) can also be defined as follows:

25 b) amino acids at the same position according to such an alignment are compared regarding their evolutionary similarity by any standard program known in the art, whereas the degree of similarity provided by such program is set at the lowest possible value and the amino acid which is the most similar for at least half of the sequences used for the comparison is selected for the corresponding position in the amino acid sequence of the consensus protein.

In another preferred embodiment the consensus sequence is used in order to improve

30 a specific protein. In this process first a consensus sequence is determined from a number of highly homologous sequences according to steps a), b) and c) as described above. In a second step the amino acid sequence of another protein which is homologous to the consensus sequence is compared with the consensus sequence and in a third step only those amino acid residues are replaced in the amino acid sequence of the other protein

which clearly differ from the consensus sequence of this protein family calculated under moderately stringent conditions whereas at all positions of the alignment where no preferred single amino acid can be determined under moderately stringent conditions the amino acids of the other protein remain unchanged.

5 By using this preferred embodiment the consensus sequence derived from a number of highly homologous sequences is used in order to replace only certain amino acid residues in the protein in such a manner that only those amino acid residues are replaced which clearly and unambiguously differ from the corresponding consensus sequence of this protein family which has been calculated on moderately stringent conditions. At all other  
10 positions of the alignment, however, where the method of the present invention is not able to determine clearly a preferred amino acid residue under moderately stringent conditions the amino acid residues of the other protein are maintained unchanged.

A further preferred embodiment is a process wherein at first a consensus sequence is determined from homologous sequences as described above. In a second step the active  
15 center of the protein comprising all amino acid residues that are involved in forming the active center is determined in the consensus sequence and in the sequence of a single homologous protein as well. The single homologous protein may have preferred properties like high specific activity or different pH dependency of enzymatic activity. In a third step some or all amino acid residues that are involved in forming the active centre of the  
20 homologueous protein are inserted into the backbone of the consensus sequence. The result thereof is a chimeric protein having the active centre derived from a single protein and the backbone of the consensus sequence.

The active centre of the protein can be determined e.g. by using any analysis of the three-dimensional structure of the protein, e.g. by homology modelling on the basis of a  
25 known 3D-structure of a known protein. Frequently the single homologueous protein is an enzyme.

It is furthermore an object of the present invention to provide such a process, wherein the program used for the comparison of amino acids at a defined position regarding their evolutionary similarity is the program "PRETTY". It is more specifically an  
30 object of the present invention to provide such a process, wherein the defined protein family is the family of phytases, especially wherein the phytases are of fungal origin.

It is furthermore an object of the present invention to provide such processes, wherein the host cell is of eukaryotic, especially fungal, preferably *Aspergillus* or yeast, preferably *Saccharomyces* or *Hansenula* origin.



It is also an object of the present invention to provide a consensus protein obtainable preferably obtained, by such processes and specifically the consensus protein, which has the amino acid *sequences shown in Figures 2, 4 and 6* or a variant thereof. A "variant" refers in the context of the present invention to a consensus protein with amino acid  
5 sequence shown in Figure 2, 5, 7, and 8 wherein at one or more positions amino acids have been deleted, added or replaced by one or more other amino acids with the proviso that the resulting sequence provides for a protein whose basic properties like enzymatic activity (type of and specific activity), thermostability, activity in a certain pH-range (pH-stability) have not significantly been changed. "Significantly" means in this context that a man  
10 skilled in the art would say that the properties of the variant may still be different but would not be unobvious over the ones of the consensus protein with the amino acid sequence of Figure 2 itself.

A "mutein" refers in the context of the present invention to replacements of the  
15 amino acid in the amino acid sequences of the consensus proteins shown in Figure 2 which lead to consensus proteins with further improved properties e.g. activity. Such muteins can be defined and prepared on the basis of the teachings given in European Patent Application number 97810175.6, e. g. Q50L, Q50T, Q50G, Q50L-Y51N, or Q50T-Y51N. "Q50L" means in this context that at position 50 of the amino acid sequence (Figure 2) the amino  
20 acid Q has been replaced by amino acid L.

In addition, a food, feed or pharmaceutical composition comprising a consensus protein as defined above is also an object of the present invention.

In this context "at least three preferably four amino acid sequences of such defined protein family" means that three, four, five, six to 12, 20, 50 or even more sequences can  
25 be used for the alignment and the comparison to create the amino acid sequence of the consensus protein. "Sequences of a defined protein family" means that such sequences fold into a three dimensional structure, wherein the alpha-helices, the beta-sheets and beta-turns are at the same position so that such structures are, as called by the man skilled in the art, largely superimposable. Furthermore these sequences characterize proteins which show the  
30 same type of biological activity, e.g. a defined enzyme class, e.g. the phytases. As known in the art, the three dimensional structure of one of such sequences is sufficient to allow the modelling of the structure of the other sequences of such a family. An example, how this can be effected, is given in the Reference Example of the present case. "Evolutionary similarity" in the context of the present invention refers to a scheme which classifies amino  
35 acids regarding their structural similarity which allows that one amino acid can be replaced by another amino acid with a minimal influence on the overall structure, as this is done e.g.

by programs, like "PRETTY", known in the art. The phrase "the degree of similarity provided by such a program...is set to less stringent number" means in the context of the present invention that values for the parameters which determine the degree of similarity in the program used in the practice of the present invention are chosen in a way to allow the  
5 program to define a common amino acid for a maximum of positions of the whole amino acid sequence, e. g. in case of the program PRETTY a value of 2 or 3 for the THRESHOLD and a value of 2 for the PLURALITY can be choosen. Furthermore, "a vote weight of one divided by the number of such sequences" means in the context of the present invention that the sequences which define a group of sequences with a higher  
10 degree of similarity as the other sequences used for the determination of the consensus sequence only contribute to such determination with a factor which is equal to one divided by a number of all sequences of this group.

As mentioned before should the program not allow to select the most similar amino acid, the most frequent amino acid is selected, should the latter be impossible the man  
15 skilled in the art will select an amino acid from all the sequences used for the comparison which is known in the art for its property to improve the thermostability in proteins as discussed e.g. by

Janecek, S. (1993), *Process Biochem.* 28, 435-445 or

Fersht, A. R. & Serrano, L. (1993), *Curr. Opin. Struct. Biol.* 3, 75-83.

20 Alber, T. (1989), *Annu. Rev. Biochem.* 58, 765-798 or

Matthews, B. W. (1987), *Biochemistry* 26, 6885-6888.

Matthews, B. W. (1991), *Curr. Opin. Struct. Biol.* 1, 17-21.

The stability of an enzyme is a critical factor for many industrial applications. Therefore, a lot of attempts, more or less successful, have been made to improve the  
25 stability, preferably the thermostability of enzymes by rational (van den Burg *et al.*, 1998) or irrational approaches (Akanuma *et al.*, 1998). The forces influencing the thermostability of a protein are the same as those that are responsible for the proper folding of a peptide strand (hydrophobic interactions, van der Waals interactions, H-bonds, salt bridges, conformational strain (Matthews, 1993). Furthermore, as shown by Matthews *et al.* (1987),  
30 the free energy of the unfolded state has also an influence on the stability of a protein. Enhancing of protein stability means to increase the number and strength of favorable interactions and to decrease the number and strength of unfavorable interactions. It has been possible to introduce disulfide linkages (Sauer *et al.*, 1986) to replace glycine with

alanine residues or to increase the proline content in order to reduce the free energy of the unfolded state (Margarit et al, 1992; Matthews, 1987a). Other groups concentrated on the importance of additional H-bonds or salt bridges for the stability of a protein (Blaber et al, 1993) or tried to fill cavities in the protein interior to increase the buried hydrophobic surface area and the van der Waals interactions (Karpusas et al, 19898). Furthermore, the stabilization of secondary structure elements, especially  $\alpha$ -helices, for example, by improved helix capping, was also investigated (Munoz & Serrano, 1995).

However, there is no fast and promising strategy to identify amino acid replacements which will increase the stability, preferably the thermal stability of a protein. Commonly, the 3D structure of a protein is required to find locations in the molecule where an amino acid replacement possibly will stabilize the protein's folded state. Alternative ways to circumvent this problem are either to search for a homologous protein in a thermo- or hyperthermophile organism or to detect stability-increasing amino acid replacements by a random mutagenesis approach. This latter possibility succeeds in only  $10^3$  to  $10^4$  mutations and is restricted to enzymes for which a fast screening procedure is available (Arase et al, 1993; Risse et al, 1992). For all these approaches, success was variable and unpredictable and, if successful, the thermostability enhancements nearly always were rather small.

Here we present an alternative way to improve the thermostability of a protein. Imanaka et al (1986) were among the first to use the comparisons of homologous proteins to enhance the stability of a protein. They used a comparison of proteases from thermophilic with homologous ones of mesophilic organisms to enhance the stability of a mesophilic protease. Serrano et al (1993) used the comparison of the amino acid sequences of two homologous mesophilic RNases to construct a more thermostable Rnase. They mutated individually all of the residues that differ between the two and combined the mutations that increase the stability in a multiple mutant. Pantoliano et al (1989) and, in particular, Steipe et al (1994) suggested that the most frequent amino acid at every position of an alignment of homologous proteins contribute to the largest amount to the stability of a protein. Steipe et al (1994) proved this for a variable domain of an immunoglobulin, whereas Pantoliano et al (1989) looked for positions in the primary sequence of subtilisin in which the sequence of the enzyme chosen to be improved for higher stability was singularly divergent. Their approach resulted in the replacement M50F which increased the  $T_m$  of subtilisin by 1.8 °C.

Steipe et al. (1994) proved on a variable domain of immunoglobulin that it is possible to predict a stabilizing mutation with better than 60% success rate just by using a statistical method which determines the most frequent amino acid residue at a certain position of this domain. It was also suggested that this method would provide useful results

not only for stabilization of variable domains of antibodies but also for domains of other proteins. However, it was never mentioned that this method could be extended to the entire protein. Furthermore, nothing is said about the program which was used to calculate the frequency of amino acid residues at a distinct position or whether scoring matrices were  
5 used as in the present case.

It is therefore an object of the present invention to provide a process for the preparation of a consensus protein comprising a process to calculate an amino acid residue for nearly all positions of a so-called consensus protein and to synthesize a complete gene from this sequence that could be expressed in a pro- or eukaryotic expression system.

10 DNA sequences of the present invention can be constructed starting from genomic or cDNA sequences coding for proteins, e.g. phytases known in the art [for sequence information see references mentioned above, e.g. EP 684 313 or sequence data bases, for example like Genbank (Intelligenetics, California, USA), European Bioinformatics Institute (Hinxton Hall, Cambridge, GB), NBRF  
15 (Georgetown University, Medical Centre, Washington DC, USA) and Vecbase (University of Wisconsin, Biotechnology Centre, Madison, Wisconsin, USA) or disclosed in the figures by methods of in vitro mutagenesis [see e.g. Sambrook et al., Molecular Cloning, Cold Spring Harbor Laboratory Press, New York]. A widely used strategy for "site directed mutagenesis", as originally outlined by Hurchinson and Edgell [J. Virol. 8, 181 (1971)],  
20 involves the annealing of a synthetic oligonucleotide carrying the desired nucleotide substitution to a target region of a single-stranded DNA sequence wherein the mutation should be introduced [for review see Smith, Annu. Rev. Genet. 19, 423 (1985) and for improved methods see references 2-6 in Stanssen et al., Nucl. Acid Res., 17, 4441-4454 (1989)]. Another possibility of mutating a given DNA sequence which is also preferred for  
25 the practice of the present invention is the mutagenesis by using the polymerase chain reaction (PCR). DNA as starting material can be isolated by methods known in the art and described e.g. in Sambrook et al. (Molecular Cloning) from the respective strains. For strain information see, e.g. EP 684 313 or any depository authority indicated below. *Aspergillus niger* [ATCC 9142], *Myceliophthora thermophila* [ATCC 48102],  
30 *Talaromyces thermophilus* [ATCC 20186] and *Aspergillus fumigatus* [ATCC 34625] have been redeposited according to the conditions of the Budapest Treaty at the American Type Culture Cell Collection under the following accession numbers: ATCC 74337, ATCC 74340, ATCC 74338 and ATCC 74339, respectively. It is however, understood that DNA encoding a consensus protein in accordance with the present invention can also be  
35 prepared in a synthetic manner as described, e.g. in EP 747 483 or the examples by methods known in the art.

The process of the present invention can preferably be used in order to improve the thermostability of the enzyme phytase. After having constructed different consensus phytase sequences it was possible to decide whether single amino acid replacements had a positive or a negative effect on the protein stability. It is therefore another subject of the present invention to improve the thermostability of a phytase.

In this embodiment single amino acids are changed in the sequence of the phytase by the introduction of at least one mutation selected from the group consisting of

E58A	F54Y
D69K	I73V
D197N	K94A
T214L	R101A
E222T	N153K
E267D	V158I
R291I	A203G
R329H	S205G
S364T	V217A
A379K	A227V
G404A	V234L
	P238A
	Q277E
	A287H
	A292Q
	V366I
	A396S
	E415Q
	G437A
	E451R

In the above-given mutations the number represents the position in the consensus phytase-1-sequence as given in Figure 2 and the letter before the number represents the amino acid in the phytase which is replaced by the respective amino acid behind the number. The numbers given correspond to the consensus phytase sequence or relate to a  
5 corresponding residue calculated by an alignment as shown in Figure 1 when 26 amino acids (signal sequence) are added to the sequences shown in Fig. 1. Those mutations can be introduced into consensus sequences or into sequences of specific enzymes which have been improved by a process of the present invention. The above-mentioned amino acid replacements have a positive effect on the protein stability.

10 Once complete DNA sequences of the present invention have been obtained they can be integrated into vectors by methods known in the art and described e.g. in Sambrook et al. (s.a.) to overexpress the encoded polypeptide in appropriate host systems. However, a man skilled in the art knows that also the DNA sequences themselves can be used to transform the suitable host systems of the invention to get overexpression of the encoded  
15 polypeptide. Appropriate host systems are for example fungi, like Aspergilli, e.g. *Aspergillus niger* [ATCC 9142] or *Aspergillus ficuum* [NRRL 3135] or like *Trichoderma*, e.g. *Trichoderma reesei* or yeasts, like *Saccharomyces*, e.g. *Saccharomyces cerevisiae* or *Pichia*, like *Pichia pastoris*, or *Hansenula polymorpha*, e.g. *H. polymorpha* (DSM5215) or plants, as described, e.g. by Pen et al., *Bio/Technology* 11, 811-814 (1994). A man skilled  
20 in the art knows that such microorganisms are available from depository authorities, e.g. the American Type Culture Collection (ATCC), the Centraalbureau voor Schimmelcultures (CBS) or the Deutsche Sammlung für Mikroorganismen und Zellkulturen GmbH (DSM) or any other depository authority as listed in the Journal "Industrial Property" [(1991) 1, pages 29-40]. Bacteria which can be used are e.g. *E. coli*,  
25 *Bacilli* as, e.g. *Bacillus subtilis* or *Streptomyces*, e.g. *Streptomyces lividans* (see e.g. Anné and Mallaert in *FEMS Microbiol. Letters* 114, 121 (1993). *E. coli*, which could be used are *E. coli* K12 strains e.g. M15 [described as DZ 291 by Villarejo et al. in *J. Bacteriol.* 120, 466-474 (1974)], HB 101 [ATCC No. 33694] or *E. coli* SG13009 [Gottesman et al., *J. Bacteriol.* 148, 265-273 (1981)].

30 Vectors which can be used for expression in fungi are known in the art and described e.g. in EP 420 358, or by Cullen et al. [*Bio/Technology* 5, 369-376 (1987)] or Ward in *Molecular Industrial Mycology, Systems and Applications for Filamentous Fungi*, Marcel Dekker, New York (1991), Upshall et al. [*Bio/Technology* 5, 1301-1304 (1987)] Gwynne et al. [*Bio/Technology* 5, 71-79 (1987)], Punt et al. [*J. Biotechnol.* 17, 19-34 (1991)] and  
35 for yeast by Sreekrishna et al. [*J. Basic Microbiol.* 28, 265-278 (1988), *Biochemistry* 28,

4117-4125 (1989)], Hitzemann et al. [Nature 293, 717-722 (1981)] or in EP 183 070, EP 183 071, EP 248 227, EP 263 311. Suitable vectors which can be used for expression in *E. coli* are mentioned, e.g. by Sambrook et al. [s.a.] or by Fiers et al. in Proc'd. 8th Int. Biotechnology Symposium" [Soc. Franc. de Microbiol., Paris (Durand et al., eds.), pp. 680-697 (1988)] or by Bujard et al. in Methods in Enzymology, eds. Wu and Grossmann, Academic Press, Inc. Vol. 155, 416-433 (1987) and Stüber et al. in Immunological Methods, eds. Lefkovits and Pernis, Academic Press, Inc., Vol. IV, 121-152 (1990). Vectors which could be used for expression in *Bacilli* are known in the art and described, e.g. in EP 405 370, Proc'd. Natl. Acad. Sci. USA 81, 439 (1984) by Yansura and Henner, Meth. Enzymol. 185, 199-228 (1990) or EP 207 459. Vectors which can be used for the expression in *H. Polymorpha* are known in the art and described, e.g. in Gellissen et al., Biotechnology 9, 291-295 (1991).

Either such vectors already carry regulatory elements, e.g. promoters, or the DNA sequences of the present invention can be engineered to contain such elements. Suitable promoter elements which can be used are known in the art and are, e.g. for *Trichoderma reesei* the *cbh1*- [Haarki et al., Biotechnology 7, 596-600 (1989)] or the *pki1*-promotor [Schindler et al., Gene 130, 271-275 (1993)], for *Aspergillus oryzae* the *amy*-promotor [Christensen et al., Abstr. 19th Lunteren Lectures on Molecular Genetics F23 (1987), Christensen et al., Biotechnology 6, 1419-1422 (1988), Tada et al., Mol. Gen. Genet. 229, 301 (1991)], for *Aspergillus niger* the *glaA*- [Cullen et al., Bio/Technology 5, 369-376 (1987), Gwynne et al., Bio/Technology 5, 713-719 (1987), Ward in Molecular Industrial Mycology, Systems and Applications for Filamentous Fungi, Marcel Dekker, New York, 83-106 (1991)], *alcA*- [Gwynne et al., Bio/Technology 5, 718-719 (1987)], *suc1*- [Boddy et al., Curr. Genet. 24, 60-66 (1993)], *aphA*- [MacRae et al., Gene 71, 339-348 (1988), MacRae et al., Gene 132, 193-198 (1993)], *tpiA*- [McKnight et al., Cell 46, 143-147 (1986), Upshall et al., Bio/Technology 5, 1301-1304 (1987)], *gpdA*- [Punt et al., Gene 69, 49-57 (1988), Punt et al., J. Biotechnol. 17, 19-37 (1991)] and the *pkiA*-promotor [de Graaff et al., Curr. Genet. 22, 21-27 (1992)]. Suitable promoter elements which could be used for expression in yeast are known in the art and are, e.g. the *pho5*-promotor [Vogel et al., Mol. Cell. Biol., 2050-2057 (1989); Rudolf and Hinnen, Proc. Natl. Acad. Sci. 84, 1340-1344 (1987)] or the *gap*-promotor for expression in *Saccharomyces cerevisiae* and for *Pichia pastoris*, e.g. the *aox1*-promotor [Koutz et al., Yeast 5, 167-177 (1989); Sreekrishna et al., J. Basic Microbiol. 28, 265-278 (1988)], or the FMD promoter [Hollenberg et al., EPA No. 0299108] or *MOX*-promotor [Ledeboer et al., Nucleic Acids Res. 13, 3063-3082 (1985)] for *H. polymorpha*.

Accordingly vectors comprising DNA sequences of the present invention, preferably for the expression of said DNA sequences in bacteria or a fungal or a yeast host and such transformed bacteria or fungal or yeast hosts are also an object of the present invention.

It is also an object of the present invention to provide a system which allows for high  
5 expression of proteins, preferably phytases like the consensus phytase of the present invention in *Hansenula* characterized therein that the codons of the encoding DNA sequence of such a protein have been selected on the basis of a codon frequency table of the organism used for expression, e.g. yeast as in the present case (see e.g. in Example 3) and optionally the codons for the signal sequence have been selected in a manner as  
10 described for the specific case in Example 3. That means that a codon frequency table is prepared on the basis of the codons used in the DNA sequences which encode the amino acid sequences of the defined protein family. Then the codons for the design of the DNA sequence of the signal sequence are selected from a codon frequency table of the host cell used for expression whereby always codons of comparable frequency in both tables are  
15 used.

Once such DNA sequences have been expressed in an appropriate host cell in a suitable medium the encoded protein can be isolated either from the medium in the case the protein is secreted into the medium or from the host organism in case such protein is present intracellularly by methods known in the art of protein purification or described in  
20 case of a phytase, e.g. in EP 420 358. Accordingly a process for the preparation of a polypeptide of the present invention characterized in that transformed bacteria or a host cell as described above is cultured under suitable culture conditions and the polypeptide is recovered therefrom and a polypeptide when produced by such a process or a polypeptide encoded by a DNA sequence of the present invention are also an object of the present  
25 invention.

Once obtained the polypeptides of the present invention can be characterized regarding their properties which make them useful in agriculture any assay known in the art and described e.g. by Simons et al. [Br. J. Nutr. 64, 525-540 (1990)], Schöner et al. [J. Anim. Physiol. a. Anim. Nutr. 66, 248-255 (1991)], Vogt [Arch. Geflügelk. 56, 93-98  
30 (1992)], Jongbloed et al. [J. Anim. Sci., 70, 1159-1168 (1992)], Perney et al. [Poultry Sci. 72, 2106-2114 (1993)], Farrell et al., [J. Anim. Physiol. a. Anim. Nutr. 69, 278-283 (1993)], Broz et al., [Br. Poultry Sci. 35, 273-280 (1994)] and Dünghoef et al. [Animal Feed Sci. Technol. 49, 1-10 (1994)] can be used.

In general the polypeptides of the present invention can be used without being  
35 limited to a specific field of application, e.g. in case of phytases for the conversion of inositol polyphosphates, like phytate to inositol and inorganic phosphate.



Furthermore the polypeptides of the present invention can be used in a process for the preparation of a pharmaceutical composition or compound food or feeds wherein the components of such a composition are mixed with one or more polypeptides of the present invention. Accordingly compound food or feeds or pharmaceutical compositions comprising one or more polypeptides of the present invention are also an object of the present invention. A man skilled in the art is familiar with their process of preparation. Such pharmaceutical compositions or compound foods or feeds can further comprise additives or components generally used for such purpose and known in the state of the art.

It is furthermore an object of the present invention to provide a process for the reduction of levels of phytate in animal manure characterized in that an animal is fed such a feed composition in an amount effective in converting phytate contained in the feedstuff to inositol and inorganic phosphate.

Before describing the present invention in more detail a short explanation of the Figures enclosed is given below.

**Figure 1:** Design of the consensus phytase sequence. The letters represent the amino acid residues in the one-letter code. The following sequences were used for the alignment: *phyA* from *Aspergillus terreus* 9A-1 (Mitchell et al, 1997; from amino acid (aa) 27), *phyA* from *A. terreus* cbs116.46; (van Loon et al., 1998; from aa 27), *phyA* from *Aspergillus niger* var. *awamori* (Piddington et al, 1993; from aa 27), *phyA* from *A. niger* T213; from aa 27), *phyA* from *A. niger* strain NRRL3135 (van Hartingsveldt et al, 1993; from aa 27), *phyA* from *Aspergillus fumigatus* ATCC 13073 (Pasamontes et al, 1993; from aa 25), *phyA* from *A. fumigatus* ATCC 32722 (van Loon et al, 1998; from aa 27), *phyA* from *A. fumigatus* ATCC 58128 (van Loon et al., 1998; from aa 27), *phyA* from *A. fumigatus* ATCC 26906 (van Loon et al, 1998; from aa 27), *phyA* from *A. fumigatus* ATCC 32239 (van Loon et al, 1998; from aa 30), *phyA* from *Emmericella nidulans* (Pasamontes et al, 1997a; from aa 25), *phyA* from *Talaromyces thermophilus* (Pasamontes et al, 1997a; from aa 24), and *phyA* from *Myceliophthora thermophila* (Mitchell et al, 1997; from aa 19). The alignment was calculated using the program PILEUP. The location of the gaps was refined by hand. Capitalized amino acid residues in the alignment at a given position belong to the amino acid coalition that establish the consensus residue. In bold, beneath the calculated consensus sequence, the amino acid sequence of the finally constructed consensus phytase (Fcp) is shown. The gaps in the calculated consensus sequence were filled by hand according to principals stated in Example 1.

**Figure 2:** DNA sequence of the consensus phytase-1 gene (*fcp*) and of the primers used for the gene construction. The calculated amino acid sequence (Figure 1) was converted into a DNA sequence using the program BACKTRANSLATE (Devereux *et al.*, 1984) and the codon frequency table of highly expressed yeast genes (GCG program package, 9.0). The signal peptide of the phytase from *A. terreus* cbs.116.46 was fused to the *N*-terminus. The bold bases represent the sequences of the oligonucleotides used to generate the gene. The names of the respective oligonucleotides are alternately noted above or below the sequence. The underlined bases represent the start and stop codon of the gene. The bases written in italics show the two introduced *Eco* RI sites.

**Figure 3:** Alignment and consensus sequence of five *Basidiomycetes* phytases. The letters represent the amino acid residues in the one-letter code. The amino acid sequences of the phytases from *Paxillus involutus*, phyA1 (aa 21) and phyA2 (aa 21, WO 98/28409), *Trametes pubescens* (aa 24, WO 98/28409), *Agrocybe pediades* (aa 19, WO 98/28409), and *Peniophora lycii* (aa 21, WO 98/28409) starting with the amino acid residues mentioned in parentheses, were used for the alignment and the calculation of the corresponding consensus sequence called "Basidio" (Example 2). The alignment was performed by the program PILEUP. The location of the gaps was refined by hand. The consensus sequence was calculated by the program PRETTY. While a vote weight of 0.5 was assigned to the two *P. involutus* phytases, all other genes were used with a vote weight of 1.0 for the consensus sequence calculation. At positions, where the program was not able to determine a consensus residues, the Basidio sequence contains a dash. Capitalized amino acid residues in the alignment at a given position belong to the amino acid coalition that establish the consensus residue.

**Figure 4:** Design of consensus phytase-10 amino acid sequence. Adding the phytase sequence of *Thermomyces lanuginosa* (Berka *et al.*, 1998) and the consensus sequence of the phytases from five *Basidiomycetes* to the alignment of Figure 1, an improved consensus sequence was calculated by the program PRETTY. Additionally, the amino acid sequence of *A. niger* T213 was omitted, therefore, using a vote weight of 0.5 for the remaining *A. niger* phytase sequences. For further information see Example 2.

**Figure 5:** DNA and amino acid sequence of consensus phytase-10. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The sequence of the oligonucleotides which were used to assemble the gene are in bold letters. The label of oligonucleotides and the amino acids, which were changed compared to those for consensus phytase -1, are underlined and their corresponding triplets are highlighted in

small cases. The *fcp10* gene was assembled from the following oligonucleotides: CP-1, CP-2, CP-3.10, CP-4.10, CP-5.10, CP-6, CP-7.10, CP-8.10, CP-9.10, CP-10.10, CP-11.10, CP-12.10, CP-13.10, CP-14.10, CP-15.10, CP-16.10, CP-17.10, CP-18.10, CP-19.10, CP-20.10, CP-21.10, CP-22.10. The newly synthesized oligonucleotides are additionally  
5 marked by number 10. The phytase contains the following 32 exchanges: Y54F, **E58A**, D69K, D70G, A94K, N134Q, I158V, S187A, Q188N, **D197N**, S204A, T214L, D220E, L234V, A238P, D246H, T251N, Y259N, **E267D**, E277Q, A283D, **R291I**, A320V, **R329H**, **S364T**, I366V, **A379K**, S396A, **G404A**, Q415E, A437G, A463E. The mutations accentuated in bold letters revealed a stabilizing effect on consensus phytase-1 as tested as  
10 single mutation in consensus phytase-1.

**Figure 6:** Alignment for the design of consensus phytase-11. In contrast to the design of consensus phytase-10, for the design of the amino acid sequence of consensus phytase-11, all *Basidiomycetes* phytases were used as independent sequences using an assigned vote weight of 0.2 for each *Basidiomycetes* sequence. Additionally, the amino acid sequence of  
15 *A. niger* T213 was used in that alignment, again.

**Figure 7:** DNA and amino acid sequence of consensus phytase-1-thermo[8]-Q50T-K91A. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (\*).

20 **Figure 8:** DNA and amino acid sequence of consensus phytase-10-thermo[3]-Q50T-K91A. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (\*).

**Figure 9:** DNA and amino acid sequence of *A. fumigatus* ATCC 13073 phytase a-mutant.  
25 The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (\*).

**Figure 10:** DNA and amino acid sequence of consensus phytase-7. The amino acids are written above the corresponding DNA sequence using the one-letter code. The sequence of  
30 the oligonucleotides used to assemble the gene are in bold letters. Oligonucleotides and amino acids that were exchanged are underlined and their corresponding triplets are highlighted in small cases. The *fcp7* gene was assembled from the following

oligonucleotides: CP-1, CP-2, CP-3, CP-4.7, CP-5.7, CP-6, CP-7, CP-8.7, CP-9, CP-10.7, CP-11.7, CP-12.7, CP-13.7, CP-14.7, CP-15.7, CP-16, CP-17.7, CP-18.7, CP-19.7, CP-20, CP-21, CP-22. The newly synthesized oligonucleotides are additionally marked by number 7. The phytase contains the following 24 exchanges in comparison to the original  
 5 consensus phytase: S89D, S92G, A94K, D164S, P201S, G203A, G205S, H212P, G224A, D226T, E255T, D256E, V258T, P265S, Q292H, G300K, Y305H, A314T, S364G, M365I, A397S, S398A, G404A, and A405S.

**Figure 11:** Differential scanning calorimetry (DSC) of consensus phytase-1 and consensus phytase-10. The protein samples were concentrated to ca. 50-60 mg/ml and extensively  
 10 dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-10 (upper graph) yielded a melting temperature of 85.4 °C, which is 7.3 °C higher than the melting point of consensus phytase-1 (78.1 °C, lower graph).

**Figure 12:** Differential scanning calorimetry (DSC) of consensus phytase-10-thermo-Q50T  
 15 and consensus phytase-10-thermo-Q50T-K91A. The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-10-thermo-Q50T (upper graph) yielded a melting temperature of 88.6 °C, while the melting point of consensus phytase-10-thermo-Q50T-K91A was found at 89.3 °C.

**Figure 13:** Comparison of the temperature optimum between consensus phytase-1,  
 20 consensus phytase-10 and consensus phytase-10-thermo-Q50T. For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 86 °C. The diluted supernatant of transformed *S. cerevisiae* strains was used for the determination. The other components of the supernatant showed  
 25 no influence on the determination of the temperature optimum: △, consensus phytase-1; ◇, consensus phytase-10; ■, consensus phytase 10-thermo-Q50T.

**Figure 14:** pH-dependent activity profile and substrate specificity of consensus phytase-10  
 and its variants thermo-Q50T and thermo-Q50T-K91A. The phytase activity was  
 determined using the standard assay in appropriate buffers (see Example 9) at different pH-  
 30 values. Graph a) shows the pH-dependent activity profile of consensus phytase-10 (□), consensus phytase-10-thermo-Q50T (•), and consensus phytase-10-thermo-Q50T-K91A (△). Graph b) shows the corresponding substrate specificity tested by replacement of phytate by the indicated compounds in the standard assay; open bars, consensus phytase-10

(grey bars, consensus phytase-10-thermo-Q50T; dark bars, consensus phytase-10-thermo-Q50T-K91A). The numbers correspond to the following compounds: 1, phytate; 2, *p*-nitrophenyl phosphate; 3, phenyl phosphate; 4, fructose-1,6-bisphosphate; 5, fructose-6-phosphate; 6, glucose-6-phosphate; 7, ribose-5-phosphate; 8, DL-glycerol-3-phosphate; 9, glycerol-2-phosphate; 10, 3-phosphoglycerate; 11, phosphoenolpyruvate; 12, AMP; 13, ADP; 14, ATP.

**Figure 15:** pH-dependent activity profile and substrate specificity of consensus phytase-1-thermo[8]-Q50T and of consensus phytase-1-thermo[8]-Q50T-K91A. The phytase activity was determined using the standard assay in appropriate buffers (see Example 9) at different pH-values. Graph a) shows the pH-dependent activity profile of the Q50T- (■) and the Q50T-K91A-variant (•). Graph b) shows the corresponding substrate specificities tested by replacement of phytate by the indicated compounds in the standard assay (open bars, consensus phytase-1-thermo[8]-Q50T; filled bars, consensus phytase-1-thermo[8]-Q50T-K91A.). The substrates are listed in the legend of Figure 14.

**Figure 16:** Differential scanning calorimetry (DSC) of consensus phytase-1-thermo[8]-Q50T and consensus phytase-1-thermo[8]-Q50T-K91A. The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-1-thermo[8]-Q50T (upper graph) showed a melting temperature of 84.7 °C, while the melting point of consensus phytase-1-thermo[8]-Q50T-K91A was found at 85.7 °C.

**Figure 17:** Comparison of the temperature optimum between consensus phytase-1, consensus phytase-1-thermo[3] and consensus phytase-1-thermo[8]. For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 86 °C. Purified protein from the supernatant of transformed *S. cerevisiae* strains was used for the determination. O, consensus phytase-1; □, consensus phytase-1-thermo[3]; ▲, consensus phytase 1-thermo[8].

**Figure 18:** Comparison of the pH-dependent activity profile and substrate specificity of consensus phytase-1, consensus phytase-7, and of the phytase from *A. niger* NRRL 3135. The phytase activity was determined using the standard assay in appropriate buffers (see Example 9) at different pH-values. Graph a) shows the pH-dependent activity profile of consensus phytase-1 (■), the phytase from *A. niger* NRRL 3135 (○), and of consensus phytase-7 (▲). Graph b) shows the corresponding substrate specificity tested by replacement of phytate by the indicated compounds in the standard assay (black bars, *A.*

*niger* NRRL 3135 phytase; grey bars, consensus phytase-1, dashed bars, consensus phytase-7). The substrates are listed in the legend of Figure 14.

**Figure 19:** Differential scanning calorimetry (DSC) of the phytase from *A. fumigatus* ATCC 13073 and of its stabilized  $\alpha$ -mutant, which contains the following amino acid exchanges F55Y, V100I, F114Y, A243L, S265P, N294D.

The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus *A. fumigatus* 13073 phytase (upper graph) revealed a melting temperature of 62.5 °C, while the melting point of the  $\alpha$ -mutant was found at 67.0 °C

**Figure 20:** Comparison of the temperature optimum of *A. fumigatus* 13073 wild-type, its *A. fumigatus*  $\alpha$ -mutant, and a further stabilized  $\alpha$ -mutant (E59A-S126N-R329H-S364T-G404A). For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 75 °C. The diluted supernatant of transformed *S. cerevisiae* strains was used for the determination. The other components of the supernatant showed no influence on the determination of the temperature optimum.

○, *A. fumigatus* ATCC 13073 phytase; ▲, *A. fumigatus* ATCC 13073  $\alpha$ -mutant; □, *A. fumigatus* ATCC 13073 alpha-mutant-(E59A-S126N-R329H-S364T-G404A)-Q27T; ■, *A. fumigatus* ATCC 13073  $\alpha$ -mutant-(E59A-S126N-R329H-S364T-G404A)-Q27T-K68A. Q27T and K68A corresponds to consensus phytase-1 Q50T and K91A, respectively.

**Figure 21:** Amino acid sequence of consensus phytase 12 (consphy12) which contains a number of active site residues transferred from the "basidio" consensus sequence to consensus phytase-10-thermo-Q50T-K91A.

**Example 1:**

**Design of the amino acid sequence of consensus phytase-1**

**Alignment of the amino acid sequences**

The alignment was calculated using the program PILEUP from the Sequence  
5 Analysis Package Release 9.0 (Devereux *et al.*, 1984) with the standard parameter (gap  
creation penalty 12, gap extension penalty 4). The location of the gaps was refined using a  
text editor. Table 1 shows the sequences (see Figure 1) without the signal sequence that  
were used for the performance of the alignment starting with the amino acid (aa) as  
mentioned in Table 1.

10 **Table 1: Origin and vote weight of the phytase amino acid sequences used for the design of  
consensus phytase-1**

- *phyA* from *Aspergillus terreus* 9A-1, aa 27, vote weight 0.5 (Mitchell *et al.*, 1997)
- *phyA* from *Aspergillus terreus* cbs116.46, aa 27, vote weight 0.5 (van Loon *et al.*, 1998)
- *phyA* from *Aspergillus niger* var. *awamori*, aa 27, vote weight 0.33 (Piddington *et al.*,  
15 1993)
- *phyA* from *Aspergillus niger* T213, aa 27, vote weight 0.33
- *phyA* from *Aspergillus niger* strain NRRL3135, aa 27, vote weight 0.33 (van  
Hartingsveldt *et al.*, 1993)
- *phyA* from *Aspergillus fumigatus* ATCC 13073, aa 26, vote weight 0.2 (Pasamontes *et*  
20 *al.*, 1997)
- *phyA* from *Aspergillus fumigatus* ATCC 32722, aa 26, vote weight 0.2 (van Loon *et al.*,  
1998)
- *phyA* from *Aspergillus fumigatus* ATCC 58128, aa 26, vote weight 0.2 (van Loon *et al.*,  
1998)
- 25 - *phyA* from *Aspergillus fumigatus* ATCC 26906, aa 26, vote weight 0.2 (van Loon *et al.*,  
1998)
- *phyA* from *Aspergillus fumigatus* ATCC 32239, aa 30, vote weight 0.2 (van Loon *et al.*,  
1998)
- *phyA* from *Emericella nidulans* , aa 25, vote weight 1.0 (Roche Nr. R1288, Pasamontes  
30 *et al.*, 1997a)
- *phyA* from *Talaromyces thermophilus* ATCC 20186, aa 24, vote weight 1.0 (Pasamontes  
*et al.*, 1997a)
- *phyA* from *Myceliophthora thermophila*, aa 19, vote weight 1.0 (Mitchell *et al.*, 1997)

### Calculation of the amino acid sequence of consensus phytase-1

Using the refined alignment as input, the consensus sequence was calculated by the program PRETTY from the Sequence Analysis Package Release 9.0 (Devereux *et al.*,  
5 1984). PRETTY prints sequences with their columns aligned and can display a consensus sequence for an alignment. A vote weight that pays regard to the similarity between the amino acid sequences of the phytases aligned was assigned to all sequences. The vote weight was set such as the combined impact of all phytases from one sequence subgroup (same species, but from different strains), e. g. the amino acid sequences of all phytases  
10 from *A. fumigatus*, on the election was set one, that means that each sequence contributes with a value of 1 divided by the number of strain sequences (see Table 1). By this means, it was possible to prevent that very similar amino acid sequences, e. g. of the phytases from different *A. fumigatus* strains, dominate the calculated consensus sequence.

The program PRETTY was started with the following parameters: The plurality  
15 defining the number of votes below which there is no consensus was set on 2.0. The threshold, which determines the scoring matrix value below which an amino acid residue may not vote for a coalition of residues, was set on 2. PRETTY used the PrettyPep.Cmp consensus scoring matrix for peptides.

Ten positions of the alignment (position 46, 66, 82, 138, 162, 236, 276, 279, 280,  
20 308; Figure 1), for which the program was not able to determine a consensus residue, were filled by hand according to the following rules: if a most frequent residue existed, this residue was chosen (138, 236, 280); if a prevalent group of similar or phylogenetically equivalent residues occurred, the most frequent or, if not available, one residues of this group was selected (46, 66, 82, 162, 276, 308). If there was either a prevalent residue nor a  
25 prevalent group, one of the occurring residues was chosen according to common assumption on their influence on the protein stability (279). Eight other positions (132, 170, 204, 211, 275, 317, 384, 447; Figure 1) were not filled with the amino acid residue selected by the program but normally with amino acids that occur with the same frequency as the residues that were chosen by the program. In most cases, the slight underrating of  
30 the three *A. niger* sequences (sum of the vote weights: 0.99) was eliminated by this corrections.

### Conversion of the consensus phytase-1 amino acid sequence to a DNA sequence

The first 26 amino acid residues of *A. terreus* cbs116.46 phytase were used as signal  
35 peptide and, therefore, fused to the N-terminus of all consensus phytases. For this stretch,



we used a special method to calculate the corresponding DNA sequence. Purvis et al (1987) proposed that the incorporation of rare codons in a gene has an influence on the folding efficiency of the protein. Therefore, at least the distribution of rare codons in the signal sequence of *A. terreus* cbs1 16.46, which was used for the consensus phytase and  
5 which is very important for secretion of the protein, but converted into the *S. cerevisiae* codon usage, was transferred into the new signal sequence generated for expression in *S. cerevisiae*. For the remaining parts of the protein, we used the codon frequency table of highly expressed *S. cerevisiae* genes, obtained from the GCG program package, to translate the calculated amino acid sequence into a DNA sequence.

10 The resulting sequence of the *fcp* gene is shown in Figure 2.

### Construction and cloning of the consensus phytase-1 gene

The calculated DNA sequence of consensus phytase-1 (*fcp*) was divided into oligonucleotides of 85 bp, alternately using the sequence of the sense and the anti-sense strand. Every oligonucleotide overlaps 20 bp with its previous and its following  
15 oligonucleotide of the opposite strand. The location of all primers, purchased by Microsynth, Balgach (Switzerland) and obtained in a PAGE-purified form, is indicated in Figure 2.

### PCR-Reactions

In three PCR reactions, the synthesized oligonucleotides were composed to the entire  
20 gene. For the PCR, the High Fidelity Kit from Boehringer Mannheim (Boehringer Mannheim, Mannheim, Germany) and the thermo cycler The Protokol™ from AMS Biotechnology (Europe) Ltd. (Lugano, Switzerland) was used.

Oligonucleotide CP-1 to CP-10 (Mix 1, Figure 2) were mixed to a concentration of 0.2 pMol/μl of each oligonucleotide. A second oligonucleotide mixture (Mix 2) was  
25 prepared with CP-9 to CP-22 (0.2 pMol/μl of each oligonucleotide). Additionally, four short primers were used in the PCR reactions:

CP-a: *Eco* RI

5'-TATATGAATTCATGGGCGTGTTCGTC-3'

CP-b:  
30 5'-TGAAAAGTTCATTGAAGGTTTC-3'

CP-c:  
5'-TCTTCGAAAGCAGTACAAGTAC-3'

CP-e:

*Eco* RI

5'-TATATGAATTCTTAAGCGAAAC-3'

5      PCR reaction *a*:                      10 µl Mix 1 (2.0 pmol of each oligonucleotide)  
   2 µl nucleotides (10 mM each nucleotide)  
   2 µl primer CP-a (10 pmol/µl)  
   2 µl primer CP-c (10 pmol/µl)  
   10,0 µl PCR buffer  
10    0.75 µl polymerase mixture  
   73.25 µl H<sub>2</sub>O

                 PCR reaction *b*:                      10 µl Mix 2 (2.0 pmol of each oligonucleotide)  
   2 µl nucleotides (10 mM each nucleotide)  
   2 µl primer CP-b (10 pmol/µl)  
   2 µl primer CP-e (10 pmol/µl)  
15    10,0 µl PCR buffer  
   0.75 µl polymerase mixture (2.6 U)  
   73.25 µl H<sub>2</sub>O

Reaction conditions for PCR reaction *a* and *b*:

20    step 1    2 min - 45°C  
   step 2    30 sec - 72°C  
   step 3    30 sec - 94°C  
   step 4    30 sec - 52°C  
   step 5    1 min - 72°C

Step 3 to 5 were repeated 40-times.

25                      The PCR products (670 and 905 bp) were purified by an agarose gel electrophoresis  
(0.9% agarose) and a following gel extraction (QIAEX II Gel Extraction Kit, Qiagen,  
Hilden, Germany). The purified DNA fragments were used for the PCR reaction *c*.

30                      PCR reaction *c*:                      6 µl PCR product of reaction *a* (≈50 ng)  
   6 µl PCR product of reaction *b* (≈50 ng)  
   2 µl primer CP-a (10 pmol/µl)  
   2 µl primer CP-e (10 pmol/µl)  
   10,0 µl PCR buffer  
   0.75 µl polymerase mixture (2.6 U)  
   73.25 µl H<sub>2</sub>O

35      Reaction conditions for PCR reaction *c*:

step 1 2 min - 94°C  
step 2 30 sec - 94°C  
step 3 30 sec - 55°C  
step 4 1 min - 72°C

5 Step 2 to 4 were repeated 31-times.

The resulting PCR product (1.4 kb) was purified as mentioned above, digested with *Eco* RI, and ligated in an *Eco* RI-digested and dephosphorylated pBsk(-)-vector (Stratagene, La Jolla, CA, USA). 1 µl of the ligation mixture was used to transform *E. coli* XL-1 competent cells (Stratagene, La Jolla, CA, USA). All standard procedures were  
10 carried out as described by Sambrook *et al.* (1987). The DNA sequence of the constructed consensus phytase gene (*fcp*, Figure 2) was controlled by sequencing as known in the art.

### Example 2

#### Design of an improved consensus phytase (consensus phytase-10) amino acid sequence

15 The alignments used for the design of consensus phytase-10 were calculated using the program PILEUP from the Sequence Analysis Package Release 9.0 (Devereux *et al.*, 1984) with the standard parameter (gap creation penalty 12, gap extension penalty 4). The location of the gaps was refined using a text editor.

The following sequences were used for the alignment of the *Basidiomycetes* phytases  
20 starting with the amino acid (aa) mentioned in Table 2:

Table 2: Origin and vote weight of five *Basidiomycetes* phytases used for the calculation of the corresponding amino acid consensus sequence (basidio)

- *phyA1* from *Paxillus involutus* NN005693, aa 21, vote weight 0.5 (WO 98/28409)
- *phyA2* from *Paxillus involutus* NN005693, aa 21, vote weight 0.5 (WO 98/28409)
- 25 - *phyA* from *Trametes pubescens* NN9343, aa 24, vote weight 1.0 (WO 98/28409)
- *phyA* from *Agrocybe pediades* NN009289, aa 19, vote weight 1.0 (WO 98/28409)
- *phyA* from *Peniophora lycii* NN006113, aa 21, vote weight 1.0 (WO 98/28409)

The alignment is shown in Figure 3.

In Table 3 the genes, which were used for the performance of the final alignment, are  
30 arranged. The first amino acid (aa) of the sequence which is used in the alignment is mentioned behind the organism designation.

**Table 3: Origin and vote weight of the phytase sequences used for the design of consensus phytase 10**

- *phyA* from *Aspergillus terreus* 9A-1, aa 27, vote weight 0.5 (Mitchell *et al.*, 1997)
- *phyA* from *Aspergillus terreus* cbs116.46, aa 27, vote weight 0.5 (van Loon *et al.*, 1998)
- 5 - *phyA* from *Aspergillus niger* var. *awamori*, aa 27, vote weight 0.5 (Piddington *et al.*, 1993)
- *phyA* from *Aspergillus niger* strain NRRL3135, aa 27, vote weight 0.5 (van Hartingsveldt *et al.*, 1993)
- *phyA* from *Aspergillus fumigatus* ATCC 13073, aa 26, vote weight 0.2 (Pasamontes *et al.*, 1997)
- 10 - *phyA* from *Aspergillus fumigatus* ATCC 32722, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)
- *phyA* from *Aspergillus fumigatus* ATCC 58128, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)
- 15 - *phyA* from *Aspergillus fumigatus* ATCC 26906, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)
- *phyA* from *Aspergillus fumigatus* ATCC 32239, aa 30, vote weight 0.2 (van Loon *et al.*, 1998)
- *phyA* from *Emmericella nidulans*, aa 25, vote weight 1.0 (Roche Nr. R1288, Pasamontes *et al.*, 1997a)
- 20 - *phyA* from *Talaromyces thermophilus* ATCC 20186, aa 24, vote weight 1.0 (Pasamontes *et al.*, 1997a)
- *phyA* from *Myceliophthora thermophila*, aa 19, vote weight 1.0 (Mitchell *et al.*, 1997)
- *phyA* from *Thermomyces lanuginosa*, aa 36, vote weight 1.0 (Berka *et al.*, 1998)
- 25 - Consensus sequence of five *Basidiomycetes* phytases, vote weight 1.0 (Basidio, Figure 3)

The corresponding alignment is shown in Figure 4.

#### **Calculation of the amino acid sequence of consensus-10**

To improve the alignment, we added the original consensus sequence of five phytases from four different *Basidiomycetes*, called Basidio, still containing the undefined sequence positions (see Figure 3), nearly all phytase sequences used for calculation of the original consensus phytase and one new phytase sequence from the *Ascomycete* *Thermomyces lanuginosa* to a larger alignment. Using the consensus sequence of the basidiomycetal phytase sequences, does not pay regard to the diversity among the five amino acid sequences, but pays regard to the common and different amino acid residues between the phytases from the *Ascomycetes* and the *Basidiomycetes*.

We set plurality on 2.0 and threshold on 3. The used vote weight are listed in Table 3. The alignment and the corresponding consensus sequence is presented in Figure 4. The new consensus phytase sequence has 32 different amino acids in comparison to the original consensus phytase. Positions for which the program PRETTY was not able to calculate a  
5 consensus amino acid residue were filled according to rules mentioned in Example 1. None of the residues suggested by the program was replaced.

Furthermore, we included all *Basidiomycetes* phytases as single amino acid sequences but assigning a vote weight of 0.2 in the alignment. The corresponding alignment is shown in Figure 6. The calculated consensus amino acid sequence (consensus phytase-11) has the  
10 following differences to the sequence of consensus phytase-10. Letter X means that the program was not able to calculate a consensus amino acid; the amino acid in parenthesis corresponds to the amino acid finally included into the consensus phytase-10.

D35X, X(K)69K, X(E)100E, A101R, Q134N, X(K)153N, X(H)190H, X(A)204S,  
X(E)220D, E222T, V227A, X(R)271R, H287A, X(D)288D, X(K)379K, X(T)389I, E390X,  
15 X(E)415E, X(A)416A, X(R)446L, E463A, whereas the numbering is as in Fig. 5.

We also checked single amino acid replacements suggested by the improved consensus sequences 10 and 11 on their influence on the stability of the original consensus phytase. The approach is described in example 3.

#### 20                    **Conversion of consensus phytase-10 amino acid sequence to a DNA sequence**

The first 26 amino acid residues of *A. terreus* cbs116.46 phytase were used as signal peptide and, therefore, fused to the *N*-terminus of consensus phytase-10. The used procedure is further described in Example 1.

The resulting sequence of the *fcp10* gene is shown in Figure 5.

25

#### **Construction and cloning of the consensus phytase-10 gene (*fcp10*)**

The calculated DNA sequence of *fcp10* was divided into oligonucleotides of 85 bp, alternately using the sequence of the sense and the anti-sense strand. Every oligonucleotide overlaps 20 bp with its previous and its following oligonucleotide of the opposite strand.

30 The location of all primers, purchased by Microsynth, Balgach (Switzerland) and obtained in a PAGE-purified form, is indicated in Figure 5.

### PCR-Reactions

In three PCR reactions, the synthesized oligonucleotides were composed to the entire gene. For the PCR, the High Fidelity Kit from Boehringer Mannheim (Boehringer Mannheim, Mannheim, Germany) and the thermo cycler The Protokol<sup>TM</sup> from AMS Biotechnology (Europe) Ltd. (Lugano, Switzerland) was used. The following oligonucleotides were used in a concentration of 0.2 pMol/ml.

Mix 1.10: CP-1, CP-2, CP-3.10, CP-4.10, CP-5.10, CP-6, CP-7.10, CP-8.10, CP-9.10, CP-10.10

Mix 2.10: CP-9.10, CP-11.10, CP-12.10, CP-13.10, CP-14.10, CP-15.10, CP-16.10, CP-17.10, CP18.10, CP-19.10, CP-20.10, CP-21.10, CP-22.10

The newly synthesized oligonucleotides are marked by number 10. The phytase contains the following 32 exchanges, which are underlined in Figure 5, in comparison to the original consensus phytase: Y54F, E58A, D69K, D70G, A94K, N134Q, I158V, S187A, Q188N, D197N, S204A, T214L, D220E, L234V, A238P, D246H, T251N, Y259N, E267D, E277Q, A283D, R291I, A320V, R329H, S364T, I366V, A379K, S396A, G404A, Q415E, A437G, A463E.

Four short PCR primer were used for the assembling of the oligonucleotides:

CP-a: *Eco* RI  
5'-TATATGAATTCATGGGCGTGTTCGTC-3'

CP-b: 5'-TGAAAAGTTCATTGAAGGTTTC-3'

CP-c.10: 5'-TCTTCGAAAGCAGTACACAAAC-3'

CP-e: *Eco* RI  
5'-TATATGAATTCTTAAGCGAAAC-3'

PCR reaction  $\alpha$ :  
10  $\mu$ l Mix 1.10 (2.0 pmol of each oligonucleotide)  
2  $\mu$ l nucleotides (10 mM each nucleotide)  
2  $\mu$ l primer CP-a (10 pmol/ml)  
2  $\mu$ l primer CP-c.10 (10 pmol/ml)  
10,0  $\mu$ l PCR buffer  
0.75  $\mu$ l polymerase mixture  
73.25  $\mu$ l H<sub>2</sub>O

PCR reaction *b*:  
10 µl Mix 2.10 (2.0 pmol of each oligonucleotide)  
2 µl nucleotides (10 mM each nucleotide)  
2 µl primer CP-b (10 pmol/ml)  
2 µl primer CP-e (10 pmol/ml)  
5 10,0 µl PCR buffer  
0.75 µl polymerase mixture (2.6 U)  
73.25 µl H<sub>2</sub>O

Reaction conditions for PCR reaction *a* and *b*:

10 step 1 2 min - 45 °C  
step 2 30 sec - 72 °C  
step 3 30 sec - 94 °C  
step 4 30 sec - 52 °C  
step 5 1 min - 72 °C

Step 3 to 5 were repeated 40-times.

15 The PCR products (670 and 905 bp) were purified by an agarose gel electrophoresis (0.9% agarose) and a following gel extraction (QIAEX II Gel Extraction Kit, Qiagen, Hilden, Germany). The purified DNA fragments were used for the PCR reaction *c*.

PCR reaction *c*:  
6 µl PCR product of reaction *a* ≈50 ng)  
6 µl PCR product of reaction *b* ≈50 ng)  
20 2 µl primer CP-a (10 pmol/ml)  
2 µl primer CP-e (10 pmol/ml)  
10,0 µl PCR buffer  
0.75 µl polymerase mixture (2.6 U)  
73.25 µl H<sub>2</sub>O

25 Reaction conditions for PCR reaction *c*:

step 1 2 min - 94 °C  
step 2 30 sec - 94 °C  
step 3 30 sec - 55 °C  
step 4 1 min - 72 °C

30 Step 2 to 4 were repeated 31-times.

The resulting PCR product (1.4 kb) was purified as mentioned above, digested with *Eco* RI, and ligated in an *Eco* RI-digested and dephosphorylated pBsk(-)-vector (Stratagene, La Jolla, CA, USA). 1 µl of the ligation mixture was used to transform *E. coli* XL-1 competent cells (Stratagene, La Jolla, CA, USA). All standard procedures were  
35 carried out as described by Sambrook *et al.* (1987). The DNA sequence of the constructed gene (*fcp10*) was checked by sequencing as known in the art.

Example 3

Increasing the thermostability of consensus phytase-1 by introduction of single mutations suggested by the amino acid sequence of consensus phytase-10 and consensus phytase-11

5 In order to increase the thermostability of homologous genes, it is also possible to test the stability effect of each differing amino acid residue between the protein of interest and the calculated consensus sequence and to combine all stabilizing mutations into the protein of interest. We used the consensus phytase as protein of interest and tested the effect on the protein stability of 34 amino acid residues, differing to consensus phytase 10  
10 and/or 11 as single mutations.

To construct muteins for expression in *A. niger*, *S. cerevisiae*, or *H. polymorpha*, the corresponding expression plasmid containing the consensus phytase gene was used as template for site-directed mutagenesis (see Example 6-8). Mutations were introduced using the "quick exchange<sup>TM</sup> site-directed mutagenesis kit" from Stratagene (La Jolla, CA,  
15 USA) following the manufacturer's protocol and using the corresponding primers. All mutations made and their corresponding primers are summarized in Table 4. Plasmids harboring the desired mutation were identified by DNA sequence analysis as known in the art.

Table 4: Primers used for site-directed mutagenesis of consensus phytase

20 (Exchanged bases are highlighted in bold. The introduction of a restriction site is marked above the sequence. When a restriction site is written in parenthesis, the mentioned site was destroyed by introduction of the mutation.)

mutation	Primer set
25 Q50T	<div style="text-align: center;"><i>Kpn</i> I</div> <div>5'-CACTTGTGGGG<b>TAC</b>CTACTCTCCATACTTCTC-3'</div> <div>5'-GAGAAGTATGGAGAGTAG<b>GT</b>ACCCCAAGTG-3'</div>
30 Y54F	<div>5'-GGTCAATACTCTCCATTCTTCTTTGGAAG-3'</div> <div>5'-CTTCCAAAGAGAAGAATGGAGAGTATTGACC-3'</div>
E58A	<div>5'-CATACTTCTCTTTGGCAGACGAATCTGC-3'</div> <div>5'-GCAGATTCGTCTGCCAAAGAGAAGTATG-3'</div>



		<i>Aat</i> II
	D69K	5'-CTCCAGACGTCCCAAAGGACTGTAGAGTTAC-3' 5'-GTA ACTCTACAGTCCTTTGGGACGTCTGGAG-3'
		<i>Aat</i> II
5	D70G	5'-CTCCAGACGTCCAGACGGCTGTAGAGTTAC-3' 5'-GTA ACTCTACAGCCGTCTGGGACGTCTGGAG-3'
	K91A	5'-GATACCCAACTTCTTCTGCGTCTAAGGCTTACTCTG-3' 5'-CAGAGTAAGCCTTAGACGCAGAAGAAGTTGGGTATC-3'
10		<i>Sca</i> I
	A94K	5'-CTTCTAAGTCTAAGAAGTACTCTGCTTTG-3' 5'-CAAAGCAGAGTACTTCTTAGACTTAGAAG-3'
	A101R	5'-GCTTACTCTGCTTTGATTGAACGGATTCAAAAGAACGCTAC-3' 5'-GTAGCGTTCTTTTGAATCCGTTCAATCAAAGCAGAGTAAGC-3'
15		
	N134Q	5'-CCATTCGGTGAACAGCAAATGGTTAACTC-3' 5'-GAGTTAACCATTGTGCTGTTACCGAATGG-3'
		<i>Nru</i> I
20	K153N	5'-GATACAAGGCTCTCGCGAGAAACATTGTTC-3' 5'-GGAACAATGTTTCTCGCGAGAGCCTTGTATC-3'
		<i>Bss</i> HI
	I158V	5'-GATTGTTCCATTTCGTGCGCGCTTCTGGTTC-3' 5'-GAACCAGAAGCGCGCACGAATGGAACAATC-3'
		<i>Bcl</i> I
25	D197N	5'-CTCCAGTTATTAACGTGATCATTCCAGAAGG-3' 5'-CCTTCTGGAATGATCACGTTAATAACTGGAG-3'
		<i>Apa</i> I
	S187A	5'-GGCTGACCCAGGGGCCCAACCACACCAAGC-3' 5'-GCTTGGTGTGGTTGGGCCCCTGGGTCAGCC-3'
30		
	T214L	5'-CACTTTGGACCATGGTCTTTGTACTGCTTTTCG-3' 5'-CGAAAGCAGTACAAAGACCATGGTCCAAAGTG-3'
		<i>Avr</i> II
35	E222T	5'-GCTTTCGAAGACTCTACCCTAGGTGACGACGTTG-3' 5'-CAACGTCGTCACCTAGGGTAGAGTCTTCGAAAGC-3'

	V227A	5'-GGTGACGACGCTGAAGCTAACTTCAC-3' 5'-GTGAAGTTAGCTTCAGCGTCGTCACC-3'
		<i>Sac II</i>
5	L234V	5'-CTAACTTCACCGCGGTGTTTCGCTCCAG-3' 5'-CTGGAGCGAACACCGCGGTGAAGTTAG-3'
	A238P	5'-GCTTTGTTTCGCTCCACCTATTAGAGCTAGATTGG-3' 5'-CCAATCTAGCTCTAATAGGTGGAGCGAACAAAGC-3'
10		<i>Hpa I</i>
	T251N	5'-GCCAGGTGTTAACTTGACTGACGAAG-3' 5'-TTCGTCAGTCAAGTTAACACCTGGC-3'
		<i>Aat II</i>
15	Y259N	5'-GACGAAGACGTCGTAACTTGATGGAC-3' 5'-GTCCATCAAGTTAACGACGTCCTTCGTC-3'
		<i>Asp I</i>
	E267D	5'-GTCCATTTCGACACTGTCGCTAGAACTT C-3' 5'-GAAGTTCTAGCGACAGTGTCGAATGGAC-3'
20	E277Q	5'-CTGACGCTACTCAGCTGTCTCCATTC-3' 5'-GAATGGAGACAGCTGAGTAGCGTCAG-3'
	A283D	5'-GTCTCCATTCTGTGATTTGTTCACTCAC-3' 5'-GTGAGTGAACAAATCACAGAATGGAGAC-3'
25		<i>Ksp I</i>
	H287A	5'-GCTTTGTTCAACCGCGGACGAATGGAG-3' 5'-CTCCATTTCGTCGCGGTGAACAAAGC-3'
		<i>Bam HI</i>
30	R291I	5'-CACGACGAATGGATCCAATACGACTAC-3' 5'-GTAGTCGTATTGGATCCATTCGTCGTG-3'
		<i>Bsi WI</i>
	Q292A	5'-GACGAATGGAGAGCGTACGACTACTTG-3' 5'-CAAGTAGTCGTACGCTCTCCATTCGTC-3'
		<i>Hpa I</i>
35	A320V	5'-GGTGTTGGTTTCGTTAACGAATTGATTGC-3' 5'-GCAATCAATTCGTTAACGAAACCAACACC-3'
		( <i>Bgl II</i> )
	R329H	5'-GCTAGATTGACTCACTCTCCAGTTCAAG-3' 5'-CTTGAAGTGGAGAGTGAGTCAATCTAGC-3'

		<i>Eco</i> RV	
	S364T	5'-CTCACGACAACACTATGATATCTATTTTCTTC-3'	5'-GAAGAAAATAGATATCATAGTGTGTCGTGAG-3'
		<i>Nco</i> I	
5	I366V	5'-CGACAACTCCATGGTTTCTATTTTCTTCGC-3'	5'-GCGAAGAAAATAGAAACCATGGAGTTGTCG-3'
		<i>Kpn</i> I	
	A379K	5'-GTACAACGGTACCAAGCCATTGTCTAC-3'	5'-GTAGACAATGGCTTGGTACCGTTGTAC-3'
10	S396A	5'-CTGACGGTTACGCTGCTTCTTGGAC-3'	5'-GTCCAAGAAGCAGCGTAACCGTCAG-3'
	G404A	5'-CTGTTCCATTTCGCTGCTAGAGCTTAC-3'	5'-GTAAGCTCTAGCAGCGAATGGAACAG-3'
15	Q415E	5'-GATGCAATGTGAAGCTGAAAAGGAACC-3'	5'-GGTTCCTTTTCAGCTTCACATTGCATC-3'
		<i>Sal</i> I	
20	A437G	5'-CACGGTTGTGGTGTGCGACAAGTTGGG-3'	5'-CCCAACTTGTGCGACACCACAACCGTG-3'
		<i>Mun</i> I	
	A463E	5'-GATCTGGTGGCAATTGGGAGGAATGTTTCG-3'	5'-CGAAACATTCCTCCCAATTGCCACCAGATC-3'

25 and accordingly for other mutations.

The temperature optimum of the purified phytases, expressed in *Saccharomyces cerevisiae* (Example 7), was determined as outlined in Example 9. Table 5 shows the effect on the stability of consensus phytase for each mutation introduced.

30

Table 5: Stability effect of the individual amino acid replacements in consensus phytase-1  
 (+ or - means a positive, respectively, negative effect on the protein stability up to 1 °C, ++  
 and -- means a positive, respectively, negative effect on the protein stability between 1 and  
 3 °C; the number 10 or 11 corresponds to the consensus phytase sequence that suggests the  
 35 amino acid replacement.)

stabilizing		neutral		destabilizing	
mutation	effect	mutation	effect	mutation	effect
E58A (10)	+	D69A	±	Y54F (10)	-
D69K (11)	+	D70G (10)	±	V73I	-
D197N (10)	+	N134Q (10)	±	A94K (10)	-
T214L (10)	++	G186H	±	A101R (11)	-
E222T (11)	++	S187A (10)	±	K153N (11)	-
E267D (10)	+	T214V	±	I158V (10)	--
R291I*	+	T251N (10)	±	G203A	--
R329H (10)	+	Y259N (10)	±	G205S	-
S364T (10)	++	A283D (10)	±	A217V	-
A379K (11)	+	A320V (10)	±	V227A (11)	--
G404A (10)	++	K445T	±	L234V (10)	-
		A463E (10)	±	A238P (10)	--
				E277Q (10)	-
				H287A (11)	-
				Q292A (10)	-
				I366V (10)	-
				S396A (10)	--
				Q415E (11)	-
				A437G (10)	--
				E451R	--

\*: This amino acid replacement was found in another round of mutations.

We combined eight positive mutations (E58A, D197N, E267D, R291I, R329H, S364T, A379K, G404A) in the consensus phytase using the primers and the technique mentioned above in this example. Furthermore, the mutations Q50T and K91A were introduced which mainly influence the catalytical characteristics of the phytase (see patent application EP 97810175.6 and EP 97112688 as well as Example 9). The DNA and amino acid sequence of the resulting phytase gene (consensus phytase-thermo[8]-Q50T-K91A) is shown in Figure 7. In this way, the temperature optimum and the melting point of the consensus phytase was increased by 7 °C (Figure 15, 16, 17).

Using the results of Table 5, we further improved the thermostability of consensus phytase 10 by the following back mutations K94A, V158I, and A396S that revealed a strong negative influence on the stability of consensus phytase. The resulting protein is phytase-10-thermo [3]. Furthermore, we introduced the mutations Q50T and K91A which mainly influence the catalytical characteristics of consensus phytase (see patent application EP 97810175.6 and EP 97112688 as well as Example 9 and Figure 14 and 15). The resulting DNA and amino acid sequence is shown in Figure 8. The optimized phytase showed a 4 °C higher temperature optimum and melting point than consensus phytase 10 (Figure 12 and 13). Furthermore, the phytase has also a strongly increased specific activity with phytate as substrate of 250 U/mg at pH 5.5 (Figure 14).

#### Example 4

Stabilization of the phytase of *A. fumigatus* ATCC 13073 by replacement of amino acid residues with the corresponding consensus phytase-1 and consensus phytase-10 residues

At six typical positions where the *A. fumigatus* 13073 is the only or nearly the only phytase in the alignment of Figure 1 that does not contain the corresponding consensus phytase amino acid residue, the non-consensus amino acid residue was replaced by the consensus one. In a first round, the following amino acids were substituted in *A. fumigatus* 13073 phytase, containing the Q27T substitution and the signal sequence of *A. terreus* cbs.116.46 phytase (see European Patent Application No. 97810175.6 and Figure 9):

F55(28)Y, V100(73)I, F114(87)Y, A243(220)L, S265(242)P, N294(282)D.

The numbers in parentheses confer to the numbering of Figure 1.

In a second round, four of the seven stabilizing amino acid exchanges (E59A, R329H, S364T, G404A) found in the consensus phytase-10 sequence and, tested as single mutation in consensus phytase-1 (Table 5), were additionally introduced into the *A. fumigatus* a-

mutant. Furthermore, the amino acid replacement S126N, shown to reduce the protease susceptibility of the phytase, was introduced.

The mutations were introduced as described in example 3 (see Table 6) and expressed as described in example 6 to 8. The resulting *A. fumigatus* 13073 phytase variants were called  
5 a-mutant and  $\beta$ -mutant-E59A-S126N-R329H-S364T-G404A.

The temperature optimum (60 °C, Figure 20) and the melting point (67.0 °C, Figure 19) of the *A. fumigatus* 13073 phytase  $\beta$ -mutant was increased by 5 °C in comparison to the values of the wild-type (temperature optimum: 55 °C,  $T_m$ : 60 °C). The five additional amino acid replacements further increased the temperature optimum by 3 °C (Figure 20).

10 Table 6: Mutagenesis primers for stabilization of *A. fumigatus* phytase ATCC 13073

Mutation	Primer
F55Y	5'-CACGTA <del>CT</del> CGCCATACTTTTCGCTCGAG-3' 5'-CTCGAGCGAAAAGTATGGCGAGTACGTG-3'
	( <i>Xho</i> I)
15 E58A	5'-CCATACTTTTCGCTCGCGGACGAGCTGTCCGTG-3' 5'-CACGGACAGCTCGTCCGCGAGCGAAAAGTAGG-3'
V100I	5'-GTATAAGAAGCTTATTACGGCGATCCAGGCC-3' 5'-GGCCTGGATCGCCGTAATAAGCTTCTTATAC-3'
20 F114Y	5'-CTTCAAGGGCAAGTACGCCTTTTTGAAGACG-3' 5'-CGTCTTCAAAAAGGCGTACTTGCCCTTGAAG-3'
A243L	5'-CATCCGAGCTCGCCTCGAGAAGCATCTTC-3'
25 S265P	5'-GAAGATGCTTCTCGAGGCGAGCTCGGATG-3' 5'-CTAATGGA TGTGTCCGTTTGATACGGTAG-3' 5'-CTACCGTATCAAACGGACACATGTCCATTAG-3'

N294D 5'-GTGGAAGAAGTACGACTACCTTCAGTC-3'  
5'-GACTGAAGGTAGTCGTA CTTCTTCCAC-3'

(Mlu I)

5 R329H 5'-GCCCCGGTTGACGCAATTCGCCAGTGCAGG-3'  
5'-CCTGCACTGGCGAATGCGTCAACCGGGC-3'

Nco I

S364T 5'-CACACGACAACACCATGGTTTCCATCTTC-3'  
5'-GAAGATGGAAACCATGGTGTGTCGTGTG-3'

10 (Bss HI)

G404A 5'-GTGGTGCCTTTCGCCGCGGAGCCTACTTC-3'  
5'-GAAGTAGGCTCGCGCGCGAAAGGCACCAC-3'

#### Example 5

15 Introduction of the active site amino acid residues of the *A. niger* NRRL 3135  
phytase into the consensus phytase-1

We used the crystal structure of the *Aspergillus niger* NRRL 3135 phytase to define all active site amino acid residues (see Reference Example and EP 97810175.6). Using the alignment of Figure 1, we replaced the following active site residues and additionally the not identical adjacent ones of the consensus phytase by that of the *A. niger* phytase:

20 S89D, S92G, A94K, D164S, P201S, G203A, G205S, H212P, G224A, D226T, E255T, D256E, V258T, P265S, Q292H, G300K, Y305H, A314T, S364G, M365I, A397S, S398A, G404A, and A405S

The new protein sequence consensus phytase -7 was backtranslated into a DNA sequence (Figure 10) as described in Example 1. The corresponding gene (*fcp7*) was generated as  
25 described in Example 1 using the following oligonucleotide mixes:

Mix 1.7: CP-1, CP-2, CP-3, CP-4.7, CP-5.7, CP-6, CP-7, CP-8.7, CP-9, CP-10.7

Mix 2.7: CP-9, CP-10.7, CP-11.7, CP-12.7, CP-13.7, CP-14.7, CP-15.7, CP-16, CP-17.7, CP-18.7, CP-19.7, CP-20, CP-21, CP-22.

The DNA sequences of the oligonucleotides are indicated in Figure 3. The newly  
30 synthesized oligonucleotides are additionally marked by number 7. After assembling of the

oligonucleotides using the same PCR primers as mentioned in Example 1, the gene was cloned into an expression vector as described in Examples 6-8.

The pH-profile determined after expression in *H. polymorpha* and purification was shifted into the acidic range of the pH-spectrum showing an optimum at pH 4.5-5.0 (see Figure 18). The enzyme had a broad pH-optimum reaching at least 60% of its maximum activity from pH 2.5 to pH 6.0. Up to pH 5.0, the profile resembled the profile of the *A. niger* NRRL 3135 phytase. However, below pH 5.0 it lacked the typical low at pH 4.0 of the profile of *A. niger* phytase.

#### Example 6

#### 10 Expression of the consensus phytase genes in *Hansenula polymorpha*

The phytase expression vectors, used to transform *H. polymorpha* RB11 (Gellissen *et al.*, 1994), was constructed by inserting the *Eco* RI fragment of pBsk-*fcp* or variants thereof into the multiple cloning site of the *H. polymorpha* expression vector pFPMT121, which is based on an *ura3* selection marker from *S. cerevisiae*, a formate dehydrogenase (FMD) promoter element and a methanol oxidase (MO) terminator element from *H.*  
15 *polymorpha*. The 5' end of the *fcp* gene is fused to the FMD promoter, the 3' end to the MOX terminator (Gellissen *et al.*, 1996; EP 0299 108 B). The resulting expression vector are designated pFPMT*fcp*, pFPMT*fcp10*, pFPMT*fcp7*.

The constructed plasmids were propagated in *E. coli*. Plasmid DNA was purified  
20 using standard state of the art procedures. The expression plasmids were transformed into the *H. polymorpha* strain RP11 deficient in orotidine-5'-phosphate decarboxylase (*ura3*) using the procedure for preparation of competent cells and for transformation of yeast as described in Gellissen *et al.* (1996). Each transformation mixture was plated on YNB (0.14% w/v Difco YNB and 0.5% ammonium sulfate) containing 2% glucose and 1.8%  
25 agar and incubated at 37 °C. After 4 to 5 days individual transformant colonies were picked and grown in the liquid medium described above for 2 days at 37 °C. Subsequently, an aliquot of this culture was used to inoculate fresh vials with YNB-medium containing 2% glucose. After seven further passages in selective medium, the expression vector integrates into the yeast genome in multimeric form. Subsequently, mitotically stable transformants  
30 were obtained by two additional cultivation steps in 3 ml non-selective liquid medium (YPD, 2% glucose, 10 g yeast extract, and 20 g peptone). In order to obtain genetically homogeneous recombinant strains an aliquot from the last stabilization culture was plated on a selective plate. Single colonies were isolated for analysis of phytase expression in YNB containing 2% glycerol instead of glucose to derepress the *fmd* promoter. Purification  
35 of the consensus phytases was done as described in Example 7.



Example 7

Expression of the consensus phytase genes in *Saccharomyces cerevisiae* and purification of the phytases from culture supernatant

The consensus phytase genes were isolated from the corresponding Bluescript-plasmid (pBsk $\overline{fcp}$ , pBSK $\overline{fcp10}$ , pBsk $\overline{fcp7}$ ) and ligated into the *Eco* RI sites of the expression cassette of the *Saccharomyces cerevisiae* expression vector pYES2 (Invitrogen, San Diego, CA, USA) or subcloned between the shortened GAPFL (glyceraldehyde-3-phosphate dehydrogenase) promoter and the *pho5* terminator as described by Janes *et al.* (1990). The correct orientation of the gene was checked by PCR. Transformation of *S. cerevisiae* strains. e. g. INVSc1 (Invitrogen, San Diego, CA, USA) was done according to Hinnen *et al.* (1978). Single colonies harboring the phytase gene under the control of the GAPFL promoter were picked and cultivated in 5 ml selection medium (SD-uracil, Sherman *et al.*, 1986) at 30°C under vigorous shaking (250 rpm) for one day. The preculture was then added to 500 ml YPD medium (Sherman *et al.*, 1986) and grown under the same conditions. Induction of the *gal1* promoter was done according to manufacturer's instruction. After four days of incubation cell broth was centrifuged (7000 rpm, GS3 rotor, 15 min, 5°C) to remove the cells and the supernatant was concentrated by way of ultrafiltration in Amicon 8400 cells (PM30 membranes) and ultrafree-15 centrifugal filter devices (Biomax-30K, Millipore, Bedford, MA, USA). The concentrate (10 ml) was desalted on a 40 ml Sephadex G25 Superfine column (Pharmacia Biotech, Freiburg, Germany), with 10 mM sodium acetate, pH 5.0, serving as elution buffer. The desalted sample was brought to 2 M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> and directly loaded onto a 1 ml Butyl Sepharose 4 Fast Flow hydrophobic-interaction chromatography column (Pharmacia Biotech, Freiburg, Germany) which was eluted with a linear gradient from 2 M to 0 M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> in 10 mM sodium acetate, pH 5.0. Phytase was eluted in the break-through, concentrated and loaded on a 120 ml Sephacryl S-300 gel permeation chromatography column (Pharmacia Biotech, Freiburg, Germany). Consensus phytase and consensus phytase -7 eluted as a homogeneous symmetrical peak and was shown by SDS-PAGE to be approx. 95% pure.

Example 8

Expression of the consensus phytase genes in *Aspergillus niger*

The Bluescript-plasmids pBsk $\overline{fcp}$ , pBSK $\overline{fcp10}$ , and pBsk $\overline{fcp7}$  were used as template for the introduction of a *Bsp* HI-site upstream of the start codon of the genes and an *Eco* RV-site downstream of the stop codon. The Expand™ High Fidelity PCR Kit (Boehringer Mannheim, Mannheim, Germany) was used with the following primers:

Primer Asp-1:

*Bsp* HI

5'-TATATCATGAGCGTGTTCGTCGTGCTACTGTTC-3'

Primer Asp-2 used for cloning of *fcp* and *fcp7*:

5

*Eco* RV

3'-ACCCGACTTACAAAGCGAATTCTATAGATATAT-5'

Primer Asp-3 used for cloning of *fcp10*:

*Eco* RV

3'-ACCCTTCTTACAAAGCGAATTCTATAGATATAT-5'

- 10        The reaction was performed as described by the supplier. The PCR-amplified *fcp*-genes had a new *Bsp* HI site at the start codon, introduced by primer Asp-1, which resulted in a replacement of the second amino acid residue glycine by serine. Subsequently, the DNA-fragment was digested with *Bsp* HI and *Eco* RV and ligated into the *Nco* I site downstream of the glucoamylase promoter of *Aspergillus niger* (*glaA*) and the *Eco* RV site
- 15        upstream of the *Aspergillus nidulans* tryptophan C terminator (*trpC*) (Mullaney *et al.*, 1985). After this cloning step, the genes were sequenced to detect possible failures introduced by PCR. The resulting expression plasmids which basically corresponds to the pGLAC vector as described in Example 9 of EP 684 313, contained the orotidine-5'-phosphate decarboxylase gene (*pyr4*) of *Neurospora crassa* as a selection marker.
- 20        Transformation of *Aspergillus niger* and expression of the consensus phytase genes was done as described in EP 684 313. The consensus phytases were purified as described in Example 7.

#### Example 9

##### Determination of phytase activity and of temperature optimum

- 25        Phytase activity was determined basically as described by Mitchell *et al* (1997). The activity was measured in an assay mixture containing 0.5% phytic acid ( $\approx 5$  mM) in 200 mM sodium acetate, pH 5.0. After 15 min of incubation at 37 °C, the reaction was stopped by addition of an equal volume of 15% trichloroacetic acid. The liberated phosphate was quantified by mixing 100  $\mu$ l of the assay mixture with 900  $\mu$ l H<sub>2</sub>O and 1 ml of 0.6 M
- 30        H<sub>2</sub>SO<sub>4</sub>, 2% ascorbic acid and 0.5% ammonium molybdate. Standard solutions of potassium phosphate were used as reference. One unit of enzyme activity was defined as the amount of enzyme that releases 1  $\mu$ mol phosphate per minute at 37 °C. The protein

concentration was determined using the enzyme extinction coefficient at 280 nm calculated according to Pace et al (1995): consensus phytase, 1.101; consensus phytase 7, 1.068; consensus phytase 10, 1.039.

5 In case of pH-optimum curves, purified enzymes were diluted in 10 mM sodium acetate, pH 5.0. Incubations were started by mixing aliquots of the diluted protein with an equal volume of 1% phytic acid ( $\approx 10$  mM) in a series of different buffers: 0.4 M glycine/HCl, pH 2.5; 0.4 M acetate/NaOH, pH 3.0, 3.5, 4.0, 4.5, 5.0, 5.5; 0.4 M imidazole/HCl, pH 6.0, 6.5; 0.4 M Tris/HCl pH 7.0, 7.5, 8.0, 8.5, 9.0. Control experiments showed that pH was only slightly affected by the mixing step. Incubations were performed for 15 min at 37 °C as  
10 described above.

For determinations of the substrate specificities of the phytases, phytic acid in the assay mixture was replaced by 5 mM concentrations of the respective phosphate compounds. The activity tests were performed as described above.

For determination of the temperature optimum, enzyme (100  $\mu$ l) and substrate  
15 solution (100  $\mu$ l) were pre-incubated for 5 min at the given temperature. The reaction was started by addition of the substrate solution to the enzyme. After 15 min incubation, the reaction was stopped with trichloroacetic acid and the amount of phosphate released was determined.

The pH-optimum of the original consensus phytase was around pH 6.0-6.5 (70  
20 U/mg). By introduction of the Q50T mutation, the pH-optimum shifted to pH 6.0 (130 U/mg). After introduction of K91A, the pH optimum shifted one pH-unit into the acidic pH-range showing a higher specific activity between pH 2.5 and pH 6.0. That was shown for the stabilized mutants and for consensus phytase-10, too (Figure 14 and 15).

Consensus phytase-7, which was constructed to transfer the catalytic characteristics of the  
25 *A. niger* phytase NRRL 3135 into the consensus phytase, had a pH-profile which is shifted into the acidic range of the pH-spectrum showing an optimum between pH 4.5 and 5.0 (see Figure 19). The enzyme had a broad pH-optimum reaching at least 60% of its increased maximum activity from pH 2.5 to pH 6.0. The substrate spectrum, too, resemble more to that of the *A. niger* NRRL 3135 phytase than to the consensus phytase-1.

30 The temperature optimum of consensus phytase-1 (71 °C) was 16-26 °C higher than the temperature optimum of the wild-type phytases (45-55 °C, Table 7) which were used to calculate the consensus sequence. The improved consensus phytase-10 showed a further

increase of its temperature optimum to 80 °C (Figure 11). The temperature optimum of the consensus phytase-1-thermo[8] was found in the same range (78 °C) using the supernatant of an overproducing *S. cerevisiae* strain. The highest temperature optimum reached of 82 °C was determined for consensus phytase-10-thermo-Q50T-K91A.

- 5 **Table 7:** Temperature optimum and  $T_m$ -value of consensus phytase and of the phytases from *A. fumigatus*, *A. niger*, *E. nidulans*, and *M. thermophila*. The determination of the temperature optimum was performed as described in Example 9. The  $T_m$ -values were determined by differential scanning calorimetry as described in Example 10.

phytase	temperature optimum [°C]	$T_m$ [°C]
Consensus phytase-10-thermo-Q50T-K91A	82	89.3
Consensus phytase-10-thermo-Q50T	82	88.6
Consensus phytase-10	80	85.4
Consensus phytase-1-thermo[8]-Q50T	78	84.7
Consensus phytase-1-thermo[8]-Q50T-K91A	78	85.7
Consensus phytase-1	71	78.1
<i>A. niger</i> NRRL3135	55	63.3
<i>A. fumigatus</i> 13073	55	62.5
<i>A. fumigatus</i> 13073 $\alpha$ -mutant	60	67.0
<i>A. fumigatus</i> 13073 $\alpha$ -mutant (optimized)	63	-
<i>A. terreus</i> 9A-1	49	57.5
<i>A. terreus</i> cbs.116.46	45	58.5
<i>E. nidulans</i>	45	55.7
<i>M. thermophila</i>	55	n. d.
<i>T. thermophilus</i>	45	n. d.

### Example 10

#### Determination of the melting point by differential scanning calorimetry (DSC)

In order to determine the unfolding temperature of the phytases, differential scanning  
5 calorimetry was applied as previously published by Brugger et al (1997). Solutions of 50-  
60 mg/ml homogeneous phytase were used for the tests. A constant heating rate of 10 °  
C/min was applied up to 90-95 °C.

The determined melting points reflect the results obtained for the temperature  
optimums (Table 7). The most stable consensus phytase designed is consensus phytase-10-  
10 thermo-Q50T-K91A showing a melting temperature under the choosen condition of 89.3 °  
C. This is 26 to 33.6 °C higher than the melting point of the wild-type phytases used.

### Example 11

#### Transfer of basidiomycete phytase active site into consensus phytase-10-thermo- Q50T-K91A

15 As described previously (Example 3), mutations derived from the basidiomycete  
phytase active site were introduced into the consensus phytase 10. The following five  
constructs a) to e) were prepared:

- a) This construct is called consensus phytase 12, and it comprises a selected number of  
active site residues of the basidio consensus sequence, its amino acid sequence  
20 (consphy12) is shown in Fig. 21 (the first 26 amino acids forms the signal peptide,  
amended positions are underlined);
- b) a cluster of mutations (Cluster II) was transferred to the consensus 10 sequence, viz.:  
S80Q, Y86F, S90G, K91A, S92A, K93T, A94R, Y95I;
- c) analogously, another cluster of mutations (Cluster III) was transferred, viz.: T129V,  
25 E133A, Q143N, M136S, V137S, N138Q, S139A;
- d) analogously, a further cluster of mutations (Cluster IV) was transferred, viz.: A168D,  
E171T, K172N, F173W;

e) and finally, a further cluster of mutations (Cluster V) was transferred, viz.: Q297G, S298D, G300D, Y305T.

These constructs were expressed as described in Examples 6 to 8.

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Claims

1. A process for the preparation of a consensus protein, whereby such process is characterized by the following steps:

- 5 a) at least three, preferably four amino acid sequences are aligned by any standard alignment program known in the art,
- b) amino acids at the same position according to such alignment are compared regarding their evolutionary similarity by any standard program known in the art, whereas the degree of similarity provided by such a program which defines the least similarity of the amino acids that is used for the determination of an amino acid of corresponding positions is set to a less stringent number and the parameters are set in such a way that it is possible for the program to determine from only 2 identical amino acids at a corresponding position an amino acid for the consensus protein; however, if among the compared amino acid sequences are sequences that show a much higher degree of similarity to each other than to the residual sequences, these sequences are represented by their consensus sequence determined as defined in the same way as in the present process for the consensus sequence of the consensus protein or a vote weight of 1 divided by the number of such sequences is assigned to every of -those sequences,
- 10 c) in case no common amino acid at a defined position is identified by the program, any of the amino acids, preferably the most frequent amino acid of all such sequences is selected,
- 20 d) once the consensus sequence has been defined, such sequence is back-translated into a DNA sequence, preferably by using a codon frequency table of the organism in which expression should take place,
- 25 e) the DNA sequence is synthesized by methods known in the art and used either integrated into a suitable expression vector or by itself to transform an appropriate host cell,
- f) the transformed host cell is grown under suitable culture conditions and the consensus protein is isolated from the host cell or its culture medium by methods known in the art.
- 30

2. A process as claimed in claim 1 wherein the program used for the comparison of amino acids at a defined position regarding their evolutionary similarity is the program "PRETTY".

3. A process as claimed in claims 1 or 2, wherein

in a first step a consensus sequence is determined from a number of highly homologous sequences according to steps a), b) and c) of claim 1,

in a second step the amino acid sequence of another protein which is homologous to the  
5 consensus sequence is compared with the consensus sequence and

in a third step only those amino acid residues are replaced in the amino acid sequence of the other protein which clearly differ from the consensus sequence of this protein family calculated under moderately stringent conditions whereas at all positions of the alignment where no preferred single amino acid can be determined under moderately stringent  
10 conditions the amino acids of the other protein remain unchanged.

4. A process as claimed in any one of claims 1-3, wherein

in a first step a consensus sequence is determined from homologous sequences according to steps a), b) and c) of claim 1,

in a second step the active center of the protein comprising all amino acid residues that are  
15 involved in forming the active center is determined in the consensus sequence and in the sequence of a homologous protein as well and

in a third step some or all of the amino acids that form the active center of the homologous protein are inserted in the backbone of the consensus sequence.

5. A process as claimed in claim 4, wherein the active center of the protein is  
20 determined by using an analysis of the three-dimensional structure of the protein.

6. A process as claimed in claims 4 and 5, wherein the homologous protein is an enzyme.

7. A process as claimed in claims 1 to 6, wherein the defined protein family is the family of phytases.

25 8. A process as claimed in claim 7, wherein the phytases are of fungal origin.

9. A process as claimed in claims 7 or 8, wherein the amino acid sequence of the phytase is changed by the introduction of at least one mutation selected from the group consisting of

E58A	F54Y
D69K	I73V
D197N	K94A
T214L	R101A
E222T	N153K
E267D	V158I
R291I	A203G
R329H	S205G
S364T	V217A
A379K	A227V
G404A	V234L
	P238A
	Q277E
	A287H
	A292Q
	V366I
	A396S
	E415Q
	G437A
	E451R

whereby the number represents the position in the consensus phytase sequence or a corresponding residue according to an alignment as shown in Fig. 1 when 26 amino acids (signal sequence) are added to the sequences shown in Fig. 1 and the letter before the number represents the amino acid in the phytase which is replaced by the amino acid  
5 behind the number.

10. A process as claimed in any one of claims 1 to 9, wherein the host cell is of eukaryotic origin.

11. A process as claimed in claim 10, wherein eukaryotic means fungal, preferably *Aspergillus* or yeast, preferably *Saccharomyces* or *Hansenula*.

12. A consensus protein obtainable preferably obtained by a process as claimed in any one of claims 1 to 11.

5        13. A consensus protein which comprises the amino acid sequence shown in Figure 2 or any variants or muteins thereof (consensus phytase-1).

14. A mutein of the consensus protein of claim 13 characterized therein that in the amino acid sequence of Figure 2 the following replacements have been effected Q50L, Q50T, Q50G, Q50T-Y51N, Q50L-Y51N or Q50T-K91A.

10        15. A consensus protein which comprises the amino acid sequence shown in Figure 4 having the designation consensus phytase 10 (Fcp10) and any variants or muteins thereof.

16. A consensus protein which comprises the amino acid sequence shown in Figure 6 having the designation Consensus seq. 11 and any variant or mutein thereof.

15        17. A consensus protein which comprises the amino acid sequence shown in Figure 10 (consensus phytase 7) and any variant or mutein thereof.

18. A consensus protein which comprises the amino acid sequence shown in Figure 21 (consensus phytase 12) and any variant or mutein thereof.

19. A consensus protein which comprises the amino acid sequence shown in Figure 3 (basidio consensus) and any variant or mutein thereof.

20        20. A phytase being selected from amongst: *A. fumigatus* ATCC 13073 alpha-mutant; *A. fumigatus* ATCC 13073 alpha-mutant-(E59A-S126N-R329H, S364T-G404A)-Q27T; *A. fumigatus* ATCC 13073 alpha-mutant-(E59A, S126N-R329H-S364T-G404A)-Q27T-K68A, preferably the latter.

25        21. A food, feed or pharmaceutical composition comprising a consensus protein as claimed in any of the claims 12 to 17.

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Figure 1

		1
50		
5	A. terreus 9A-1	KhSDCNSVDh GYQCFPELSH kWGLYAPYFS
	LQDESPFPID VPEDChITFV	
	A. terreus cbs	NhsDCTSVDr GYQCFPELSH kWGLYAPYFS
	LQDESPFPID VPDDChITFV	
	A. niger var. awamori	NqsTCDTVDQ GYQCFSETSH LWGQYAPFFS
10	LANESAISPD VPAGCrVTFA	
	A. niger T213	NqsSCDTVdQ GYQCFSETSH LWGQYAPFFS
	LANESVISPD VPAGCrVTFA	
	A. niger NRRL3135	NqsSCDTVdQ GYQCFSETSH LWGQYAPFFS
	LANESVISPE VPAGCrVTFA	
15	A. fumigatus 13073	GskSCDTVd1 GYQCsPATSH LWGQYSPFFS
	LEDELSVSSK LPKDCrITLV	
	A. fumigatus 32722	GskSCDTVd1 GYQCsPATSH LWGQYSPFFS
	LEDELSVSSK LPKDCrITLV	
	A. fumigatus 58128	GskSCDTVd1 GYQCsPATSH LWGQYSPFFS
20	LEDELSVSSK LPKDCrITLV	
	A. fumigatus 26906	GskSCDTVd1 GYQCsPATSH LWGQYSPFFS
	LEDELSVSSK LPKDCrITLV	
	A. fumigatus 32239	GskACDTVE1 GYQCsPGTSH LWGQYSPFFS
	LEDELSVSSD LPKDCrVTFV	
25	E. nidulans	QNHSCNTADG GYQCFPNVSH VWGQYSPYFS
	IEQESAISd VPHGCeVTFV	
	T. thermophilus	DSHSCNTVEG GYQCrPEISH sWGQYSPFFS
	LADQSEISPD VPQNCKITFV	
	M. thermophila	ESRPCDTpD1 GFQCgTAISH FWGQYSPYFS
30	VpSElDaS.. IPDDCeVTFA	
	Consensus	NSHSCDTVdG GYQCFPEISH LWGQYSPYFS
	LEDESAISPD VPDDC-VTFV	
	Consensus phytase	NSHSCDTVdG GYQCFPEISH LWGQYSPYFS
35	LEDESAISPD VPDDCrVTFV	
		51
100		
40	A. terreus 9A-1	QVLARHGArS PThSKtKAYA AtIAAIQKSA
	TaFpGKYAFL QSYNYSLDSE	
	A. terreus cbs	QVLARHGArS PTDSKtKAYA AtIAAIQKNA
	TaLpGKYAFL KSYNYSMGSE	
	A. niger var. awamori	QVLSRHGARY PTESKgKkYS ALIEEIQQNV
45	TtFDGKYAFL KTYNYSLGAD	
	A. niger T213	QVLSRHGARY PTESKgKkYS ALIEEIQQNV
	TtFDGKYAFL KTYNYSLGAD	
	A. niger NRRL3135	QVLSRHGARY PTDSKgKkYS ALIEEIQQNA
	TtFDGKYAFL KTYNYSLGAD	
50	A. fumigatus 13073	QVLSRHGARY PTSSKsKkYK kLVTAIQaNA
	TdFKGKFAFL KTYNYTLGAD	
	A. fumigatus 32722	QVLSRHGARY PTSSKsKkYK kLVTAIQaNA
	TdFKGKFAFL KTYNYTLGAD	
	A. fumigatus 58128	QVLSRHGARY PTSSKsKkYK kLVTAIQaNA
55	TdFKGKFAFL KTYNYTLGAD	

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*A. fumigatus* 26906 QVLSRHGARY PTSSKsKkYK kLVTAIQaNA  
 TdFKGKFAFL KTYNYTLGAD  
*A. fumigatus* 32239 QVLSRHGARY PTASKsKkYK kLVTAIQKNA  
 TeFKGKFAFL ETYNYTLGAD  
 5 *E. nidulans* QVLSRHGARY PTESKsKAYS GLIEAIQKNA  
 TsFwGQYAFI ESYNYTLGAD  
*T. thermophilus* QLLSRHGARY PTSSKtELYS QLISrIQKTA  
 TaYKGyYAFI KDYrYqLGAN  
*M. thermophila* QVLSRHGARA PtlKRaaSYv DLIDrIHhGA  
 10 IsYgPgYEFL RTYDYTLGAD  
  
 Consensus QVLSRHGARY PTSSK-KAYS ALIEAIQKNA T-  
 FKGYAFI KTYNYTLGAD  
 Consensus phytase QVLSRHGARY PTSSKSKAYS ALIEAIQKNA  
 15 TAFKGYAFI KTYNYTLGAD

101

150  
 20 *A. terreus* 9A-1 ELTPFGGrNQL rDlGaQFYeR YNALTRhInP  
 FVRATDASRV hESAekFVEG  
*A. terreus* cbs NLTPFGGrNQL qDlGaQFYRR YDTLTRhInP  
 FVRAADSSRV hESAekFVEG  
*A. niger* var. *awamori* DLTPFGEQEL VNsgIKFYQR YESLTRNIIP  
 25 FIRSSGSSRV IASGEKFIEG  
*A. niger* T213 DLTPFGEQEL VNsgIKFYQR YESLTRNIIP  
 FIRSSGSSRV IASGEKFIEG  
*A. niger* NRRL3135 DLTPFGEQEL VNsgIKFYQR YESLTRNIVP  
 FIRSSGSSRV IASGKKFIEG  
 30 *A. fumigatus* 13073 DLTPFGEQQL VNsgIKFYQR YKALARSVVP  
 FIRASGSDRV IASGEKFIEG  
*A. fumigatus* 32722 DLTPFGEQQL VNsgIKFYQR YKALARSVVP  
 FIRASGSDRV IASGEKFIEG  
*A. fumigatus* 58128 DLTPFGEQQL VNsgIKFYQR YKALARSVVP  
 35 FIRASGSDRV IASGEKFIEG  
*A. fumigatus* 26906 DLTAfGEQQL VNsgIKFYQR YKALARSVVP  
 FIRASGSDRV IASGEKFIEG  
*A. fumigatus* 32239 DLTPFGEQQM VNsgIKFYQK YKALAgSVVP  
 FIRSSGSDRV IASGEKFIEG  
 40 *E. nidulans* DLTiFGENQM VDsgAKFYRR YKNLARKnTP  
 FIRASGSDRV VASAEKFIEG  
*T. thermophilus* DLTPFGENQM IQlGIKFYnH YKSLARNAVP  
 FVRCSGSDRV IASGrIFIEG  
*M. thermophila* ELTRtGQQQM VNsgIKFYRR YRALARKsIP  
 45 FVRTAGqDRV VhSAENFTQG  
  
 Consensus DLTPFGENQM VNsgIKFYRR YKALARK-VP  
 FVRASGSDRV IASAEKFIEG  
 Consensus phytase DLTPFGENQM VNsgIKFYRR YKALARKIVP  
 50 FIRASGSDRV IASAEKFIEG

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		151
	200	
	<i>A. terreus</i> 9A-1	FQTARqDDHh ANpHQSPPrV DVaIPEGSAY
	NNTLEHSICT AFES...STV	
5	<i>A. terreus</i> cbs	FQNARqGDPH ANpHQSPPrV DVVIPEGTAY
	NNTLEHSICT AFEA...STV	
	<i>A. niger</i> var. <i>awamori</i>	FQSTKLkDPr AqpgQSSPkI DVVISEASSs
	NNTLDPGTCT VFED...SEL	
	<i>A. niger</i> T213	FQSTKLkDPr AqpgQSSPkI DVVISEASSs
10	NNTLDPGTCT VFED...SEL	
	<i>A. niger</i> NRRL3135	FQSTKLkDPr AqpgQSSPkI DVVISEASSs
	NNTLDPGTCT VFED...SEL	
	<i>A. fumigatus</i> 13073	FQqAKLADPG A.TNRAAPAI SVIIPESETF
	NNTLDHGVCT kFEA...SQL	
15	<i>A. fumigatus</i> 32722	FQqAKLADPG A.TNRAAPAI SVIIPESETF
	NNTLDHGVCT kFEA...SQL	
	<i>A. fumigatus</i> 58128	FQqAKLADPG A.TNRAAPAI SVIIPESETF
	NNTLDHGVCT kFEA...SQL	
	<i>A. fumigatus</i> 26906	FQqAKLADPG A.TNRAAPAI SVIIPESETF
20	NNTLDHGVCT kFEA...SQL	
	<i>A. fumigatus</i> 32239	FQqANVADPG A.TNRAAPVI SVIIPESETY
	NNTLDHSVCT NFEA...SEL	
	<i>E. nidulans</i>	FRKAQLhDHG S..gQATPVV NVIIPeIDGF
	NNTLDHSTCV SFEN...DER	
25	<i>T. thermophilus</i>	FQSAKvldPh SDkHDAPPTI NVIIeEGPSY
	NNTLDtGSCP VFED...SSg	
	<i>M. thermophila</i>	FHSALLADRG STvRPTlPyd mVVIPETAGa
	NNTLHNDlCT AFEEgpySTI	
30	Consensus	FQSAKLADPG S-PHQASPVI NVIIPESGSGY
	NNTLDHGTCT AFED---SEL	
	Consensus phytase	FQSAKLADPG SQPHQASPVI DVIIPESGSGY
	NNTLDHGTCT AFED...SEL	
35		
		201
	250	
	<i>A. terreus</i> 9A-1	GDDAVANFTA VFAPAIaQRL EADLPGVqLS
	TDDVVnLMAM CPFETVSlTD	
40	<i>A. terreus</i> cbs	GDAADNFTA VFAPAIakRL EADLPGVqLS
	ADDVVnLMAM CPFETVSlTD	
	<i>A. niger</i> var. <i>awamori</i>	ADTVEANFTA TFAPSIRQRL ENDLsgVTLT
	DTEVTyLMDM CSFDTIstST	
	<i>A. niger</i> T213	ADTVEANFTA TFAPSIRQRL ENDLsgVTLT
45	DTEVTyLMDM CSFDTIstST	
	<i>A. niger</i> NRRL3135	ADTVEANFTA TFVPSIRQRL ENDLsgVTLT
	DTEVTyLMDM CSFDTIstST	
	<i>A. fumigatus</i> 13073	GDEVAANFTA lFAPDIRARA EkHLPGVTLT
	DEDVVsLMDM CSFDTVARTS	
50	<i>A. fumigatus</i> 32722	GDEVAANFTA lFAPDIRARA EkHLPGVTLT
	DEDVVsLMDM CSFDTVARTS	
	<i>A. fumigatus</i> 58128	GDEVAANFTA lFAPDIRARA EkHLPGVTLT
	DEDVVsLMDM CSFDTVARTS	
	<i>A. fumigatus</i> 26906	GDEVAANFTA lFAPDIRARA KkHLPGVTLT
55	DEDVVsLMDM CSFDTVARTS	
	<i>A. fumigatus</i> 32239	GDEVEANFTA lFAPAIRARI EkHLPGVqLT
	DDDVVsLMDM CSFDTVARTA	



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E. nidulans      ADEiEANFTA IMGPPiRKRL ENDLPGIKLT
NENViYLMDM CSFDTMARTA
T. thermophilus  GHDAQEKFAK qFAPAiLEKI KDHLPGVDLA
vSDVpyLMDL CPFETLARNh
5 M. thermophila  GDDAQDTYiS TFAGPiTARV NANLPGANLT
DADTVaLMDL CPFETVAsSS

Consensus        GDDAEANFTA TFAPAiRARL EADLPGVTiLT DEDVV-
LMDM CPFETVARTS
10 Consensus phytase  GDDVEANFTA LFAPAiRARL EADLPGVTiLT
DEDVVYLMDM CPFETVARTS

251
15 300
A. terreus 9A-1  ..... DAhTLSPFC DLFTAtEWtq
YNYLlSLDKY YGYGGGNPLG
A. terreus cbs  ..... DAhTLSPFC DLFTAaEWtq
YNYLlSLDKY YGYGGGNPLG
20 A. niger var. awamori ..... vDTKLSPFC DLFTHdEWih
YDYLQSLkKY YGHGAGNPLG
A. niger T213   ..... vDTKLSPFC DLFTHdEWih
YDYLRSLkKY YGHGAGNPLG
A. niger NRRL3135 ..... vDTKLSPFC DLFTHdEWin
YDYLQSLkKY YGHGAGNPLG
25 A. fumigatus 13073 ..... DASQLSPFC QLFTHnEWkk
YNYLQSLGKY YGYGAGNPLG
A. fumigatus 32722 ..... DASQLSPFC QLFTHnEWkk
YNYLQSLGKY YGYGAGNPLG
30 A. fumigatus 58128 ..... DASQLSPFC QLFTHnEWkk
YNYLQSLGKY YGYGAGNPLG
A. fumigatus 26906 ..... DASQLSPFC QLFTHnEWkk
YNYLQSLGKY YGYGAGNPLG
A. fumigatus 32239 ..... DASELSPFC AIFTHnEWkk
35 YDYLQSLGKY YGYGAGNPLG
E. nidulans     ..... HGTELSPFC AIFTEkEWlq
YDYLQSLSKY YGYGAGSPLG
T. thermophilus ..... TDT.LSPFC ALStQeEWqa
YDYYQSLGKY YGnGGGNPLG
40 M. thermophila  sdpatadagg gNGrPLSPFC rLFSEsEWra
YDYLQSVGKW YGYGPGNPLG

Consensus        ----- -DATELSPFC ALFTE-EW--
YDYLQSLGKY YGYGAGNPLG
45 Consensus phytase ..... .DATELSPFC ALFTHDEWRQ
YDYLQSLGKY YGYGAGNPLG

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	350		301
	<i>A. terreus</i> 9A-1	PVQGVGWaNE	LMARLTRAPV HDHTCVNNTL
	DASPATFPLN ATLYADFSHD		
5	<i>A. terreus</i> cbs	PVQGVGWaNE	LIARLTRSPV HDHTCVNNTL
	DANPATFPLN ATLYADFSHD		
	<i>A. niger</i> var. <i>awamori</i>	PTQGVGYaNE	LIARLTHSPV HDDTSSNHTL
	DSNPATFPLN STLYADFSHD		
	<i>A. niger</i> T213	PTQGVGYaNE	LIARLTHSPV HDDTSSNHTL
10	DSNPATFPLN STLYADFSHD		
	<i>A. niger</i> NRRL3135	PTQGVGYaNE	LIARLTHSPV HDDTSSNHTL
	DSSPATFPLN STLYADFSHD		
	<i>A. fumigatus</i> 13073	PAQGIGFtNE	LIARLTRSPV QDHTSTNStL
	vSNPATFPLN ATMYVDFSHD		
15	<i>A. fumigatus</i> 32722	PAQGIGFtNE	LIARLTRSPV QDHTSTNStL
	vSNPATFPLN ATMYVDFSHD		
	<i>A. fumigatus</i> 58128	PAQGIGFtNE	LIARLTRSPV QDHTSTNStL
	vSNPATFPLN ATMYVDFSHD		
	<i>A. fumigatus</i> 26906	PAQGIGFtNE	LIARLTRSPV QDHTSTNStL
20	vSNPATFPLN ATMYVDFSHD		
	<i>A. fumigatus</i> 32239	PAQGIGFtNE	LIARLTNSPV QDHTSTNStL
	DSDPATFPLN ATTYVDFSHD		
	<i>E. nidulans</i>	PAQGIGFtNE	LIARLTQSPV QDNTSTNHTL
	DSNPATFPLD rKLYADFSHD		
25	<i>T. thermophilus</i>	PAQGVGFvNE	LIARMTSPV QDYTTVNHTL
	DSNPATFPLN ATLYADFSHD		
	<i>M. thermophila</i>	PTQGVGFvNE	LLARLAgvPV RDgTSTNRTL
	DGDPrTFPLG rPLYADFSHD		
30	Consensus	PAQGVGF-NE	LIARLTHSPV QDHTSTNHTL
	DSNPATFPLN ATLYADFSHD		
	Consensus phytase	PAQGVGFANE	LIARLTRSPV QDHTSTNHTL
	DSNPATFPLN ATLYADFSHD		
35			
	400		351
	<i>A. terreus</i> 9A-1	SNLVSIFWAL	GLYNGTAPLS qTSVESVSQT
	DGYAAAWTVP FAARAYVEMM		
40	<i>A. terreus</i> cbs	SNLVSIFWAL	GLYNGTkPLS qTTVEDITrT
	DGYAAAWTVP FAARAYIEMM		
	<i>A. niger</i> var. <i>awamori</i>	NGIISILFAL	GLYNGTkPLS TTTVENITQT
	DGFSSAWTVP FASRLYVEMM		
	<i>A. niger</i> T213	NGIISILFAL	GLYNGTkPLS TTTVENITQT
45	DGFSSAWTVP FASRLYVEMM		
	<i>A. niger</i> NRRL3135	NGIISILFAL	GLYNGTkPLS TTTVENITQT
	DGFSSAWTVP FASRLYVEMM		
	<i>A. fumigatus</i> 13073	NSMVSIFFAL	GLYNGTEPLS rTSVESaKEl
	DGYSASWVVP FGARAYFetM		
50	<i>A. fumigatus</i> 32722	NSMVSIFFAL	GLYNGTGPLS rTSVESaKEl
	DGYSASWVVP FGARAYFetM		
	<i>A. fumigatus</i> 58128	NSMVSIFFAL	GLYNGTEPLS rTSVESaKEl
	DGYSASWVVP FGARAYFetM		
	<i>A. fumigatus</i> 26906	NSMVSIFFAL	GLYNGTEPLS rTSVESaKEl
55	DGYSASWVVP FGARAYFetM		
	<i>A. fumigatus</i> 32239	NGMIPIFFAM	GLYNGTEPLS qTSeESTKES
	NGYSASWAVP FGARAYFetM		

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*E. nidulans* NSMISIFFAM GLYNGTQPLS mDSVESIQEm  
 DGYAASWTVP FGARAYFELM  
*T. thermophilus* NTMTSIFaAL GLYNGTAKLS TTEIKSIEET  
 DGYSAAWTVP FGGRAYIEMM  
 5 *M. thermophila* NDMMGVLgAL GaYDGVPPLD KTArrDpEEI  
 GGYAASWAVP FAARIYVEKM  
  
 Consensus NSMISIFFAL GLYNGTAPLS TTSVESIEET  
 DGYAASWTVP FGARAYVEMM  
 10 Consensus phytase NSMISIFFAL GLYNGTAPLS TTSVESIEET  
 DGYSASWTVP FGARAYVEMM  
  
 401  
 15 450  
*A. terreus* 9A-1 QC..... RAEKE PLVRVLVNDR  
 VMPLHGCPD KLGRCKrDAF  
*A. terreus* cbs QC..... RAEKQ PLVRVLVNDR  
 VMPLHGCAVD NLGRCKrDDF  
 20 *A. niger* var. *awamori* QC..... QAEQE PLVRVLVNDR  
 VVPLHGCPID aLGRCTrDSF  
*A. niger* T213 QC..... QAEQE PLVRVLVNDR  
 VVPLHGCPID aLGRCTrDSF  
*A. niger* NRRL3135 QC..... QAEQE PLVRVLVNDR  
 25 VVPLHGCPVD aLGRCTrDSF  
*A. fumigatus* 13073 QC..... KSEKE PLVRALINDR  
 VVPLHGCDVD KLGRCKLNDF  
*A. fumigatus* 32722 QC..... KSEKE PLVRALINDR  
 VVPLHGCDVD KLGRCKLNDF  
 30 *A. fumigatus* 58128 QC..... KSEKE SLVRALINDR  
 VVPLHGCDVD KLGRCKLNDF  
*A. fumigatus* 26906 QC..... KSEKE PLVRALINDR  
 VVPLHGCDVD KLGRCKLNDF  
*A. fumigatus* 32239 QC..... KSEKE PLVRALINDR  
 35 VVPLHGCAVD KLGRCKLNDF  
*E. nidulans* QC..... E.KKE PLVRVLVNDR  
 VVPLHGCAVD KFGRCTLDLDDW  
*T. thermophilus* QC..... DDSDE PVVRVLVNDR  
 VVPLHGCEVD SLGRCKrDDF  
 40 *M. thermophila* RCsggggggg gggrQEKE eMVRVLVNDR  
 VMTLkGCGAD ErGMCTLErF  
  
 Consensus QC----- QAEKE PLVRVLVNDR  
 VVPLHGCAVD KLGRCKLDDF  
 45 Consensus phytase QC..... QAEKE PLVRVLVNDR  
 VVPLHGCAVD KLGRCKRDDF  
  
 451  
 471  
 50 *A. terreus* 9A-1 VAGLSFAQAG GNWADCF---  
*A. terreus* cbs VEGLSFARAG  
 GNWAECF---  
*A. niger* var. *awamori* VrGLSFARSG GDWAECsA--  
*A. niger* T213 VrGLSFARSG GDWAECFA--  
 55 *A. niger* NRRL3135 VrGLSFARSG  
 GDWAECFA--  
*A. fumigatus* 13073 VKGLSWARSG GNWGECS--  
*A. fumigatus* 32722 VKGLSWARSG GNWGECS--

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	A. fumigatus 58128	VKGLSWARSG GNWGECS-- -
	A. fumigatus 26906	VKGLSWARSG GNWGECS-- -
	A. fumigatus 32239	VKGLSWARSG
	GNSEQSFS-- -	
5	E. nidulans	VEGLNFARSG GNWkTCFTl~ -
	T. thermophilus	VrGLSFARqG GNWEGCYAas e
	M. thermophila	IESMAFARGN GKWDlCFA-- -
	Consensus	VEGLSFARSG GNWAECS-- -
10	Consensus phytase	VEGLSFARSG GNWAECS... .

Figure 2

CP-1

15 Eco RI M G V F V V L L S I A T L F G S T  
TATATGAATTCTATGGCGTGTTCGTCGTGCTACTGTCCATTGCCACCTTGTTCCGGTTCCA

1 -----+-----+-----+-----+-----+ 60

ATATACTTAAGTACCCGCACAAGCAGCAGCATGACAGGTAACGGTGGAACAAGCCAAGGT

20 S G T A L G P R G N S H S C D T V D G G  
CATCCGGTACCGCCTTGGGTCCTCGTGGTAATTCTCACTCTTGACACTGTTGACGGTG

61 -----+-----+-----+-----+-----+ 120

GTAGGCCATGGCGGAACCCAGGAGCACCATTAAAGAGTGAGAACACTGTGACAACTGCCAC

CP-2

CP-3

25 Y Q C F P E I S H L W G Q Y S P Y F S L  
GTTACCAATGTTTCCAGAAATTTCTCACTTGTGGGGTCAATACTCTCCATACTTCTCTT

121 -----+-----+-----+-----+-----+ 180

CAATGGTTACAAAGGGTCTTTAAAGAGTGAACACCCAGTTATGAGAGGTATGAAGAGAA

30 E D E S A I S P D V P D D C R V T F V Q  
TGGAAGACGAATCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTCGTTT

181 -----+-----+-----+-----+-----+ 240

ACCTTCTGCTTAGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAG

CP-4

CP-5

35 V L S R H G A R Y P T S S K S K A Y S A  
AAGTTTGTCTAGACACGGTGCTAGATACCCAACTTCTTCTAAGTCTAAGGCTTACTCTG

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241 -----+-----+-----+-----+-----+-----+ 300

TTCAAAACAGATCTGTGCCACGATCTATGGGTTGAAGAAGATTGAGATTCCGAATGAGAC

L I E A I Q K N A T A F K G K Y A F L K

5 CTTTGATTGAAGCTATTCAAAGAACGCTACTGCTTCAAGGGTAAGTACGCTTTCTTGA

301 -----+-----+-----+-----+-----+-----+ 360

GAAACTAACTTCGATAAGTTTCTTGCGATGACGAAAGTTCCCATTTCATGCGAAAGAACT

CP-6

CP-7

10 T Y N Y T L G A D D L T P F G E N Q M V

AGACTTACAACCTACACTTTGGGTGCTGACGACTTGACTCCATTGCGTGAAAACCAAATGG

361 -----+-----+-----+-----+-----+-----+ 420

TCTGAATGTTGATGTGAAAACCCACGACTGCTGAACTGAGGTAAGCCACTTTTGGTTTACC

15 N S G I K F Y R R Y K A L A R K I V P F

TTAACTCTGGTATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCAT

421 -----+-----+-----+-----+-----+-----+ 480

AATTGAGACCATAATTCAAGATGCTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTA

CP-8

20

CP-9

I R A S G S D R V I A S A E K F I E G F

TCATTAGAGCTTCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTTCATTGAAGGTT

481 -----+-----+-----+-----+-----+-----+ 540

AGTAATCTCGAAGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACTTCCAA

25

Q S A K L A D P G S Q P H Q A S P V I D

TCCAATCTGCTAAGTTGGCTGACCCAGGTTCTCAACCACACCAAGCTTCTCCAGTTATTG

541 -----+-----+-----+-----+-----+-----+ 600

AGGTTAGACGATTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTTGAAGAGGTCAATAAC

30

CP-10

CP-11

V I I P E G S G Y N N T L D H G T C T A

ACGTTATTATTCCAGAAGGATCAGGTTACAACACACTTTGGACCACGGTACTTGTACTG

601 -----+-----+-----+-----+-----+-----+ 660

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TGCAATAATAAGGTCTTCctAGgCCAATGTTGTTGTGAAACCTGGTGCCATGAACATGAC

F E D S E L G D D V E A N F T A L F A P

CTTTCGAAGACTCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTTTGTTGCTC

5 661 -----+-----+-----+-----+-----+-----+ 720

GAAAGCTTCTGAGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGAAACAAGCGAG

CP-12

A I R A R L E A D L P G V T L T D E D V

10 CAGCTATTAGAGCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGACGAAGACG

721 -----+-----+-----+-----+-----+-----+ 780

GTCGATAATCTCGATCTAACCTTCGACTGAACGGTCCACAATGAACTGACTGCTTCTGC

CP-13

15 V Y L M D M C P F E T V A R T S D A T E

TTGTTTACTTGATGGACATGTGTCCATTGAACTGTTGCTAGAACTTCTGACGCTACTG

781 -----+-----+-----+-----+-----+-----+ 840

AACAAATGAACTACCTGTACACAGGTAAGCTTTGACAACGATCTTGAAGACTGCGATGAC

20 L S P F C A L F T H D E W R Q Y D Y L Q

AATTGTCTCCATTCTGTGCTTTGTTCACTCACGACGAATGGAGACAATACGACTACTTGC

841 -----+-----+-----+-----+-----+-----+ 900

TTAACAGAGGTAAGACACGAAACAAGTGAGTGCTGCTTACCTCTGTTATGCTGATGAACG

CP-14

25

CP-15

S L G K Y Y G Y G A G N P L G P A Q G V

AATCTTTGGGTAAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTG

901 -----+-----+-----+-----+-----+-----+ 960

TTAGAAACCCATTTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCAC

30

G F A N E L I A R L T R S P V Q D H T S

TTGGTTTCGCTAACGAATTGATTGCTAGATTGACTAGATCTCCAGTTCAAGACCACACTT

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961 -----+-----+-----+-----+-----+-----+  
1020  
AACCAAAGCGATTGCTTAACTAACGATCTAACTGATCTAGAGGTCAAGTTCTGGTGTGAA  
CP-16  
5 CP-17  
T N H T L D S N P A T F P L N A T L Y A  
CTACTAACCACACTTTGGACTCTAACCAGCTACTTTCCCATTTGAACGCTACTTTGTACG  
1021 -----+-----+-----+-----+-----+-----+  
1080  
10 GATGATTGGTGTGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCGATGAAACATGC  
D F S H D N S M I S I F F A L G L Y N G  
CTGACTTCTCTCACGACAACTCTATGATTCTATTTTCTTCGCTTTGGGTTTGTACAACG  
1081 -----+-----+-----+-----+-----+-----+  
15 1140  
GACTGAAGAGAGTGCTGTTGAGATACTAAAGATAAAAGAAGCGAAACCCAAACATGTTGC  
CP-18  
CP-19  
T A P L S T T S V E S I E E T D G Y S A  
20 GTACTGCTCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTG  
1141 -----+-----+-----+-----+-----+-----+  
1200  
CATGACGAGGTAACAGATGATGAAGACAACTTAGATAACTTCTTTGACTGCCAATGAGAC  
25 S W T V P F G A R A Y V E M M Q C Q A E  
CTTCTTGGACTGTTCCATTGCGTGCTAGAGCTTACGTTGAAATGATGCAATGTCAAGCTG  
1201 -----+-----+-----+-----+-----+-----+  
1260  
GAAGAACCTGACAAGGTAAGCCACGATCTCGAATGCAACTTTACTACGTTACAGTTCGAC  
30 CP-20  
CP-21  
K E P L V R V L V N D R V V P L H G C A  
AAAAGGAACCATTGGTTAGAGTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG  
1261 -----+-----+-----+-----+-----+-----+  
35 1320

V D K L G R C K R D D F V E G L S F A R

5 1321 1380

CP-22

S G G N W A E C F A \* Eco RI

1381 -----+-----+-----+-----+----- 1426

CTAGACCACCATTTGACCCGACTTACAAAGCGAATTCTTAAGTATAT

15

**Figure 3**

1

50

20 P. involutus (phyA1) SvP.KnTAPt FPIPeseQrn WSPYSPYFPL AeYkAPPAGC  
QInQVNIIQR

*P. involutus* (phyA2) SvP.RniAPK FSIPeseQrn WSPYSPYFPL AeYkAPPAGC  
EInQVNIIQR

*T. pubescens* hiPlRdTSAc LdVTrDvQqs WSmYSPYFPa' AtYvAPPASC  
QInQVHIQR

25    *A. pediades*                    GgvvQaTfvQ pfFPPQiQds WAAYTPYYPV qaYtPPPkDC  
         KitQVNIIQR

*P. lycii* StQfsfvAAQ LPiPaQntsn WGPYdPFFPV EpYaAPPEGC  
tVtQVNLIQR

30 Basidio S-P-R-TAAQ LPIP-Q-Q-- WSPYSPYFPV A-Y-APPAGC QI-  
QVNIIR



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100

*P. involutus* (phyA1) HGARFPTSGA TTRIKAGLTK LQGVqnfTDA KFNFIksfky  
dLGnsDLVPP

5 *P. involutus* (phyA2) HGARFPTSGA ATRIKAGLSK LQSVqnfTDP KFDFIksfTY  
dLGtsDLVPP

*T. pubescens* HGARFPTSGA AKRIQTAVAK LKAAsnyTDP lLAFVtNyTY  
sLGqDsLveL

10 *A. pediades* HGARFPTSGA GTRIQAaVKK LQSAktyTDP RLDFLtnTY  
tLGhDDLVPF

*P. lycii* HGARWPTSGA rSRqvAAVAK IQmArpfTDP KYEFLnDfvY  
kFGvADLLPF

15 **Basidio** HGARFPTSGA ATRIQAaVAK LQSA---TDP KLDfL-N-TY -LG-  
**DDLVPF**

101

150

20 *P. involutus* (phyA1) GAaQSfDAGQ EAFARYSkLV SkNNLPFIRA dGSDRVVDSA  
TNWTAGFAsA

*P. involutus* (phyA2) GAaQSfDAGl EvFARYSkLV SsDNLPPFIRS dGSDRVVDTA  
TNWTAGFAsA

25 *T. pubescens* GAtQSSEAGQ EAFTRYsLV SaDELpFVRA SGSDRVVATA  
nNWTAGFAlA

*A. pediades* GAlQSSQAGE ETfQRYsfLV SkENLPFVRA SSSNRVVDSA  
TNWTEGFSaA

*P. lycii* GAnQShQTGt DmYTRYStLf egGDVPFVRA AGdQRVVDSS  
TNWTAGFGdA

30 **Basidio** GA-QSSQAGQ EAFTRYs-LV S-DNLpFVRA SGSDRVVDSA  
**TNWTAGFA-A**

35

151

200

*P. involutus* (phyA1) ShNTvqPkLn LILPQtGNDT LEDNMCPaAG DSDPQvNaWL  
AVafPSITAR

40 *P. involutus* (phyA2) SrNAiqPkLd LILPQtGNDT LEDNMCPaAG ESDPQvDaWL  
AsafPSVTAQ

	<i>T. pubescens</i> AqFAPPMTAR	SsNSitPvLs VIISEaGNDT LDDNMCPaAG DSDPQvNqWL
	<i>A. pediades</i> SIYGTPIAnR	ShHvlnPiLf VILSEsINDT LDDaMCPnAG sSDPQtGiWt
5	<i>P. lycii</i> GVFAPnITAR	SgETv1PtLq VVLqEeGNcT LcNNMCPnEv DGDest.tWL
	<b>Basidio</b> AVFAPPITAR	<b>S-NT--P-L- VILSE-GNDT LDDNMCP-AG DSDPQ-N-WL</b>
10		
		201
	250	
15	<i>P. involutus</i> (phyA1) giPGsFeAFa	LNAAAPSVNL TDtDAfNLvs LCAFlTVSke kkSdFctLFE
	<i>P. involutus</i> (phyA2) giPGsFeAFa	LNAAAPGANL TDaDAfNLvs LCPFmTVSke qkSdFctLFE
	<i>T. pubescens</i> elQAE.dAFa	LNAGAPGANL TDtDTyNLlt LCPFETVatE rrSeFCDIYE
20	<i>A. pediades</i> .tPEEFaqFe	LNqqAPGANI TAAdvsNLip LCAFETivke tpSpFCNLF.
	<i>P. lycii</i> .tABEYvSYe	LNAAAPSANL SDsDAItLmd MCPFDTLSSg naSpFCDLF.
25	<b>Basidio</b> AF-	<b>LNAAAPGANL TD-DA-NL-- LCPFETVS-E --S-FCDLFE --PEEF-</b>
		251
30	300	
	<i>P. involutus</i> (phyA1) NTQTNRTLDA	YgGDLdkFYG TGYGQeLGPV QGVGYVNELI ARLTnsAVRD
	<i>P. involutus</i> (phyA2) NTQTNRTLDA	YaGDLdkFYG TGYGQALGPV QGVGYINELL ARLTnsAVnD
35	<i>T. pubescens</i> HTQTNsTLDS	YnADLDKfYg TGYGQPLGPV QGVGYINELI ARLTaQnVsD
	<i>A. pediades</i> NTQTNRTLDS	YfGDLDKfYg TGYGQPLGPV QGVGYINELL ARLTemPVrd
40	<i>P. lycii</i> ETQTNRTLDS	YyyDLdkYYG TGpGNALGPV QGVGYVNELL ARLTgQAVRD

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Basidio Y-GDLDFYFG TGYGQPLGPV QGVGYINELL ARLT-QAVRD  
 NTQTNRTLDS

5

301

350

*P. involutus* (phyA1) SPvTFPLNKT FYADFSHDN1 MVAVFSAMGL FrQPAPLsTS  
 vPNPwRTwRT

10 *P. involutus* (phyA2) APdTFPLNKT MYADFSHDN1 MVAVFSAMGL FrQSAPLsTS  
 tPDPNRTWLT

*T. pubescens* SPeTFPLNRT LYADFSHDNQ MVAIFSAMGL FNQSAPLDPT  
 tPDPaRTFLv

15 *A. pediades* SPtTFPLDRS IYADLSHDNQ MIAIFSAMGL FNQSSPLDPS  
 fPNPKRTWVT

*P. lycii* dPaTFPLNRT FYADFSHDNt MVPIFAALGL FNaTA.LDPl  
 kPDeNRLwVd

20 Basidio SP-TFPLNRT FYADFSHDNQ MVAIFSAMGL FNQSAPLDPS -  
 PDPNRTWVT

351

400

25 *P. involutus* (phyA1) SsLVPFSGRM VVERLsC..f GT..... tkV  
 RVLVQDqVQP

*P. involutus* (phyA2) SsVVPFSARM aVERLsC..a GT..... tkV  
 RVLVQDqVQP

30 *T. pubescens* kKIVPFSARM VVERLdC..g GA..... qsv  
 RLLVNDaVQP

*A. pediades* SRLtPFSARM VtERLlCqrd GTgsgggsri mrngnvqtfv  
 RILVNDALQP

*P. lycii* SKLVPFSGHM tVEKLaC... sgkeaV  
 RVLVNDaVQP

35 Basidio SKLVPFSGRM VVERL-C--- GT-----V  
 RVLVNDaVQP

40 401 441

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*P. involutus* (phyA1) LEFCGGDrNG lCTLAKFVES QtFARsDGaG DFEKCFATSa ~  
*P. involutus* (phyA2) LEFCGGDqDG lCALDkFVES QaYARsGGaG DFEKCLATTv ~  
*T. pubescens* LAFCGADtsG vCTLDaFVES QaYARNDGEG DFEKCFAT-- ~  
*A. pediades* LKFCGGDmDS lCTLEAFVES QkYAREDGQG DFEKCFD--- ~  
5 *P. lycii* LEFCGG.vDG vCeLsAFVES QtYARENGQG DfAKCgfvPs e  
  
*Basidio* LEFCGGD-DG -CTLDaFVES Q-YAREDGQG DFEKCFATP- -

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Figure 4

		1	
50			
5	<i>A. terreus</i> 9a1 VPeDCHITFV	KhsdCNSVDh GYQCfPELSH kWGLYAPYFS LqDESPFP1D	
	<i>A. terreus</i> cbs VPdDCHITFV	NhsdCtSVDr GYQCfPELSH kWGLYAPYFS LqDESPFP1D	
10	<i>A. niger</i> var. <i>awamori</i> VPaGCRVTFa	NqsTCDTVdG GYQCfSEtSH LWGQYAPFFS LANESAISPD	
	<i>A. niger</i> NRRL3135 VPaGCRVTFa	NqsSCDTVdG GYQCfSEtSH LWGQYAPFFS LANESvISPE	
	<i>A. fumigatus</i> 13073 LPkDCRITLV	GskSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDELSVSSK	
15	<i>A. fumigatus</i> 32722 LPkDCRITLV	GskSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDELSVSSK	
	<i>A. fumigatus</i> 58128 LPkDCRITLV	GskSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDELSVSSK	
20	<i>A. fumigatus</i> 26906 LPkDCRITLV	GskSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDELSVSSK	
	<i>A. fumigatus</i> 32239 LPkDCRVTFV	GskACDTVE1 GYQCSPGtSH LWGQYSPFFS LEDELSVSSD	
	<i>E. nidulans</i> VPhGCeVTFV	QNHSCNTaDG GYQCfPNVSH VWGQYSPYFS IEQESAISeD	
25	<i>T. thermophilus</i> VPqNCKITFV	DSHSCNTVEG GYQCrPEISH sWGQYSPFFS LADQSEISPD	
	<i>T. lanuginosa</i> VPkGCRVeFV	----- ----nvDIAR hWGQYSPFFS LAEvSEISPA	
30	<i>M. thermophila</i> IPdDCeVTFa	ESRPCDTpD1 GFQCgTAISH FWGQYSPYFS VPSElDaS..	
	Basidio pPaGCQIxqV	xSxPxrxTAA qLPipxQxqx xWSPYSPYFP VAXyxA....	
35	Consensus GCRVTFV	NSHSCDTVdG GYQC-PEISH LWGQYSPFFS LADESAISPD VP-	
	Fcp10 VPKGCRVTFV	NSHSCDTVdG GYQCfPEISH LWGQYSPFFS LADESAISPD	

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		51	
100			
5	<i>A. terreus</i> 9a1 QSYNYSLDSE	QVLARHGARS PThSKTKaYA AtIaAIQKSA TaFpGKYAFL	
	<i>A. terreus</i> cbs KSYNYSMGSE	QVLARHGARS PTdSKTKaYA AtIaAIQKNA TaLpGKYAFL	
	<i>A. niger</i> var. <i>awamori</i> KTYNYSLGAD	QVLSRHGARY PTeSKGKKYS ALIeEIQQNv TtFDGKYAFL	
10	<i>A. niger</i> NRRL3135 KTYNYSLGAD	QVLSRHGARY PTdSKGKKYS ALIeEIQQNA TtFDGKYAFL	
	<i>A. fumigatus</i> 13073 KTYNYTLGAD	QVLSRHGARY PTSSKSCKYk kLVtAIQaNA TdFKGKFAFL	
15	<i>A. fumigatus</i> 32722 KTYNYTLGAD	QVLSRHGARY PTSSKSCKYk kLVtAIQaNA TdFKGKFAFL	
	<i>A. fumigatus</i> 58128 KTYNYTLGAD	QVLSRHGARY PTSSKSCKYk kLVtAIQaNA TdFKGKFAFL	
	<i>A. fumigatus</i> 26906 KTYNYTLGAD	QVLSRHGARY PTSSKSCKYk kLVtAIQaNA TdFKGKFAFL	
20	<i>A. fumigatus</i> 32239 ETNYNYTLGAD	QVLSRHGARY PTASKSCKYk kLVtAIQKNA TeFKGKFAFL	
	<i>E. nidulans</i> ESYNYTLGAD	QVLSRHGARY PTeSKSKaYS GLIeAIQKNA TsFwGQYAFL	
25	<i>T. thermophilus</i> KdYrYqLGAN	QLLSRHGARY PTSSKTELYS qLIsrIQKtA TaYKGyYAFL	
	<i>T. lanuginosa</i> RdYaYhLGAD	QVLSRHGARY PTAhKSEvYA ELLqrIQDtA TeFKGDFAFL	
	<i>M. thermophila</i> RTYDYTLGAD	QVLSRHGARA PTLkRAasYv DLIdrIHhGA isYgPgYEFL	
30	Basidio xnxtYxLGxD	NIIqRHGARF PTSGaAtRiq AaVakLQsax xxtDPKLDLFL	
	Consensus KTYNYTLGAD	QVLSRHGARY PTSSKSCKYS ALI-AIQKNA T-FKGKYAFL	
35	Fcp10 KTYNYTLGAD	QVLSRHGARY PTSSKSCKYS ALIEAIQKNA TAFKGKYAFL	

		101	
5	150		
	<i>A. terreus</i> 9a1	ELTPFGGrNQL rDlGaQFYeR YNAL.TRhIn PFVRATDAsR	
	VhESA EK FVE		
	<i>A. terreus</i> cbs	NLTPFGGrNQL qDlGaQFYRR YDTL.TRhIn PFVRAADSsR	
	VhESA EK FVE		
10	<i>A. niger</i> var. <i>awamori</i>	DLTPFGGEQEL VNSGIKFYQR YESL.TRnII PFIRSSGSsR	
	VIASGEKFIE		
	<i>A. niger</i> NRRL3135	DLTPFGGEQEL VNSGIKFYQR YESL.TRnIV PFIRSSGSsR	
	VIASGKKFIE		
15	<i>A. fumigatus</i> 13073	DLTPFGGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR	
	VIASGEKFIE		
	<i>A. fumigatus</i> 32722	DLTPFGGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR	
	VIASGEKFIE		
	<i>A. fumigatus</i> 58128	DLTPFGGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR	
	VIASGEKFIE		
20	<i>A. fumigatus</i> 26906	DLTAFGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR	
	VIASGEKFIE		
	<i>A. fumigatus</i> 32239	DLTPFGGEQQM VNSGIKFYQK YKAL.AgsVV PFIRSSGSDR	
	VIASGEKFIE		
25	<i>E. nidulans</i>	DLTiFGENQM VDSGaKFYRR YKnL.ARknt PFIRASGSDR	
	VVASAEKFIN		
	<i>T. thermophilus</i>	DLTPFGENQM IQlGIKFYnH YKSL.ARnaV PFVRCSGSDR	
	VIASGrIFIE		
	<i>T. lanuginosa</i>	NLTRFGEEQM MESGrQFYHR YREq.AReIV PFVRAAGSAR	
	VIASAEfFnr		
30	<i>M. thermophila</i>	ELTRtGQQQM VNSGIKFYRR YRAL.ARksI PFVRTAGqDR	
	VVhSAENftQ		
	Basidio	DLvPFGAXqs sQAGqEaFtR YsxLvSxdnL PFVRASGSDR	
	VVDSAtNWtA		
35	Consensus	DLTPFGGEQQM VNSGIKFYRR YKAL-AR-IV PFVRASGSDR	
	VIASAEKFIE		
	Fcp10	DLTPFGGEQQM VNSGIKFYRR YKAL.ARkIV PFVRASGSDR	
	VIASAEKFIE		

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		151	
200			
5	<i>A. terreus</i> 9a1 TAFES...St	GFQTARqDDh hAnphQPSPr VDValPEGSa YNNTLEHSLC	
	<i>A. terreus</i> cbs TAFEa...St	GFQNARqGDP hAnphQPSPr VDVVIPEGtA YNNTLEHSIC	
	<i>A. niger</i> var. <i>awamori</i> TvFEd...SE	GFQSTKLkDP rAqpgQSSPk IDVVISEAs sNNTLDpGtC	
10	<i>A. niger</i> NRRL3135 TvFEd...SE	GFQSTKLkDP rAqpgQSSPk IDVVISEAs sNNTLDpGtC	
	<i>A. fumigatus</i> 13073 TkFEa...SQ	GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC	
15	<i>A. fumigatus</i> 32722 TkFEa...SQ	GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC	
	<i>A. fumigatus</i> 58128 TkFEa...SQ	GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC	
	<i>A. fumigatus</i> 26906 TkFEa...SQ	GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC	
20	<i>A. fumigatus</i> 32239 TnFEa...SE	GFQqANVADP gAt.nRAAPV ISVIIPESeT YNNTLDHSVC	
	<i>E. nidulans</i> vSFEn...dE	GFRKAQLhDh g.s.gQATPV VNVIPEidG FNNTLDHStC	
25	<i>T. thermophilus</i> PvFEd...Ss	GFQSAKVlDP hSdKhDAPPt INVIIeEGPs YNNTLDtGsC	
	<i>T. lanuginosa</i> PAaEe...Ap	GFQdAKdrDP rSnkdQAePV INVIISEETG sNNTLDgltC	
	<i>M. thermophila</i> TAFEegPySt	GFHSALLADR gStvrPTlPy dmVVIPETaG aNNTLHNDLC	
30	Basidio .....PxAG	GFaxA.....sxntxxPx LxVILSExg. .NDTLDDNMC	
	Consensus	GFQSAKLADP -A---QASPV INVIIPEG-G YNNTLDHGLC	
	TAFE--P-SE		
35	Fcp10	GFQSAKLADP GANPHQASPV INVIIPEGAG YNNTLDHGLC	
	TAFEE...SE		

201

250



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	<i>A. terreus</i> 9a1	VGDDavANFT AVFAPAIaQr LEAdLPGVQL StDDVVNLMA
	MCPFETVSlT	
	<i>A. terreus</i> cbs	VGDAaADNFT AVFAPAIaK R LEAdLPGVQL SADDVVNLMA
	MCPFETVSlT	
5	<i>A. niger</i> var. <i>awamori</i>	LADtVEANFT AtFAPSIRqR LEndLSGVtL TDtEVtyLMD
	MCSFDTISs	
	<i>A. niger</i> NRRL3135	LADtVEANFT AtFvPSIRqR LEndLSGVtL TDtEVtyLMD
	MCSFDTISs	
10	<i>A. fumigatus</i> 13073	LGDEVAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD
	MCSFDTVArT	
	<i>A. fumigatus</i> 32722	LGDEVAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD
	MCSFDTVArT	
	<i>A. fumigatus</i> 58128	LGDEVAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD
	MCSFDTVArT	
15	<i>A. fumigatus</i> 26906	LGDEVAANFT ALFAPdIRAR aKkhLPGVtL TDEDVVSLMD
	MCSFDTVArT	
	<i>A. fumigatus</i> 32239	LGDEVEANFT ALFAPAIRAR IEkhLPGVQL TDDDVVSLMD
	MCSFDTVArT	
20	<i>E. nidulans</i>	rADEIEANFT AIMGPPIRkR LEndLPGIKL TNENViyLMD
	MCSFDTMArT	
	<i>T. thermophilus</i>	gGHDAQEKFA kqFAPAIIEK IKDhLPGVDL AvsDVpyLMD
	LCPFETLArN	
	<i>T. lanuginosa</i>	.DptqpAEfI qVFGPRVlkK ItkhMPGVNL TLEDVplfMD
	LCPFDTVGSd	
25	<i>M. thermophila</i>	IGDDaQDtYl StFAGPItAR VNAnLPGaNL TDADtVaLMD
	LCPFETVAss	
	Basidio	dSDpqxnXWl AVFAPPItAR LNAaaPGaNL TDxDaxNLxx
	LCPFETVS..	
30	Consensus	LGDDVEANFT AVFAPPIRAR LEA-LPGVNL TDEDVVNLMD
	MCPFDTVA-T	
	Fcp10	LGDDVEANFT AVFAPPIRAR LEAHLPGVNL TDEDVVNLMD
	MCPFDTVART	
35	251	
	300	
	<i>A. terreus</i> 9a1	dD..Aht... ..LSPF CDLFTa..tE WtQYNYLlSL
	dKYYGYGGGN	

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	<i>A. terreus</i> cbs	dD..Aht... ..LSPF CDLFTa..aE WtQYNYLlSL
	dKYYGYGGGN	
	<i>A. niger</i> var. <i>awamori</i> Tv..DTK... ..LSPF CDLFTH..dE WiHYDYLQSL	
	kKYYGHGAGN	
5	<i>A. niger</i> NRRL3135	Tv..DTK... ..LSPF CDLFTH..dE WiNYDYLQSL
	kKYYGHGAGN	
	<i>A. fumigatus</i> 13073	SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL
	gKYYGYGAGN	
10	<i>A. fumigatus</i> 32722	SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL
	gKYYGYGAGN	
	<i>A. fumigatus</i> 58128	SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL
	gKYYGYGAGN	
	<i>A. fumigatus</i> 26906	SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL
	gKYYGYGAGN	
15	<i>A. fumigatus</i> 32239	AD..ASE... ..LSPF CAIFTH..nE WkKYDYLQSL
	gKYYGYGAGN	
	<i>E. nidulans</i>	AH..GTE... ..LSPF CAIFTE..kE WlQYDYLQSL
	sKYYGYGAGS	
20	<i>T. thermophilus</i>	ht..DT.... ..LSPF CALsTQ..eE WqaYDYYQSL
	gKYYGnGGGN	
	<i>T. lanuginosa</i>	PvlfPrQ... ..LSPF CHLFTa..dD WmaYDYYyTL
	dKYYSHGGGS	
	<i>M. thermophila</i>	SsdpATadag ggngrrpLSPF CrLFSE..sE WrayDYDLSV
	gKWYGYGPGN	
25	Basidio	.....xexxSxF CDLFexxpeE FxaFxYxgdL
	dKFYGTgYgQ	
	Consensus	SD--ATQ--- -----LSPF CDLFTH---E W-QYDYLQSL -
	KYYGYGAGN	
30	Fcp10	SD..ATQ... ..LSPF CDLFTH..DE WiQYDYLQSL
	GKYYGYGAGN	
		301
	350	
35	<i>A. terreus</i> 9a1	PLGPvQGVGW aNELMARLTR A.PVHDHTCv NNTLDASPAT
	FPLNATLYAD	
	<i>A. terreus</i> cbs	PLGPvQGVGW aNELIARLTR S.PVHDHTCv NNTLDANPAT
	FPLNATLYAD	

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	<i>A. niger</i> var. <i>awamori</i>	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSNPAT
	FPLNSTLYAD	
	<i>A. niger</i> NRRL3135	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSSPAT
	FPLNSTLYAD	
5	<i>A. fumigatus</i> 13073	PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT
	FPLNATMYvD	
	<i>A. fumigatus</i> 32722	PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT
	FPLNATMYvD	
10	<i>A. fumigatus</i> 58128	PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT
	FPLNATMYvD	
	<i>A. fumigatus</i> 26906	PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT
	FPLNATMYvD	
	<i>A. fumigatus</i> 32239	PLGPAQGIGF tNELIARLTN S.PVQDHTST NsTLDSDPAT
	FPLNATIYvD	
15	<i>E. nidulans</i>	PLGPAQGIGF tNELIARLTQ S.PVQDNTST NHTLDSNPAT
	FPLDrkLYAD	
	<i>T. thermophilus</i>	PLGPAQGVGF vNELIARMTH S.PVQDYTTv NHTLDSNPAT
	FPLNATLYAD	
20	<i>T. lanuginosa</i>	AFGPSRGVGF vNELIARMTg NlPVKDHTTv NHTLDdNPET
	FPLDAvLYAD	
	<i>M. thermophila</i>	PLGPTQGVGF vNELLARLA. GvPVRDgTST NRTLdGDPrt
	FPLGrPLYAD	
	Basidio	PLGPvQGVGY iNELLARLTx qa.VRDNTqT NRTLDSsPxT
	FPLNrTFYAD	
25	Consensus	PLGPAQGVGF -NELIARLTH S-PVQDHTST NHTLDSNPAT
	FPLNATLYAD	
	Fcp10	PLGPAQGVGF vNELIARLTH S.PVQDHTST NHTLDSNPAT
	FPLNATLYAD	
30		
		351
	400	
	<i>A. terreus</i> 9a1	FSHDSnLVSI FWALGLYNGT aPLSqTSVE. .SvsQTDGYA
	AAWTVPFAR	
35	<i>A. terreus</i> cbs	FSHDSnLVSI FWALGLYNGT kPLSqTTVE. .ditrTDGYA
	AAWTVPFAR	
	<i>A. niger</i> var. <i>awamori</i>	FSHDNGIISI LFALGLYNGT kPLSTTTVE. .NitQTDGFS
	SAWTVPFASR	

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	<i>A. niger</i> NRRL3135 SAWTVPFASR	FSHDNGIISI LFALGLYNGT kPLSTTTVE. .NitQTDGFS
	<i>A. fumigatus</i> 13073 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKElDGYS
5	<i>A. fumigatus</i> 32722 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT gPLSrTSVE. .SaKElDGYS
	<i>A. fumigatus</i> 58128 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKElDGYS
10	<i>A. fumigatus</i> 26906 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKElDGYS
	<i>A. fumigatus</i> 32239 ASWAVPFGAR	FSHDNGMIPI FFAMGLYNGT ePLSqTSeE. .StKESNGYS
	<i>E. nidulans</i> ASWTVPFGAR	FSHDNSMISI FFAMGLYNGT qPLSmdSVE. .SiQEmDGYA
15	<i>T. thermophilus</i> AAWTVPFGR	FSHDNTMtSI FaALGLYNGT akLSTTeIK. .SiEETDGYS
	<i>T. lanuginosa</i> ASWTVPFAAR	FSHDNTMtGI FsAMGLYNGT kPLSTSkIQP pTgAAADGYA
20	<i>M. thermophila</i> ASWAVPFAAR	FSHDNdMMGV LgALGaYDgV pPLdkTA..R rdpEELGGYA
	Basidio TSklVPFSAR	FSHDNqMVAI FsAMGLFNqS aPLdPSxpDP nrt.....Wv
25	Consensus ASWTVPFAAR	FSHDNTMVSI FFALGLYNGT -PLSTTSVEP -S-EETDGYS
	Fcp10 ASWTVPFAAR	FSHDNTMVSI FFALGLYNGT KPLSTTSVE. .SiEETDGYS
30	450	401
	<i>A. terreus</i> 9a1 PLHGCPtDKL	AYVEMMQC.. ra.....EKEPL VRVLVNDVRM
	<i>A. terreus</i> cbs PLHGCAVDNL	AYIEMMQC.. ra.....EKQPL VRVLVNDVRM
35	<i>A. niger</i> var. <i>awamori</i> PLHGCPIDaL	lyVEMMQC.. Qa.....EQEPL VRVLVNDRVV
	<i>A. niger</i> NRRL3135 PLHGCPVDaL	lyVEMMQC.. Qa.....EQEPL VRVLVNDRVV

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	<i>A. fumigatus</i> 13073 PLHGCDVDKL	AYfEtMQC.. Ks..... EKEPL VRaLINDRVV
	<i>A. fumigatus</i> 32722 PLHGCDVDKL	AYfEtMQC.. Ks..... EKEPL VRaLINDRVV
5	<i>A. fumigatus</i> 58128 PLHGCDVDKL	AYfEtMQC.. Ks..... EKESL VRaLINDRVV
	<i>A. fumigatus</i> 26906 PLHGCDVDKL	AYfEtMQC.. Ks..... EKEPL VRaLINDRVV
10	<i>A. fumigatus</i> 32239 PLHGCAVDKL	AYfEtMQC.. Ks..... EKEPL VRaLINDRVV
	<i>E. nidulans</i> PLHGCAVDKF	AYfELMQC.. E..... KKEPL VRVLVNDRVV
	<i>T. thermophilus</i> PLHGCEVDsL	AYIEMMQC.. Dd..... sDEPV VRVLVNDRVV
15	<i>T. lanuginosa</i> PLHGCrVDRW	AYVELLRC.. Etetsseeee EG... EDEPF VRVLVNDRVV
	<i>M. thermophila</i> TLkGCGaDER	iYVEkMRC.. sggggggggg EGrqeKDEeM VRVLVNDRVM
20	Basidio PLEfCGgDxd	mvVErLxCxx xgtxxxxxxxx xxxxxxxxxxx VRVLVNDaVq
	Consensus PLHGCGVDKL	AYVEMMQC-- E----- EG---EKEPL VRVLVNDRVV
25	Fcp10 PLHGCGVDKL	AYVEMMQC.. EA..... EKEPL VRVLVNDRVV

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	451	482
A. terreus 9a1	GRCKrDAFVA GLSFAQAG..	GNWADCF--- --
A. terreus cbs	GRCKrDDFVE GLSFARAG..	GNWAECF--- --
A. niger var. awamori	GRCTrDsFVr GLSFARSG..	GDWAECsA-- --
5 A. niger NRRL3135	GRCTrDsFVr GLSFARSG..	GDWAECFA-- --
A. fumigatus 13073	GRCKlNDFVK GLSWARSG..	GNWGECSF-- --
A. fumigatus 32722	GRCKlNDFVK GLSWARSG..	GNWGECSF-- --
A. fumigatus 58128	GRCKlNDFVK GLSWARSG..	GNWGECSF-- --
A. fumigatus 26906	GRCKlNDFVK GLSWARSG..	GNWGECSF-- --
10 A. fumigatus 32239	GRCKlKDFVK GLSWARSG..	GNSEQSFS-- --
E. nidulans	GRCTlDDWVE GLNFARSG..	GNWktCFTl- --
T. thermophilus	GRCKrDDFVr GLSFARqG..	GNWEGCYAas e-
T. lanuginosa	GRCRrDEWIK GLTFARqG..	GHWDrCF--- --
M. thermophila	GmCtlerFIE SMAFARGN..	GKWDlCFA-- --
15 Basidio	GxCtlDAFVE SqxYAReDgq	GDFEKCFAtp xx
Consensus	GRCK-DDFVE GLSFARSG--	GNWEECFA-- --
Fcp10	GRCKRDDFVE GLSFARSG..	GNWEECFA.. ..

20

25

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Figure 5

CP-1

5           Eco RI M G V F V V L L S I A T L F G S T 17  
TATATGAATTCATGGGCGTGTTCGTGCTACTGTCCATTGCCACCTTGTTTCGGTTCCA

1 -----+-----+-----+-----+-----+-----+ 60

ATATACTTAAGTACCCGCACAAGCAGCACGATGACAGGTAACGGTGAACAAGCCAAGGT

10           S G T A L G P R G N S H S C D T V D G G 37  
CATCCGGTACCGCCTTGGGTCTCGTGGTAATTCTCACTCTTGACACTGTTGACGGTG

61 -----+-----+-----+-----+-----+-----+ 120

GTAGGCCATGGCGGAACCCAGGAGCACCATTAAAGAGTGAGAACACTGTGACAACTGCCAC

CP-2

CP-3.10

15           Y Q C F P E I S H L W G Q Y S P E F S L 57  
GTTACCAATGTTTCCAGAAATTTCTCACTTGTGGGGTCAATACTCTCCATTCTTCTCTT

121 -----+-----+-----+-----+-----+-----+ 180

CAATGGTTACAAAGGGTCTTTAAAGAGTGAACACCCCAGTTATGAGAGGTAAGAAGAGAA

20           A D E S A I S P D V P K Q C R V T F V Q 77  
TGGCTGACGAATCTGCTATTTCTCCAGACGTTCCAAAGGGTTGTAGAGTTACTTTTCGTTT

181 -----+-----+-----+-----+-----+-----+ 240

ACCGACTGCTTAGACGATAAAGAGGTCTGCAAGGTTTCCCGACATCTCAATGAAAGCAAG

CP-4.10

CP-5.10

25           V L S R H G A R Y P T S S K S K K Y S A 97  
AAGTTTGTCTAGACACGGTGCTAGATACCCAACTTCTTCTAAGTCTAAGAAGTACTCTG

241 -----+-----+-----+-----+-----+-----+ 300

TTCAAAACAGATCTGTGCCACGATCTATGGGTTGAAGAAGATTGAGATTCTTCATGAGAC

30           L I E A I Q K N A T A F K G K Y A F L K 117  
CTTTGATTGAAGCTATTCAAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGA

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301 -----+-----+-----+-----+-----+-----+-----+ 360

GAAACTAACTTCGATAAGTTTCTTGCATGACGAAAGTTCCCATTCATGCGAAAGAAGT

CP-6

CP-7.10

5 T Y N Y T L G A D D L T P F G E Q Q M V 137

AGACTTACAACACTACTTTGGGTGCTGACGACTTGACTCCATTCCGGTGAACAACAAATGG

361 -----+-----+-----+-----+-----+-----+-----+ 420

TCTGAATGTTGATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTGTGTTTACC

10 N S G I K F Y R R Y K A L A R K I V P F 157

TTAACTCTGGTATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCAT

421 -----+-----+-----+-----+-----+-----+-----+ 480

AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTA

CP-8.10

15

CP-9.10

Y R A S G S D R V I A S A E K F I E G F 177

TCGTTAGAGCTTCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCAATTGAAGGTT

481 -----+-----+-----+-----+-----+-----+-----+ 540

AGCAATCTCGAAGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAAGTTCCAA

20

Q S A K L A D P G A N P H Q A S P V I N 197

TCCAATCTGCTAAGTTGGCTGACCCAGGTGCTAACCACACCAAGCTTCTCCAGTTATTA

541 -----+-----+-----+-----+-----+-----+-----+ 600

AGGTTAGACGATTCAACCGACTGGGTCCACGATTGGGTGTGGTTCGAAGAGGTCAATAAT

25

CP-10.10

CP-11.10

V I I P E G A G Y N N T L D H G L C T A 217

ACGTTATTATTCCAGAAGGTGCTGGTTACAACAACACTTTGGACCACGGTTTGTGTACTG

601 -----+-----+-----+-----+-----+-----+-----+ 660

30

TGCAATAATAAGGTCTTCCACGACCAATGTTGTTGTGAAACCTGGTGCCAAACACATGAC

F E E S E L G D D V E A N F T A Y F A P 237

CTTTCGAAGAATCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTGTTTTCGCTC



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661 -----+-----+-----+-----+-----+-----+ 720  
GAAAGCTTCTTAGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGACAAAAGCGAG  
CP-12.10  
5 P I R A R L E A H L P G V N L T D E D V 257  
CACCTATTAGAGCTAGATTGGAAGCTCACTTGCCAGGTGTTAACTTGACTGACGAAGACG  
721 -----+-----+-----+-----+-----+-----+ 780  
GTGGATAATCTCGATCTAACCTTCGAGTGAACGGTCCACAATTGAACTGACTGCTTCTGC  
CP-13.10  
10 V N L M D M C P F D T V A R T S D A T Q 277  
TTGTAACTTGATGGACATGTGTCCATTGACACTGTTGCTAGAACTTCTGACGCTACTC  
781 -----+-----+-----+-----+-----+-----+ 840  
AACAATTGAACTACCTGTACACAGGTAAGCTGTGACAACGATCTGAAGACTGCGATGAG  
L S P F C D L F T H D E W I Q Y D Y L Q 297  
AATTGTCTCCATTCTGTGACTTGTTCACCTCACGACGAATGGATTCAATACGACTACTTGC  
841 -----+-----+-----+-----+-----+-----+ 900  
TTAACAGAGGTAAGACACTGAACAAGTGAGTGCTGCTTACCTAAGTTATGCTGATGAACG  
CP-14.10  
20 CP-15.10  
S L G K Y Y G Y G A G N P L G P A Q G V 317  
AATCTTTGGGTAAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTG  
901 -----+-----+-----+-----+-----+-----+ 960  
TTAGAAACCCATTTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCAC  
G F Y N E L I A R L T H S P V Q D H T S 337  
TTGGTTTCGTTAACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTT  
961 -----+-----+-----+-----+-----+-----+  
30 1020  
AACCAAAGCAATTGCTTAACTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAA  
CP-16.10  
CP-17.10

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T N H T L D S N P A T F P L N A T L Y A 357  
CTACTAACCACACTTTGGACTCTAACCCAGCTACTTTCCCATTGAACGCTACTTTGTACG  
1021 -----+-----+-----+-----+-----+  
1080  
5 GATGATTGGTGTGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCGATGAAACATGC  
D F S H D N T M Y S I F F A L G L Y N G 377  
CTGACTTCTCTCAGACAACACTATGGTTTCTATTTTCTTCGCTTTGGGTTTGTACAACG  
1081 -----+-----+-----+-----+-----+  
10 1140  
GACTGAAGAGAGTGCTGTTGTGATACCAAAGATAAAAGAAGCGAAACCCAAACATGTTGC  
CP-18.10  
CP-19.10  
T K P L S T T S V E S I E E T D G Y A A 397  
15 GTACTAAGCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACGCTG  
1141 -----+-----+-----+-----+-----+  
1200  
CATGATTCGGTAACAGATGATGAAGACAACCTTAGATAACTTCTTTGACTGCCAATGCGAC  
20 S W T V P F A A R A Y V E M M Q C E A E 417  
CTTCTTGGACTGTTCCATTTCGCTGCTAGAGCTTACGTTGAAATGATGCAATGTGAAGCTG  
1201 -----+-----+-----+-----+-----+  
1260  
GAAGAACCTGACAAGGTAAGCCACGATCTCGAATGCAACTTTACTACGTTACACTTCGAC  
25 CP-20.10  
CP-21.10  
K E P L V R V L V N D R V V P L H G C G 437  
AAAAGGAACCATTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG  
1261 -----+-----+-----+-----+-----+  
30 1320  
TTTTCCTTGGTAACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACAC  
V D K L G R C K R D D F V E G L S F A R 457  
GTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTGTCTTTTCGCTA

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1321 -----+-----+-----+-----+-----+-----+  
 1380  
 CACAACCTGTTCAACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGAT  
 5 S G G N W E E C F A \* Eco RI CP-22.10 467  
 GATCTGGTGGTAACTGGGAAGAATGTTTCGCTTAAGAATTCATATA  
 1381 -----+-----+-----+-----+-----+----- 1426  
 CTAGACCACCATTGACCCTTCTTACAAAGCGAATTCTTAAGTATAT

10

Figure 6

50 1  
 15 *P. involutus* (phyA1) ----- ~FPipeseqR nWSPYSPYFP LAEyKA....  
 pPaGCQInqV  
*P. involutus* (phyA2) ----- ~FsipeseqR nWSPYSPYFP LAEyKA....  
 pPaGCeInqV  
 20 *T. pubescens* ----- ~LDvtRDVqQ sWSmYSPYFP aAtyvA....  
 pPaSCQInqV  
*A. pediades* ----- ~pffpPQIQD sWAaYTPYYP VqAyTP....  
 pPKDCKITqV  
*P. lycii* ----- ~LPipAQnTs nWGPYdPFFP VEPyAA....  
 pPEGctVTqV  
 25 *A. terreus* 9a1 KhSDCNSVDh GYQCfPELSH kWGLYAPYFS LqDESFPFpD  
 VPEDCHITFV  
*A. terreus* cbs NhSDCtSVDr GYQCfPELSH kWGLYAPYFS LqDESFPFpD  
 VPDDCHITFV  
 30 *A. niger* var. awamori NqSTCDTVDq GYQCfSEtSH LWGQYAPFFS LANESAISPD  
 VPAGCRVTFa  
*A. niger* T213 NqSSCDTVDq GYQCfSEtSH LWGQYAPFFS LANESvISPD  
 VPAGCRVTFa  
*A. niger* NRRL3135 NqSSCDTVDq GYQCfSEtSH LWGQYAPFFS LANESvISPE  
 VPAGCRVTFa  
 35 *A. fumigatus* ATCC13073 GSKSCDTVDl GYQCSPAtSH LWGQYSPFFS LEDElSVSSK  
 LPKDCRITLV

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	<i>A. fumigatus</i> ATCC32722	GSkSCDTVD1 GYQcSPAtSH LWGQYSPFFS LEDElSVSSK LPKDCRITLV
	<i>A. fumigatus</i> ATCC58128	GSkSCDTVD1 GYQcSPAtSH LWGQYSPFFS LEDElSVSSK LPKDCRITLV
5	<i>A. fumigatus</i> ATCC26906	GSkSCDTVD1 GYQcSPAtSH LWGQYSPFFS LEDElSVSSK LPKDCRITLV
	<i>A. fumigatus</i> ATCC32239	GSkACDTVE1 GYQcSPGtSH LWGQYSPFFS LEDElSVSSD LPKDCRVTFV
10	<i>E. nidulans</i> VPhGCeVTFV	QNHSCNTaDg GYQcFPNVSH VWGQYSPYFS IEQESAISeD
	<i>T. thermophilus</i> VPQNCKITFV	DSHSCNTVEg GYQcRPEISH sWGQYSPFFS LADQSEISPD
	<i>T. lanuginosa</i> VPKGCRVeFV	----- ----nvDIAR hWGQYSPFFS LAEvSEISPA
15	<i>M. thermophila</i> IPDDCeVTFa	ESRPCDTpDl GFQCgTAISH FWGQYSPYFS VPSElDaS..
	Consensus Seq. 11 VPKGCRVTFV	NSHSCDTVD- GYQC-PeISH LWGQYSPFFS LADESAISPD
20		
	100	51
	<i>P. involutus</i> (phyA1) KSFKYdLGns	NIIqRHGARF PTSGaTtRik AgLtKLQgvq nftDAKFnFI
25	<i>P. involutus</i> (phyA2) KSftYdLGts	NIIqRHGARF PTSGaAtRik AgLsKLQsvq nftDPKFDFI
	<i>T. pubescens</i> tnYtYSLGqD	HIIqRHGARF PTSGaAKRiq TaVAKLKaaS nytDPlLAFV
30	<i>A. pediades</i> tnYtYTLGhD	NIIqRHGARF PTSGaGtRiq AaVKKLQsak TytDPRLDfL
	<i>P. lycii</i> NdFvYkFGvA	NLIqRHGARW PTSGarsRqv AaVAKIQmar PftDPKYEFL
	<i>A. terreus</i> 9a1 QSYNYSLDSE	QVLARHGARS PThSKTKaYA AtIAaIQKSA TaFpGKYAFL
35	<i>A. terreus</i> cbs KSYNYSMGSE	QVLARHGARS PTdSKTKaYA AtIAaIQKNA TaLpGKYAFL
	<i>A. niger</i> var. <i>awamori</i> KTYNYSLGAD	QVLSRHGARY PTeSKGKKYS ALIEeIQQNv TtFDGKYAFL

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	<i>A. niger</i> T213 KTYNYSLGAD	QVLSRHGARY PTeSKGKKYS ALIEeIQQNv TtFDGKYAFL
	<i>A. niger</i> NRRL3135 KTYNYSLGAD	QVLSRHGARY PTdSKGKKYS ALIEeIQQNA TtFDGKYAFL
5	<i>A. fumigatus</i> ATCC13073 KTYNYTLGAD	QVLSRHGARY PTSSKSsKKYk kLVtaIQaNA TdFKGKFAPL
	<i>A. fumigatus</i> ATCC32722 KTYNYTLGAD	QVLSRHGARY PTSSKSsKKYk kLVtaIQaNA TdFKGKFAPL
10	<i>A. fumigatus</i> ATCC58128 KTYNYTLGAD	QVLSRHGARY PTSSKSsKKYk kLVtaIQaNA TdFKGKFAPL
	<i>A. fumigatus</i> ATCC26906 KTYNYTLGAD	QVLSRHGARY PTSSKSsKKYk kLVtaIQaNA TdFKGKFAPL
	<i>A. fumigatus</i> ATCC32239 ETYNYTLGAD	QVLSRHGARY PTASKSsKKYk kLVtaIQKNA TeFKGKFAPL
15	<i>E. nidulans</i> ESYNYTLGAD	QVLSRHGARY PTeSKSKaYS GLIEaIQKNA TsFwGQYAFL
	<i>T. thermophilus</i> KdYrYqLGAN	QLLSRHGARY PTSSKTELYS qLIIsRIQKtA TaYKGyYAFL
20	<i>T. lanuginosa</i> RdYaYhLGAD	QVLSRHGARY PTAhKSEvYA ELLQRIQDtA TeFKGDFAFL
	<i>M. thermophila</i> RTYDYTLGAD	QVLSRHGARA PTlkRAasYv DLIDRIHhGA isYgPgYEFL
25	Consensus Seq. 11 KTYNYTLGAD	QVLSRHGARY PTSSKSsKKYS ALIERIQKNA T-FKGKYAFL

101

150

30	<i>P. involutus</i> (phyA1) VVDSAtNWtA	DLvPFGAAQs fDAGqEaFaR YskLvSKNnL PFIRAdGSDR
	<i>P. involutus</i> (phyA2) VVDtAtNWtA	DLvPFGAAQs fDAGLEvFaR YskLvSsDnL PFIRSDGSDR
	<i>T. pubescens</i> VVATANNWtA	sLveLGAtQs sEAGqEaFtR YsSLvSaDeL PFVRASGSDR
35	<i>A. pediades</i> VVDSAtNWtE	DLvPFGAlQs sQAGeEtFQR YsfLvSKEnL PFVRASSNR
	<i>P. lycii</i> VVDSStNWtA	DLlPFGANQs hQTGtDMYtR YsTLfEgGdV PFVRAAGdQR

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	<i>A. terreus</i> 9a1 VhESA EK FVE	ELTPFG rNQL rDlGaQFYeR YNAL.TRHIn PFVRATDAsR
	<i>A. terreus</i> cbs VhESA EK FVE	NLTPFG rNQL qDlGaQFYRR YDTL.TRHIn PFVRAADSsR
5	<i>A. niger</i> var. <i>awamori</i> VIASGEKFIE	DLTPFG EQEL VNSGIKFYQR YESL.TRNII PFIRSSGSsR
	<i>A. niger</i> T213 VIASGEKFIE	DLTPFG EQEL VNSGIKFYQR YESL.TRNII PFIRSSGSsR
10	<i>A. niger</i> NRRL3135 VIASGKKFIE	DLTPFG EQEL VNSGIKFYQR YESL.TRNIV PFIRSSGSsR
	<i>A. fumigatus</i> ATCC13073 VIASGEKFIE	DLTPFG EQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR
	<i>A. fumigatus</i> ATCC32722 VIASGEKFIE	DLTPFG EQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR
15	<i>A. fumigatus</i> ATCC58128 VIASGEKFIE	DLTPFG EQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR
	<i>A. fumigatus</i> ATCC26906 VIASGEKFIE	DLTAFGEQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR
20	<i>A. fumigatus</i> ATCC32239 VIASGEKFIE	DLTPFG EQQM VNSGIKFYQK YKAL.AgSVV PFIRSSGSsR
	<i>E. nidulans</i> VVASAEKFIN	DLTiFGENQM VD SGaKFYRR YKnL.ARKnt PFIRASGSDR
	<i>T. thermophilus</i> VIASGrIFIE	DLTPFGENQM IQlGIKFYnH YKSL.ARN aV PFVRCSGSDR
25	<i>T. lanuginosa</i> VIASAEfFnr	NLTRFGEEQM MESGrQFYHR YREq.AREIV PFVRAAGSAR
	<i>M. thermophila</i> VhSAENFtQ	ELTRtGQQQM VNSGIKFYRR YRAL.ARKsI PFVRTAGqDR
30	Consensus Seq. 11 VIASAEKFIE	DLTPFGENQM VNSGIKFYRR YKAL-ARNIV PFVRASGSDR
		151
	200	
35	<i>P. involutus</i> (phyA1) PAaGD.....	GFaSA..... ..shNtvqPk LNLILPQ..T gNDTLEDNMC
	<i>P. involutus</i> (phyA2) PAaGE.....	GFaSA..... ..srNaiqPk LDLILPQ..T gNDTLEDNMC

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	<i>T. pubescens</i> PAaGD.....	GFaIA..... ..ssNsITPV LSVIISE..A gNDTLDDNMC
	<i>A. pediades</i> PnaGs.....	GFsAA..... ..shHvINPI LfVILSE..S LNDTLDDAMC
5	<i>P. lycii</i> PnevD.....	GFgdA..... ..sgEtvIPt LQVVLQE..E gNcTLcNNMC
	<i>A. terreus</i> 9a1 TAFES...ST	GFQTARqDDh hAnpHQPSPr VDVaIPEGSA YNNTLEHSLC
10	<i>A. terreus</i> cbs TAFEA...ST	GFQNArqGDP hAnpHQPSPr VDVVIPEGTA YNNTLEHSIC
	<i>A. niger</i> var. <i>awamori</i> TvFED...Se	GFQSTKLkDP rAqpgQSSPk IDVWISEASS sNNTLDpGtC
	<i>A. niger</i> T213 TvFED...Se	GFQSTKLkDP rAqpgQSSPk IDVWISEASS sNNTLDpGtC
15	<i>A. niger</i> NRRL3135 TvFED...Se	GFQSTKLkDP rAqpgQSSPk IDVWISEASS sNNTLDpGtC
	<i>A. fumigatus</i> ATCC13073 TkFEA...Sq	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC
20	<i>A. fumigatus</i> ATCC32722 TkFEA...Sq	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC
	<i>A. fumigatus</i> ATCC58128 TkFEA...Sq	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC
	<i>A. fumigatus</i> ATCC26906 TkFEA...Sq	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC
25	<i>A. fumigatus</i> ATCC32239 TnFEA...Se	GFQqANVADP gAt.NRAAPV ISVIIPESeT YNNTLDHSVC
	<i>E. nidulans</i> vSFEN...de	GFRkaQLhDh g.s.gQATPV VNVIIPeIdG FNNTLDHStC
30	<i>T. thermophilus</i> PvFED...SS	GFQSAKVlDP hSdKHDPpT INVIIeEGPS YNNTLDtGsC
	<i>T. lanuginosa</i> PAaEE...AP	GFQdAKdrDP rSnkDQAePV INVIISEETG sNNTLDgltC
	<i>M. thermophila</i> TAFEEgpyST	GFHSALLADR gStvRPTlPy dmVVIPETAG aNNTLHNDLC
35		
	Consensus Seq. 11 TAFED---ST	GFQSAKLADP -A--HQASPV INVIIPEGSG YNNTLDHGLC

		201
	250	
	<i>P. involutus</i> (phyA1) LCAFlTVSK.	.SDpqvnaWl AVafPSItAR LNAaaPSVNL TDtDafNLVs
5	<i>P. involutus</i> (phyA2) LCPFmTVSK.	.SDpqvDaWl AsafPSVtAQ LNAaaPGaNL TDADafNLVs
	<i>T. pubescens</i> LCPFETVAt.	.SDpqvnQWl AqFAPPMtAR LNAgaPGaNL TDtDtyNLLt
10	<i>A. pediades</i> LCAFETivK.	.SDpqtGiWT SIYGTPIanR LNqqaPGaNI TAADVsnLIp
	<i>P. lycii</i> MCPFDTLSS.	.GDESt.tWl GVFAPnItAR LNAaaPSaNL SDsDaLtLMD
	<i>A. terreus</i> 9a1 MCPFETVS1T	VGDDAvANFT AVFAPAIaqR LEAdLPGVQL StDDVVNLMA
15	<i>A. terreus</i> cbs MCPFETVS1T	VGDAADNFT AVFAPAIakR LEAdLPGVQL SADDVVNLMA
	<i>A. niger</i> var. <i>awamori</i> MCSFDTISs	LADtveANFT AtFAPSIRqR LEndLSGVtL TDtEVtyLMD
20	<i>A. niger</i> T213 MCSFDTISs	LADtveANFT AtFAPSIRqR LEndLSGVtL TDtEVtyLMD
	<i>A. niger</i> NRRL3135 MCSFDTISs	LADtveANFT AtFvPSIRqR LEndLSGVtL TDtEVtyLMD
	<i>A. fumigatus</i> ATCC13073 MCSFDTVART	LGDEvAANFT ALFAPdIRAR aekhLPGVtL TDEDVVSLMD
25	<i>A. fumigatus</i> ATCC32722 MCSFDTVART	LGDEvAANFT ALFAPdIRAR aekhLPGVtL TDEDVVSLMD
	<i>A. fumigatus</i> ATCC58128 MCSFDTVART	LGDEvAANFT ALFAPdIRAR aekhLPGVtL TDEDVVSLMD
30	<i>A. fumigatus</i> ATCC26906 MCSFDTVART	LGDEvAANFT ALFAPdIRAR aKkhLPGVtL TDEDVVSLMD
	<i>A. fumigatus</i> ATCC32239 MCSFDTVART	LGDEvEANFT ALFAPAIRAR IEkhLPGVQL TDDDVVSLMD
	<i>E. nidulans</i> MCSFDTMART	rADEiEANFT AIMGPPIrkR LEndLPGIKL TNENViyLMD
35	<i>T. thermophilus</i> LCPFETLARN	gGHDAQEKFA kqFAPAILEK IKDhLPGVDL AvesDVpyLMD
	<i>T. lanuginosa</i> LCPFDTVGsd	.DptqpAEFl qVFGPRVlkK ItkhMPGVNL TLEDVplFMD



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	<i>M. thermophila</i> LCPFETVAsS	IGDDAQDtYl StFAGPiTAR VNAnLPGaNL TDADtVaLMD
5	Consensus Seq. 11 MCPFDTVART	LGDDAEANFT AVFAPPiRAR LEA-LPGVNL TDDEDVVNLMD
		251
	300	
10	<i>P. involutus</i> (phyA1) dKFYGTGyGQ	..... ekkSdF CtLFegiPGs FeaFAYggdL
	<i>P. involutus</i> (phyA2) dKFYGTGyGQ	..... eqkSdF CtLFegiPGs FeaFAYagdL
	<i>T. pubescens</i> dKFYGTGyGQ	..... errSeF CDiYeelqAE .daFAYnadL
15	<i>A. pediades</i> dKFYGTGyGQ	..... etpSPF CNLF..TPEE FaQFEYfgdL
	<i>P. lycii</i> dKYYGTGPGN	..... gnaSPF CDLF..TAAE YvsYEYYydL
20	<i>A. terreus</i> 9a1 dKYYGYGGGN	dD..Aht... ..LSPF CDLF..TatE WtQYNYLlSL
	<i>A. terreus</i> cbs dKYYGYGGGN	dD..Aht... ..LSPF CDLF..TAAE WtQYNYLlSL
	<i>A. niger</i> var. <i>awamori</i> kKYYGHGAGN	Tv..DTK... ..LSPF CDLF..ThDE WiHYDYlQSL
25	<i>A. niger</i> T213 kKYYGHGAGN	Tv..DTK... ..LSPF CDLF..ThDE WiHYDYlRSL
	<i>A. niger</i> NRRL3135 kKYYGHGAGN	Tv..DTK... ..LSPF CDLF..ThDE WiNYDYlQSL
30	<i>A. fumigatus</i> ATCC13073 gKYYGYGAGN	SD..ASQ... ..LSPF CQLF..ThNE WkKYNYlQSL
	<i>A. fumigatus</i> ATCC32722 gKYYGYGAGN	SD..ASQ... ..LSPF CQLF..ThNE WkKYNYlQSL
	<i>A. fumigatus</i> ATCC58128 gKYYGYGAGN	SD..ASQ... ..LSPF CQLF..ThNE WkKYNYlQSL
35	<i>A. fumigatus</i> ATCC26906 gKYYGYGAGN	SD..ASQ... ..LSPF CQLF..ThNE WkKYNYlQSL
	<i>A. fumigatus</i> ATCC32239 gKYYGYGAGN	AD..ASE... ..LSPF CAIF..ThNE WkKYDYlQSL

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	<i>E. nidulans</i> sKYYGYGAGS	AH..GTE... ..LSPF CAIF..TEKE WlQYDYLQSL
	<i>T. thermophilus</i> gKYYGnGGGN	ht..DT.... ..LSPF CALs..TqEE WqayDYyQSL
5	<i>T. lanuginosa</i> dKYYSHGGGS	PvlfPrQ... ..LSPF CHLF..TADD WmaYDYyTL
	<i>M. thermophila</i> gKWYGYGPGN	SsdpATadag ggngprLSPF CrLF..SEsE WraYDYLQSV
10	Consensus Seq. 11 KYYGYGAGN	SD--ATQ--- -----LSPF CDLF--TADK W-QYDYLQSL -
		301
	350	
15	<i>P. involutus</i> (phyA1) FPLNkTFYAD	eLGPvQGvGY vNELIARLTN S.AVRDNTqT NRTLdASPvT
	<i>P. involutus</i> (phyA2) FPLNkTMYAD	ALGPvQGvGY iNELLARLTN S.AVNDNTqT NRTLDAApDT
20	<i>T. pubescens</i> FPLNrTLyAD	PLGPvQGvGY iNELIARLTa q.nVsDHTqT NsTLdSSPET
	<i>A. pediades</i> FPLDrSIYAD	PLGPvQGvGY iNELLARLTm m.PVRDNTqT NRTLdSSPlT
	<i>P. lycii</i> FPLNrTFYAD	ALGPvQGvGY vNELLARLTg q.AVRDETqT NRTLdSDPAT
25	<i>A. terreus</i> 9a1 FPLNATLYAD	PLGPvQGvGW aNELMARLTR A.PVHDHTCv NNTLDASPAT
	<i>A. terreus</i> cbs FPLNATLYAD	PLGPvQGvGW aNELIARLTR S.PVHDHTCv NNTLDANPAT
30	<i>A. niger</i> var. <i>awamori</i> FPLNSTLYAD	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDsNPAT
	<i>A. niger</i> T213 FPLNSTLYAD	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDsNPAT
	<i>A. niger</i> NRRL3135 FPLNSTLYAD	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSSPAT
35	<i>A. fumigatus</i> ATCC13073 FPLNATMYvD	PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT
	<i>A. fumigatus</i> ATCC32722 FPLNATMYvD	PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT

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	<i>A. fumigatus</i> ATCC58128	PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT FPLNATMYvD
	<i>A. fumigatus</i> ATCC26906	PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT FPLNATMYvD
5	<i>A. fumigatus</i> ATCC32239	PLGPAQGIGF tNELIARLTN S.PVQDHTST NsTLDSNPAT FPLNATIYvD
	<i>E. nidulans</i> FPLDrkLYAD	PLGPAQGIGF tNELIARLTQ S.PVQDNTST NHTLDSNPAT
10	<i>T. thermophilus</i> FPLNATLYAD	PLGPAQGVGF vNELIARMTH S.PVQDYTTv NHTLDSNPAT
	<i>T. lanuginosa</i> FPLDAvLYAD	AFGPSRGVGF vNELIARMTg NlPVKDHTTv NHTLDdNPET
	<i>M. thermophila</i> FPLGrPLYAD	PLGPTQGVGF vNELLARLA. GvPVRDgTST NRTLGDGPrt
15	Consensus Seq. 11 FPLNATLYAD	PLGPAQGVGF -NELIARLTH S-PVQDHTST NHTLDSNPAT
		351
20	400	
	<i>P. involutus</i> (phyA1) TSSlVPFSGR	FSHDNlMVAV FsAMGLFrqP aPLSTsvpNP wrt.....Wr
	<i>P. involutus</i> (phyA2) TSSvVPFSAR	FSHDNlMVAV FsAMGLFrqS aPLSTSTpDP nrt.....Wl
25	<i>T. pubescens</i> vkkivPFASR	FSHDNqMVAI FsAMGLFNqS aPLdPTTpDP art.....Fl
	<i>A. pediades</i> TSRltPFASR	LSHDNqMIAI FsAMGLFNqS sPLdPSfpNP krt.....Wv
30	<i>P. lycii</i> DSklVPFSGH	FSHDNTMVPI FaALGLFNAT a.LdPlkpDe nrl.....Wv
	<i>A. terreus</i> 9a1 AAWTVPFAAR	FSHDSnLVSI FWALGLYNGT aPLSqTSVES Vs..QTDGYA
	<i>A. terreus</i> cbs AAWTVPFAAR	FSHDSnLVSI FWALGLYNGT KPLSqTTVED It..rTDGYA
35	<i>A. niger</i> var. <i>awamori</i> SAWTVPFASR	FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS
	<i>A. niger</i> T213 SAWTVPFASR	FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS

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	<i>A. niger</i> NRRL3135 SAWTVPFASR	FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS
	<i>A. fumigatus</i> ATCC13073 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..EldGYS
5	<i>A. fumigatus</i> ATCC32722 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT gPLSrTSVES ak..EldGYS
	<i>A. fumigatus</i> ATCC58128 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..EldGYS
10	<i>A. fumigatus</i> ATCC26906 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..EldGYS
	<i>A. fumigatus</i> ATCC32239 ASWAVPFGAR	FSHDNGMIPI FFAMGLYNGT EPLSqTSeES tk..ESNGYS
	<i>E. nidulans</i> ASWTVPFGAR	FSHDNSMISI FFAMGLYNGT QPLSmdSVES Iq..EmDGYA
15	<i>T. thermophilus</i> AAWTVPFGGR	FSHDNTMtSI FaALGLYNGT akLSTTeIKS Ie..ETDGYS
	<i>T. lanuginosa</i> ASWTVPFAAR	FSHDNTMtGI FsAMGLYNGT KPLSTSkIQP ptgaAADGYA
20	<i>M. thermophila</i> ASWAVPFAAR	FSHDNdMMGV LgALGaYDGV pPLdkTArrd ..peElGGYA
	Consensus Seq. 11 ASWTVPFAAR	FSHDNTMVSI FFALGLYNGT KPLSTTSVES I---ETDGYA
25	450	401
	<i>P. involutus</i> (phyA1) PLEfCGgDRn	mvVERLsC.. fGt..... Tk VRVLVQDQVq
30	<i>P. involutus</i> (phyA2) PLEfCGgDQd	maVERLsC.. AGt..... Tk VRVLVQDQVq
	<i>T. pubescens</i> PLafCGaDts	mvVERLDC.. GGa..... Qs VRLLVNDaVq
	<i>A. pediades</i> PLkfCGgDmd	mvteRlLCQr DGtGsGGpsr imrNgnvQTF VRILVNDaLq
35	<i>P. lycii</i> PLEfCGg.vd	mtVEkLaC.. .....sgKea VRVLVNDaVq
	<i>A. terreus</i> 9a1 PLHGCPtDKL	AYVEMMQCrA .....EK...EPL VRVLVNDRVM

	<i>A. terreus</i> cbs PLHGCAVDNL	AYIEMMQCrA ..... ..EK...QPL VRVLVNDVRVM
	<i>A. niger</i> var. <i>awamori</i> PLHGCPIDaL	1YVEMMQCQA ..... ..EQ...EPL VRVLVNDRVV
5	<i>A. niger</i> T213 PLHGCPIDaL	1YVEMMQCQA ..... ..EQ...EPL VRVLVNDRVV
	<i>A. niger</i> NRRL3135 PLHGCPVDaL	1YVEMMQCQA ..... ..EQ...EPL VRVLVNDRVV
10	<i>A. fumigatus</i> ATCC13073 PLHGCDVDKL	AYfEtMQCKS ..... ..EK...EPL VRaLINDRVV
	<i>A. fumigatus</i> ATCC32722 PLHGCDVDKL	AYfEtMQCKS ..... ..EK...EPL VRaLINDRVV
	<i>A. fumigatus</i> ATCC58128 PLHGCDVDKL	AYfEtMQCKS ..... ..EK...ESL VRaLINDRVV
15	<i>A. fumigatus</i> ATCC26906 PLHGCDVDKL	AYfEtMQCKS ..... ..EK...EPL VRaLINDRVV
	<i>A. fumigatus</i> ATCC32239 PLHGCAVDKL	AYfEtMQCKS ..... ..EK...EPL VRaLINDRVV
20	<i>E. nidulans</i> PLHGCAVDKF	AYfELMQCE. .... ..KK...EPL VRVLVNDRVV
	<i>T. thermophilus</i> PLHGCEVDsL	AYIEMMQCDD ..... ..sD...EPV VRVLVNDRVV
	<i>T. lanuginosa</i> PLHGCrVDRW	AYVELLRcET ETsSeEEeEG ..ED...EPF VRVLVNDRVV
25	<i>M. thermophila</i> TLkGCGaDEr	iYVEkMRCsG GGgGgGGgEG ..rQekdEeM VRVLVNDVRM
	Consensus Seq. 11 PLHGCGVDKL	AYVEMMQCEA GG-G-GG-EG --EK---EPL VRVLVNDRVV
30		
		451 482
	<i>P. involutus</i> (phyA1)	G1CtLAKFVE SqTFARSDga GDFEKCFats a-
	<i>P. involutus</i> (phyA2)	G1CaLDKFVE SqAYARSGga GDFEKCLAtt v-
	<i>T. pubescens</i>	GvCtLDAFVE SqAYARNDge GDFEKCFat- --
35	<i>A. pediades</i>	S1CtLEAFVE SqkYAReDgq GDFEKCFD-- --
	<i>P. lycii</i>	GvCELSAFVE SqTYAReNgq GDFAKCgfvp se

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	<i>A. terreus</i> 9a1	GRCKrDAFVA GLSFAQAG.. GNWADCF----	--
	<i>A. terreus</i> cbs	GRCKrDDFVE GLSFARAG.. GNWAECF----	--
	<i>A. niger</i> var. <i>awamori</i>	GRCtrDsFVr GLSFARSG.. GDWAECsA--	--
	<i>A. niger</i> T213	GRCtrDsFVr GLSFARSG.. GDWAECFA--	--
5	<i>A. niger</i> NRRL3135	GRCtrDsFVr GLSFARSG.. GDWAECFA--	--
	<i>A. fumigatus</i> ATCC13073	GRCKLNDFVK GLSWARSG.. GNWGECS--	--
	<i>A. fumigatus</i> ATCC32722	GRCKLNDFVK GLSWARSG.. GNWGECS--	--
	<i>A. fumigatus</i> ATCC58128	GRCKLNDFVK GLSWARSG.. GNWGECS--	--
	<i>A. fumigatus</i> ATCC26906	GRCKLNDFVK GLSWARSG.. GNWGECS--	--
10	<i>A. fumigatus</i> ATCC32239	GRCKLKDFVK GLSWARSG.. GNSEQSFS--	--
	<i>E. nidulans</i>	GRcTLDDWVE GLNFARSG.. GNWktCFTl-	--
	<i>T. thermophilus</i>	GRCKrDDFVr GLSFARqG.. GNWEGCYAas e-	
	<i>T. lanuginosa</i>	GRCRrDEWIK GLTFARqG.. GHWDrCF---	--
	<i>M. thermophila</i>	GmCtLErFIE SMAFARGN.. GKWDlCFA--	--
15			
	Consensus Seq. 11	GRCKLDDFVE GLSFARSG-- GNWAECFA--	--

20

25

Figure 7

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20 M G V F V V L L S I A T L F G S T S G T  
ATGGGCGTGTTTCGTGCTACTGTCCATTGCCACCTTGTTTCGGTTCACATCCGGTACC

5 1 ---+-----+-----+-----+-----+-----+-----  
60 TACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAAACAAGCCAAGGTGTAGGCCATGG

10 40 A L G P R G N S H S C D T V D G G Y Q C  
GCCTTGGGTCCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGTGGTTACCAATGT

120 61 ---+-----+-----+-----+-----+-----+-----  
CGGAACCCAGGAGCACCATTAAGAGTGAGAACACTGTGACAACCTGCCACCAATGGTTACA

15 60 F P E I S H L W G T Y S P Y F S L A D E  
TTCCCAGAAATTTCTCACTTGTGGGGTACCTACTCTCCATACTTCTCTTTGGCAGACGAA

20 180 121 ---+-----+-----+-----+-----+-----+-----  
AAGGGTCTTTAAAGAGTGAACACCCCATGGATGAGAGGTATGAAGAGAAACCGTCTGCTT

80 S A I S P D V P D D C R V T F V Q V L S  
TCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTTCGTTCAAGTTTGTCT

25 240 187 ---+-----+-----+-----+-----+-----+-----  
AGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAGTTCAAAACAGA

30 100 R H G A R Y P T S S A S K A Y S A L I E  
AGACACGGTGCTAGATACCCAACCTTCTTCTGCGTCTAAGGCTTACTCTGCTTTGATTGAA

300 241 ---+-----+-----+-----+-----+-----+-----  
TCTGTGCCACGATCTATGGGTTGAAGAAGACGCAGATTCCGAATGAGACGAACTAACTT

35 120 A I Q K N A T A F K G K Y A F L K T Y N

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GCTATTCAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGAAGACTTACAAC  
301 ---+-----+-----+-----+-----+-----+-----+  
360  
CGATAAGTTTTCTTGCGATGACGAAAGTTCCCATTCATGCGAAAGAACTTCTGAATGTTG  
5  
Y T L G A D D L T P F G E N Q M V N S G  
140  
TACACTTTGGGTGCTGACGACTTGACTCCATTCCGGTGAAAACCAAATGGTTAACTCTGGT  
361 ---+-----+-----+-----+-----+-----+-----+  
10 420  
ATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTTTGGTTTACCAATTGAGACCA  
I K F Y R R Y K A L A R K I V P F I R A  
160  
ATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCATTCATTAGAGCT  
15  
421 ---+-----+-----+-----+-----+-----+-----+  
480  
TAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTCTAACAAGGTAAGTAATCTCGA  
S G S D R V I A S A E K F I E G F Q S A  
20  
180  
TCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTTTCCAATCTGCT  
481 ---+-----+-----+-----+-----+-----+-----+  
540  
AGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACTTCCAAAGGTTAGACGA  
25  
K L A D P G S Q P H Q A S P V I N V I I  
200  
AAGTTGGCTGACCCAGGTTCTCAACCACACCAAGCTTCTCCAGTTATTAACGTGATCATT  
30  
541 ---+-----+-----+-----+-----+-----+-----+  
600  
TTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTCGAAGAGGTCAATAATTGCACTAGTAA  
P E G S G Y N N T L D H G T C T A F E D  
35 220  
CCAGAAGGATCCGGTTACAACAACACTTTGGACCACGGTACTTGTACTGCTTTTGAAGAC



601 ---+-----+-----+-----+-----+-----+-----  
660  
GGTCTTCCTAGGCCAATGTTGTTGTGAAACCTGGTGCCATGAACATGACGAAAGCTTCTG  
5  
240 S E L G D D V E A N F T A L F A P A I R  
TCTGAATTAGGTGACGACGTTGAAGCTAACTTCACTGCTTTGTTGCTCCAGCTATTAGA  
661 ---+-----+-----+-----+-----+-----+-----  
720  
10 AGACTTAATCCACTGCTGCAACTTCGATTGAAGTGACGAAACAAGCGAGGTCGATAATCT  
A R L E A D L P G V T L T D E D V V Y L  
260  
GCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGACGAAGACGTTGTTACTTG  
15 721 ---+-----+-----+-----+-----+-----+-----  
780  
CGATCTAACCTTCGACTGAACGGTCCACAATGAAACTGACTGCTTCTGCAACAAATGAAC  
M D M C P F D T V A R T S D A T E L S P  
20 280  
ATGGACATGTGTCCATTCGACACTGTCGCTAGAACTTCTGACGCTACTGAATTGTCTCCA  
781 ---+-----+-----+-----+-----+-----+-----  
840  
TACCTGTACACAGGTAAGCTGTGACAGCGATCTTGAAGACTGCGATGACTTAACAGAGGT  
25  
300 F C A L F T H D E W I Q Y D Y L Q S L G  
TTCTGTGCTTTGTTCACTCACGACGAATGGATCCAATACGACTACTTGCAAAGCTTGGGT  
841 ---+-----+-----+-----+-----+-----+-----  
30 900  
AAGACACGAAACAAGTGAGTGCTGCTTACCTAGGTTATGCTGATGAACGTTTCGAACCCA  
K Y Y G Y G A G N P L G P A Q G V G F A  
320  
35 AAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTGTTGGTTTCGCT  
901 ---+-----+-----+-----+-----+-----+-----  
960

Modtag  
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TTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCACAACCAAAGCGA  
N E L I A R L T H S P V Q D H T S T N H  
340  
5 AACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTTCTACTAACCAC  
961 ---+-----+-----+-----+-----+-----+-----  
1020  
TTGCTTAACTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAAGATGATTGGTG  
10 T L D S N P A T F P L N A T L Y A D F S  
360  
ACTTTGGACTCTAACCCAGCTACTTTCCCATTGAACGCTACTTTGTACGCTGACTTCTCT  
1021 ---+-----+-----+-----+-----+-----+-----  
1080  
15 TGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCGATGAAACATGCGACTGAAGAGA  
H D N I M I S I F F A L G L Y N G T K P  
380  
CACGACAACACTATGATATCTATTTTCTTCGCTTTGGGTTTGTACAACGGTACCAAGCCA  
20 1081 ---+-----+-----+-----+-----+-----+-----  
1140  
GTGCTGTTGTGATACTATAGATAAAAGAAGCGAAACCCAAACATGTTGCCATGGTTCGGT  
L S T T S V E S I E E T D G Y S A S W T  
25 400  
TTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTGCTTCTTGGACT  
1141 ---+-----+-----+-----+-----+-----+-----  
1200  
AACAGATGATGAAGACAACCTTAGATAACTTCTTTGACTGCCAATGAGACGAAGAACCTGA  
30 V P F A A R A Y V E M M Q C Q A E K E P  
420  
GTTCCATTGCTGCTAGAGCTTACGTTGAAATGATGCAATGTCAAGCTGAAAAGGAACCA  
1201 ---+-----+-----+-----+-----+-----+-----  
35 1260  
CAAGGTAAGCGACGATCTCGAATGCAACTTTACTACGTTACAGTTCGACTTTTCCTTGGT

Modtaget PD  
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440 L V R V L V N D R V V P L H G C A V D K  
TTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTGCTGTTGACAAG  
5 1261 ---+-----+-----+-----+-----+-----+-----  
1320  
AACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACACGACAACCTGTTTC  
10 460 L G R C K R D D F V E G L S F A R S G G  
TTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTTCGCTAGATCTGGTGGT  
1321 ---+-----+-----+-----+-----+-----+-----  
1380  
AACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGATCTAGACCACCA  
15 N W A E C F A \* 467  
AACTGGGCTGAATGTTTCGCTTAA  
1381 ---+-----+-----+ 1410  
TTGACCCGACTTACAAAGCGAATT

20

25

30

5

Figure 8

10		M G V F V V L L S I A T L F G S T S G T
20		ATGGGCGTGTTTCGTCTGTGCTACTGTCCATTGCCACCTTGTTCGGTTCCACATCCGGTACC
15	60	1 -----+-----+-----+-----+-----+-----+
		TACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGTGTAGGCCATGG
40		A L G P R G N S H S C D T V D G G Y Q C
20		GCCTTGGGTCCTCGTGGTAACTCTCACTCTTGTGACACTGTTGACGGTGGTTACCAATGT
	120	61 -----+-----+-----+-----+-----+-----+
		CGGAACCCAGGAGCACCATTGAGAGTGAGAACACTGTGACAACTGCCACCAATGGTTACA
25	A	F P E I S H L W G T Y S P F F S L A D E
	60	TTCCCAGAAATTTCTCACTTGTGGGGTACATACTCTCCATTCTTCTCTTTGGCTGACGAA
	180	121 -----+-----+-----+-----+-----+-----+
30		AAGGGTCTTTAAAGAGTGAACACCCCATGTATGAGAGGTAAGAAGAGAAACCGACTGCTT

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- 96 -

80 S A I S P D V P K G C R V T F V Q V L S

TCTGCTATTCTCCAGACGTTCCAAAGGGTTGTAGAGTTACTTTCGTTCAAGTTTTGTCT

5 181 -----+-----+-----+-----+-----+

240 AGACGATAAAGAGGTCTGCAAGGTTCCCAACATCTCAATGAAAGCAAGTTCAAAACAGA

10 100 R H G A R Y P T S S A S K A Y S A L I E

AGACACGGTGCTAGATACCCAACTTCTTCTGCGTCTAAGGCGTACTCTGCTTTGATTGAA

241 -----+-----+-----+-----+-----+

300 TCTGTGCCACGATCTATGGGTTGAAGAAGACGCAGATTCCGCATGAGACGAACTAACTT

15 120 A I Q K N A T A F K G K Y A F L K T Y N

GCTATTCAAAGAAGCGTACTGCTTTCAAGGGTAAGTACGCTTTCTTGAAGACTTACAAC

301 -----+-----+-----+-----+-----+

20 360 CGATAAGTTTTCTTGCATGACGAAAGTTCCCATTCATGCGAAAGAACTTCTGAATGTTG

A 140 Y T L G A D D L T P F G E Q Q M V N S G

25 TACACTTTGGGTGCTGACGACTTGACTCCATTCGGTGAACAACAAATGGTTAACTCTGGT

361 -----+-----+-----+-----+-----+

420 ATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTGTTGTTTACCAATTGAGACCA

30 160 I K F Y R R Y K A L A R K I V P F I R A

ATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCATTCATTAGAGCT

421 -----+-----+-----+-----+-----+

480 TAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTAAGTAATCTCGA

35 180 S G S D R V I A S A E K F I E G F Q S A

22 JAN. 1999

- 97 -

TCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTTTCCAATCTGCT  
481 -----+-----+-----+-----+-----+-----+  
540  
AGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACCTCCAAAGGTTAGACGA  
5  
K L A D P G A N P H Q A S P V I N V I I  
200  
AAGTTGGCTGACCCAGGTGCTAACCACACCAAGCTTCTCCAGTTATTAACGTTATTATT  
541 -----+-----+-----+-----+-----+-----+  
10 600  
TTCAACCGACTGGGTCCACGATTGGGTGTGGTTCGAAGAGGTCAATAATTGCAATAATAA  
P E G A G Y N N T L D H G L C T A F E E  
220  
CCAGAAGGTGCTGGTTACAACAACACTTTGGACCACGGTTTGTGTACTGCTTTCGAAGAA  
15  
601 -----+-----+-----+-----+-----+-----+  
660  
GGTCTTCCACGACCAATGTTGTTGTGAAACCTGGTGCCAAACACATGACGAAAGCTTCTT  
20  
S E L G D D V E A N F T A V F A P P I R  
240  
TCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTGTTTTTCGCTCCACCAATTAGA  
661 -----+-----+-----+-----+-----+-----+  
720  
AGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGACAAAAGCGAGGTGGTTAATCT  
25  
A R L E A H L P G V N L T D E D V V N L  
260  
GCTAGATTGGAAGCTCACTTGCCAGGTGTTAACTTGACTGACGAAGACGTTGTAACTTG  
30  
721 -----+-----+-----+-----+-----+-----+  
780  
CGATCTAACCTTCGAGTGAACGGTCCACAATTGAACTGACTGCTTCTGCAACAATTGAAC  
M D M C P F D T V A R T S D A T Q L S P  
35 280  
ATGGACATGTGTCCATTGACACTGTTGCTAGAACTTCTGACGCTACTCAATTGTCTCCA

22 JAN. 1953

- 98 -

840 781 -----+-----+-----+-----+-----+-----+  
TACCTGTACACAGGTAAGCTGTGACAACGATCTTGAAGACTGCGATGAGTTAACAGAGGT  
5 300 F C D L F T H D E W I Q Y D Y L Q S L G  
TTCTGTGACTTGTTCACTCACGACGAATGGATTCAATACGACTACTTGCAATCTTTGGGT  
900 841 -----+-----+-----+-----+-----+-----+  
10 AAGACACTGAACAAGTGAGTGCTGCTTACCTAAGTTATGCTGATGAACGTTAGAAACCCA  
320 K Y Y G Y G A G N P L G P A Q G V G F V  
AAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTGTTGGTTTCGTT  
15 901 -----+-----+-----+-----+-----+-----+  
960 TTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCACAAACCAAAGCAA  
20 340 N E L I A R L T H S P V Q D H T S T N H  
AACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTTCTACTAACCAC  
1020 961 -----+-----+-----+-----+-----+-----+  
TTGCTTAACTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAAGATGATTGGTG  
25 360 T L D S N P A T F P L N A T L Y A D F S  
ACTTTGGACTCTAACCAGCTACTTTCCCATTTGAACGCTACTTTGTACGCTGACTTCTCT  
1021 -----+-----+-----+-----+-----+-----+  
30 1080 TGAAACCTGAGATTGGGTCGATGAAAGGGTAACCTGCGATGAAACATGCGACTGAAGAGA  
380 H D N T M V S I F F A L G L Y N G T K P  
35 CACGACAACACTATGGTTTCTATTTTCTTCGCTTTGGGTTTGTACAACGGTACTAAGCCA  
1081 -----+-----+-----+-----+-----+-----+  
1140

Modtaget PD  
22 JAN. 1959

- 99 -

GTGCTGTTGTGATACCAAAGATAAAAGAAGCGAAACCCAAACATGTTGCCATGATTCCGGT

400 L S T T S V E S I E E T D G Y S A S W T

5 TTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTGCTTCTTGGACT

1141 -----+-----+-----+-----+-----+-----+-----+  
1200

AACAGATGATGAAGACAACCTTAGATAACTTCTTTGACTGCCAATGAGACGAAGAACCTGA

10 V P F A A R A Y V E M M Q C E A E K E P

420

GTTCCATTGCTGCTAGAGCTTACGTTGAAATGATGCAATGTGAAGCTGAAAAGGAACCA

1201 -----+-----+-----+-----+-----+-----+-----+  
1260

15 CAAGGTAAGCGACGATCTCGAATGCAACTTTACTACGTTACACTTCGACTTTTCCTTGGT

L V R V L V N D R V V P L H G C G V D K

440

TTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTGGTGTGACAAG

20 1261 -----+-----+-----+-----+-----+-----+-----+  
1320

AACCAATCTCAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACACCACAACCTGTTC

L G R C K R D D F V E G L S F A R S G G

25 460

TTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTTCGCTAGATCTGGTGGT

1321 -----+-----+-----+-----+-----+-----+-----+  
1380

AACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGATCTAGACCACCA

30

N W E E C F A \* 467

AACTGGGAAGAATGTTTCGCTTAA

1381 -----+-----+----- 1404

TTGACCCTTCTTACAAAGCGAATT

35



5

10

15

Figure 9

20

M G V F V V L L S I A T L F G S T S G T 20  
ATGGGGGTTTTTCGTCGTTCTATTATCTATCGCGACTCTGTTCGGCAGCACATCGGGCACT  
1 -----+-----+-----+-----+-----+ 60  
TACCCCCAAAAGCAGCAAGATAATAGATAGCGCTGAGACAAGCCGTCGTGTAGCCCGTGA

22 JAN. 1999

- 101 -

A L G P R G N H S K S C D T V D L G Y Q 40  
GCGCTGGGCCCCCGTGGAAATCACTCCAAGTCCTGCGATACGGTAGACCTAGGGTACCAG  
61 -----+-----+-----+-----+-----+-----+ 120  
5 CGCGACCCGGGGGCACCTTTAGTGAGGTTTCAGGACGCTATGCCATCTGGATCCCATGGTC  
  
C S P A T S H L W G T Y S P Y F S L E D 60  
TGCTCCCCCTGCGACTTCTCATCTATGGGGCACGTA CTGCCATaCTTTTCGCTCGAGGAC  
121 -----+-----+-----+-----+-----+-----+ 180  
10 ACGAGGGGACGCTGAAGAGTAGATACCCCGtgCATGAGCGGTAtGAAAAGCGAGCTCCTG  
  
E L S V S S K L P K D C R I T L V Q V L 80  
GAGCTGTCCGTGTCGAGTAAGCTTCCCAAGGATTGCCGGATCACCTTGGTACAGGTGCTA  
181 -----+-----+-----+-----+-----+-----+ 240  
15 CTCGACAGGCACAGCTCATTGGAAGGGTTCCTAACGGCCTAGTGGAACCATGTCCACGAT  
  
S R H G A R Y P T S S K S K K Y K K L I 100  
TCGCGCCATGGAGCGCGGTACCCAACCAGCTCCAAGAGCAAAAAGTATAAGAAGCTTaTt  
241 -----+-----+-----+-----+-----+-----+ 300  
20 AGCGCGGTACCTCGCGCCATGGGTTGGTTCGAGGTTCTCGTTTTTCATATCTTTCGAAtAa  
  
T A I Q A N A T D F K G K Y A F L K T Y 120  
ACGGCGATCCAGGCCAATGCCACCGACTTCAAGGGCAAGTAcGCCTTTTTGAAGACGTAC  
301 -----+-----+-----+-----+-----+-----+ 360  
25 TGCCGCTAGGTCCGTTACGGTGGCTGAAGTTCCCGTTCAtgCGGAAAAAATTCTGCATG  
  
N Y T L G A D D L T P F G E Q Q L V N S 140  
AACTATACTCTGGGTGCGGATGACCTCACTCCCTTTGGGGAGCAGCAGCTGGTGAACTCG  
361 -----+-----+-----+-----+-----+-----+ 420  
30 TTGATATGAGACCCACGCCTACTGGAGTGAGGGAAACCCCTCGTCGTCGACCACTTGAGC  
  
G I K F Y Q R Y K A L A R S V V P F I R 160  
GGCATCAAGTTCTACCAGAGGTACAAGGCTCTGGCGCGCAGTGTGGTGCCGTTTATTTCGC

22 JAN. 1999

- 102 -

421 -----+-----+-----+-----+-----+ 480  
CCGTAGTTCAAGATGGTCTCCATGTTCCGAGACCGCGCGTCACACCACGGCAAATAAGCG  
A S G S D R V I A S G E K F I E G F Q Q 180  
5 GCCTCAGGCTCGGACCGGGTTATTGCTTCGGGAGAGAAGTTCATCGAGGGGTTCCAGCAG  
481 -----+-----+-----+-----+-----+ 540  
CGGAGTCCGAGCCTGGCCCAATAACGAAGCCCTCTCTTCAAGTAGCTCCCCAAGGTCGTC  
A K L A D P G A T N R A A P A I S V I I 200  
10 GCGAAGCTGGCTGATCCTGGCGCGACGAACCGCGCCGCTCCGGCGATTAGTGTGATTATT  
541 -----+-----+-----+-----+-----+ 600  
CGCTTCGACCGACTAGGACCGCGCTGCTTGGCGCGGCGAGGCCGCTAATCACACTAATAA  
P E S E T F N N T L D H G V C T K F E A 220  
15 CCGGAGAGCGAGACGTTCAACAATACGCTGGACCACGGTGTGTGCACGAAGTTTGAGGCG  
601 -----+-----+-----+-----+-----+ 660  
GGCCTCTCGCTCTGCAAGTTGTTATGCGACCTGGTGCCACACACGTGCTTCAAACCTCCGC  
S Q L G D E V A A N F T A L F A P D I R 240  
20 AGTCAGCTGGGAGATGAGGTTGCGGCCAATTTCACTGCGCTCTTTGCACCCGACATCCGA  
661 -----+-----+-----+-----+-----+ 720  
TCAGTCGACCCTCTACTCCAACGCCGTTAAAGTGACGCGAGAAACGTGGGCTGTAGGCT  
A R L E K H L P G V T L T D E D V V S L 260  
25 GCTCGCctCGAGAAGCATCTTCCTGGCGTGACGCTGACAGACGAGGACGTTGTCAGTCTA  
721 -----+-----+-----+-----+-----+ 780  
CGAGCGgaGCTCTTCGTAGAAGGACCGCACTGCGACTGTCTGCTCCTGCAACAGTCAGAT  
M D M C P F D T V A R T S D A S Q L S P 280  
30 ATGGACATGTGTcCGTTTGATACGGTAGCGCGCACCAGCGACGCAAGTCAGCTGTCACCG  
781 -----+-----+-----+-----+-----+ 840

TACCTGTACACAgGCAAACTATGCCATCGCGCGTGGTCGCTGCGTTTCAGTCGACAGTGGC

F C Q L F T H N E W K K Y D Y L Q S L G 300

TTCTGTCAACTCTTCACTCACAATGAGTGGAGAAGTACgACTACCTTCAGTCCTTGGGC

5 841 -----+-----+-----+-----+-----+-----+ 900

AAGACAGTTGAGAAGTGAGTGTACTCACCTTCTTCATGcTGATGGAAGTCAGGAACCCG

K Y Y G Y G A G N P L G P A Q G I G F T 320

AAGTACTACGGCTACGGCGCAGGCAACCCTCTGGGACCGGCTCAGGGGATAGGGTTCCACC

10 901 -----+-----+-----+-----+-----+-----+ 960

TTCATGATGCCGATGCCGCGTCCGTTGGGAGACCCTGGCCGAGTCCCCTATCCCAAGTGG

N E L I A R L T R S P V Q D H T S T N S

340

AACGAGCTGATTGCCCCGTTGACgCGTTCCGCCAGTGCAGGACCACACCAGCACTAACTCG

15 961 -----+-----+-----+-----+-----+-----+ 1020

TTGCTCGACTAACGGGCCAACTGcGCAAGCGGTACGTCCTGGTGTGGTCGTGATTGAGC

T L V S N P A T F P L N A T M Y V D F S

20 360

ACTCTAGTCTCCAACCCGGCCACCTTCCC GTTGAACGCTACCATGTACGTCGACTTTTCA

1021 -----+-----+-----+-----+-----+-----+ 1080

TGAGATCAGAGGTTGGGCCGGTGGAAAGGGCAACTTGCGATGGTACATGCAGCTGAAAAGT

25

H D N S M V S I F F A L G L Y N G T E P

380

CACGACAACAGCATGGTTTCCATCTTCTTGCATTGGGCCTGTACAACGGCACTGAACCC

30 1081 -----+-----+-----+-----+-----+-----+ 1140

GTGCTGTTGTCGTACCAAAGGTAGAAGAAACGTAACCCGGACATGTTGCCGTGACTTGGG

L S R T S V E S A K E L D G Y S A S W V

35 400

22 JAN. 1959

- 104 -

TTGTCCCGGACCTCGGTGGAAAGCGCCAAGGAATTGGATGGGTATTCTGCATCCTGGGTG  
 1141 -----+-----+-----+-----+-----+  
 1200  
 AACAGGGCCTGGAGCCACCTTTCGCGGTTCTTAACCTACCCATAAGACGTAGGACCCAC  
 5  
 420 V P F G A R A Y F E T M Q C K S E K E P  
 GTGCCTTTCGGCGCGCGAGCCTACTTTCGAGACGATGCAATGCAAGTCGGAAAAGGAGCCT  
 1201 -----+-----+-----+-----+-----+  
 10 1260  
 CACGGAAAGCCGCGCGCTCGGATGAAGCTCTGCTACGTTACGTTACGCTTTTCCTCGGA  
 440 L V R A L I N D R V V P L H G C D V D K  
 15 CTTGTTTCGCGCTTTTGATTAATGACCGGGTTGTGCCACTGCATGGCTGCGATGTGGACAAG  
 1261 -----+-----+-----+-----+-----+  
 1320  
 GAACAAGCGCGAAACTAATTACTGGCCCAACACGGTGACGTACCGACGCTACACCTGTTC  
 20 460 L G R C K L N D F V K G L S W A R S G G  
 CTGGGGCGATGCAAGCTGAATGACTTTGTCAAGGGATTGAGTTGGGCCAGATCTGGGGGC  
 1321 -----+-----+-----+-----+-----+  
 1380  
 25 GACCCCGCTACGTTGACTTACTGAAACAGTTCCTTAACCTCAACCCGGTCTAGACCCCGG  
 N W G E C F S \* 467  
 AACTGGGGAGAGTGCTTTAGTTGA  
 1381 -----+-----+----- 1404  
 30 TTGACCCCTCTCACGAAATCAACT

Figure 10

CP-1

5           Eco RI M G V F V V L L S I A T L F G S T  
TATATGAATTCATGGGCGTGTTCGTCTGCTACTGTCCATTGCCACCTTGTTCCGGTTCCA  
1 -----+-----+-----+-----+-----+-----+ 60  
ATATACTTAAGTACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGT

10           S G T A L G P R G N S H S C D T V D G G  
CATCCGGTACCGCCTTGGGTCCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGTG  
61 -----+-----+-----+-----+-----+-----+ 120  
GTAGGCCATGGCGGAACCCAGGAGCACCATTAAAGAGTGAGAACACTGTGACAACTGCCAC

CP-2

15           CP-3  
Y Q C F P E I S H L W G Q Y S P Y F S L  
GTTACCAATGTTTCCCAGAAATTTCTCACTTGTGGGGTCAATACTCTCCATACTTCTCTT  
121 -----+-----+-----+-----+-----+-----+ 180  
CAATGGGTTACAAAGGGTCTTTAAAGAGTGAACACCCCAAGTTATGAGAGGTATGAAGAGAA

20           E D E S A I S P D V P D D C R V T F V Q  
TGGAAGACGAATCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTCTGTTT  
181 -----+-----+-----+-----+-----+-----+ 240  
ACCTTCTGCTTAGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAG

25           CP-4.7  
            CP-5.7  
V L S R H G A R Y P T D S K G K K Y S A  
AAGTTTGTCTAGACACGGTGCTAGATACCCAAGTgacTCTAAGggtAAGaagTACTCTG  
241 -----+-----+-----+-----+-----+-----+ 300  
30           TTCAAAACAGATCTGTGCCACGATCTATGGGTTGActgAGATTCCAATTCttCATGAGAC

Modtaget PD  
22 JAN. 1999

- 106 -

L I E A I Q K N A T A F K G K Y A F L K  
CTTTGATTGAAGCTATTCAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGA

301 -----+-----+-----+-----+-----+-----+ 360

5 GAAACTAACTTCGATAAGTTTTCTTGCGATGACGAAAGTCCCATTTCATGCGAAAGAAGT

CP-6

CP-7

T Y N Y T L G A D D L T P F G E N Q M V  
AGACTTACAACCTACTTTGGGTGCTGACGACTTGACTCCATTCCGGTGAAAACCAAATGG

10 361 -----+-----+-----+-----+-----+-----+ 420

TCTGAATGTTGATGTGAACCCACGACTGCTGAACTGAGGTAAGCCACTTTTGGTTTACC

N S G I K F Y R R Y K A L A R K I V P F  
TTAACTCTGGTATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCAT

15 421 -----+-----+-----+-----+-----+-----+ 480

AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTA

CP-8.7

CP-9

I R A S G S S R V I A S A E K F I E G F  
TCATTAGAGCTTCTGGTTCTTctAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTT

20 481 -----+-----+-----+-----+-----+-----+ 540

AGTAATCTCGAAGACCAAGAagaTCTCAATAACGAAGACGACTTTTCAAGTAACTTCCAA

Q S A K L A D P G S Q P H Q A S P V I D  
TCCAATCTGCTAAGTTGGCTGACCCAGGTTCTCAACCACACCAAGCTTCTCCAGTTATTG

25 541 -----+-----+-----+-----+-----+-----+ 600

AGGTTAGACGATTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTCTGAAGAGGTCAATAAC

CP-10.7

CP-11.7

V I I S E A S S Y N N T L D P G T C T A  
ACGTTATTATTtctGAcgctTCTtctTACAACAACACTTTGGACccaGGTACTTGTACTG

30 601 -----+-----+-----+-----+-----+-----+ 660

Modtaget 12

22 JAN. 1999

- 107 -

TGCAATAATAAagaCTgcgaAGGagaATGTTGTTGTGAAACCTGggtCCATGAACATGAC



Modtagei P.  
22 JAN. 1959

- 108 -

F E D S E L A D T V E A N F T A L F A P

CTTTCGAAGACTCTGAATTGgctGACactGTTGAAGCTAACTTCACTGCTTTGTTTCGCTC

661 -----+-----+-----+-----+-----+ 720

GAAAGCTTCTGAGACTTAACcgaCTGtgaCAACTTCGATTGAAGTGACGAAACAAGCGAG

5

CP-12.7

A I R A R L E A D L P G V T L T D T E V

CAGCTATTAGAGCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGACactgaaG

721 -----+-----+-----+-----+-----+ 780

10

GTCGATAATCTCGATCTAACCTTCGACTGAACGGTCCACAATGAACTGACTGtgacttc

CP-13.7

T Y L M D M C S F E T V A R T S D A T E

TTactTACTTGATGGACATGTGTtctTTCGAAACTGTTGCTAGAACTTCTGACGCTACTG

15

781 -----+-----+-----+-----+-----+ 840

AatgaATGAACTACCTGTACACAagaAAGCTTTGACAACGATCTTGAAGACTGCGATGAC

L S P F C A L F T H D E W R H Y D Y L Q

AATTGTCTCCATTCTGTGCTTTGTTCACTCACGACGAATGGAGAcacTACGACTACTTGC

20

841 -----+-----+-----+-----+-----+ 900

TTAACAGAGGTAAGACACGAAACAAGTGAGTGCTGCTTACCTCTgtgATGCTGATGAACG

CP-14.7

CP-15.7

S L K K Y Y G H G A G N P L G P T Q G V

25

AATCTTTGaagAAGTACTACGGTcacGGTGCTGGTAACCCATTGGGTCCAactCAAGGTG

901 -----+-----+-----+-----+-----+ 960

TTAGAAActtcTTCATGATGCCAgtgCCACGACCATTGGGTAACCCAGGTtgaGTTCCAC

G F A N E L I A R L T R S P V Q D H T S

30

TTGGTTTCGCTAACGAATTGATTGCTAGATTGACTAGATCTCCAGTTCAAGACCACACTT

961 -----+-----+-----+-----+-----+ 1020

AACCAAAGCGATTGCTTAACTAACGATCTAACTGATCTAGAGGTCAAGTTCTGGTGTGAA

Modtaget -  
22 JAN. 1993

- 109 -

CP-16

CP-17.7

T N H T L D S N P A T F P L N A T L Y A  
CTACTAACCACACTTTGGACTCTAACCAGCTACTTTCCCATTTGAACGCTACTTTGTACG  
5 1021 -----+-----+-----+-----+-----+-----+  
1080

GATGATTGGTGTGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCGATGAAACATGC  
D F S H D N G I I S I F F A L G L Y N G  
10 CTGACTTCTCTCAGACAACggtattATTCTATTTTCTTCGCTTTGGGTTTGTACAACG  
1081 -----+-----+-----+-----+-----+-----+  
1140

GACTGAAGAGAGTGCTGTTGccataaTAAAGATAAAAGAAGCGAAACCCAAACATGTTGC  
CP-18.7  
15 CP-19.7

T A P L S T T S V E S I E E T D G Y S S  
GTACTGCTCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTt  
1141 -----+-----+-----+-----+-----+-----+  
1200

CATGACGAGGTAACAGATGATGAAGACAACCTTAGATAACTTCTTTGACTGCCAATGAGAA  
A W T V P F A S R A Y V E M M Q C Q A E  
ctgctTGGACTGTTCCATTTCgtttctAGAGCTTACGTTGAAATGATGCAATGTCAAGCTG  
1201 -----+-----+-----+-----+-----+-----+  
25 1260

gacgaACCTGACAAGGTAAGcgaagaTCTCGAATGCAACTTTACTACGTTACAGTTTCGAC

CP-20

CP-21

K E P L V R V L V N D R V V P L H G C A  
30 AAAAGGAACCATTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG  
1261 -----+-----+-----+-----+-----+-----+  
1320

TTTTCCTTGGTAACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACAC

Modtaget PD  
22 JAN. 1999

- 110 -

V D K L G R C K R D D F V E G L S F A R

CTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTTCGCTA

1321 -----+-----+-----+-----+-----+-----+  
1380

5 GACAACTGTTCAACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGAT

S G G N W A E C F A \* Eco RI CP-22

GATCTGGTGGTAACTGGGCTGAATGTTTCGCTTAAGAATTCATATA

1381 -----+-----+-----+-----+-----+----- 1426

10 CTAGACCACCATTGACCCGACTTACAAAGCGAATTCTTAAGTATAT

Figure 11

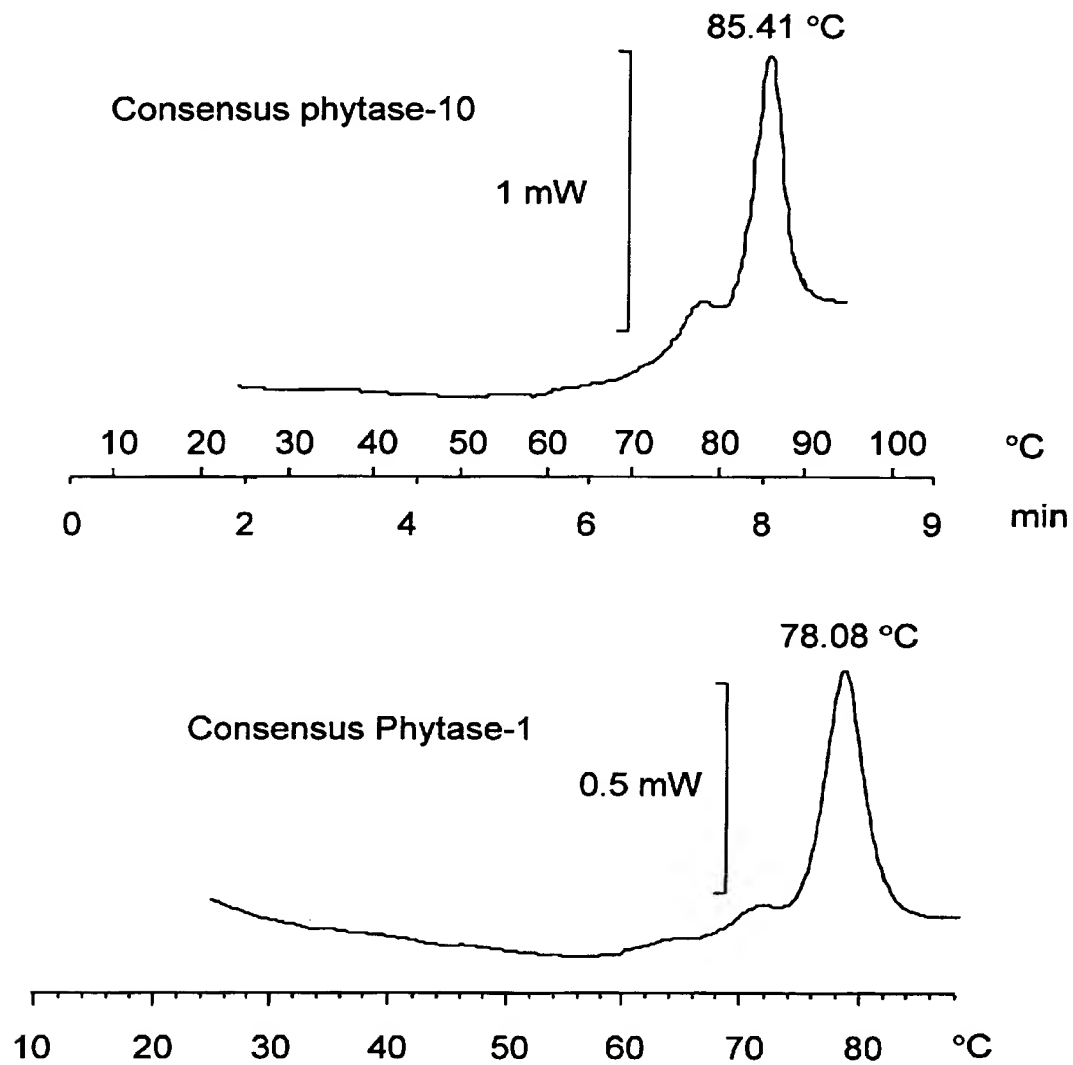


Figure 12

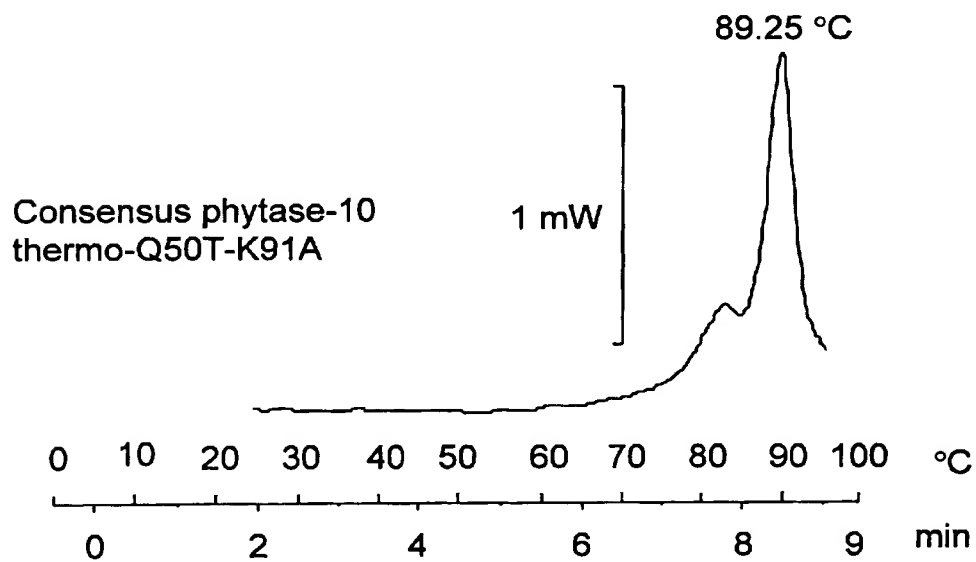
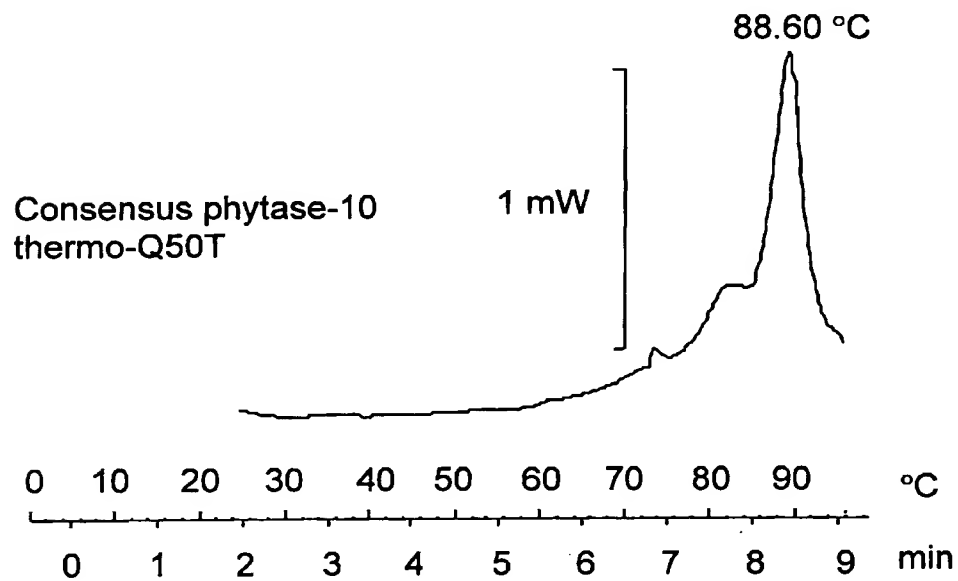


Figure 13

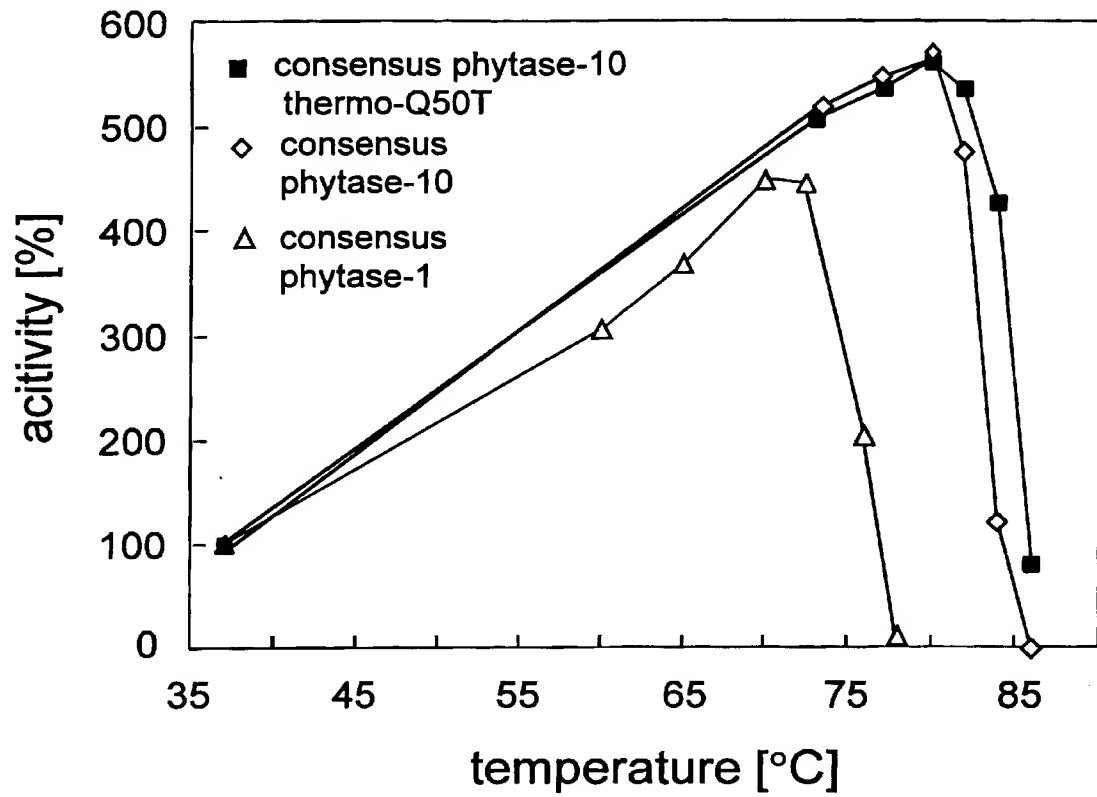


Figure 14

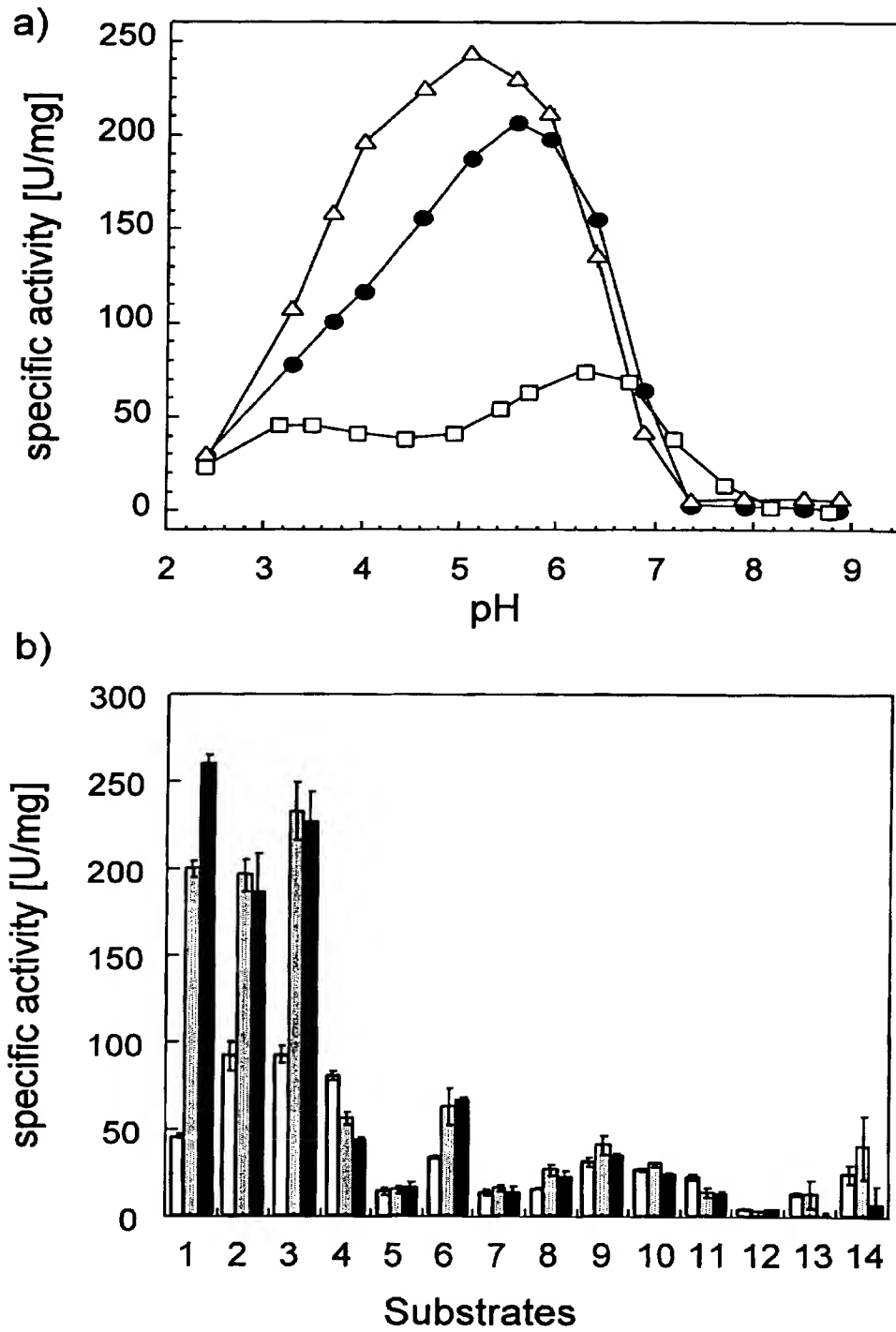


Figure 15

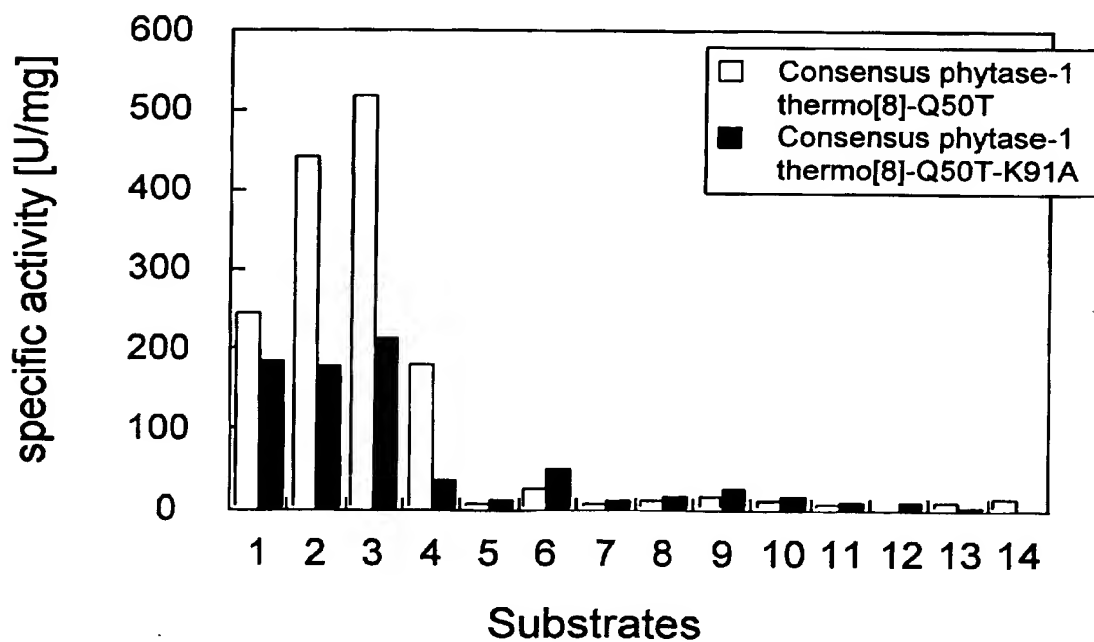
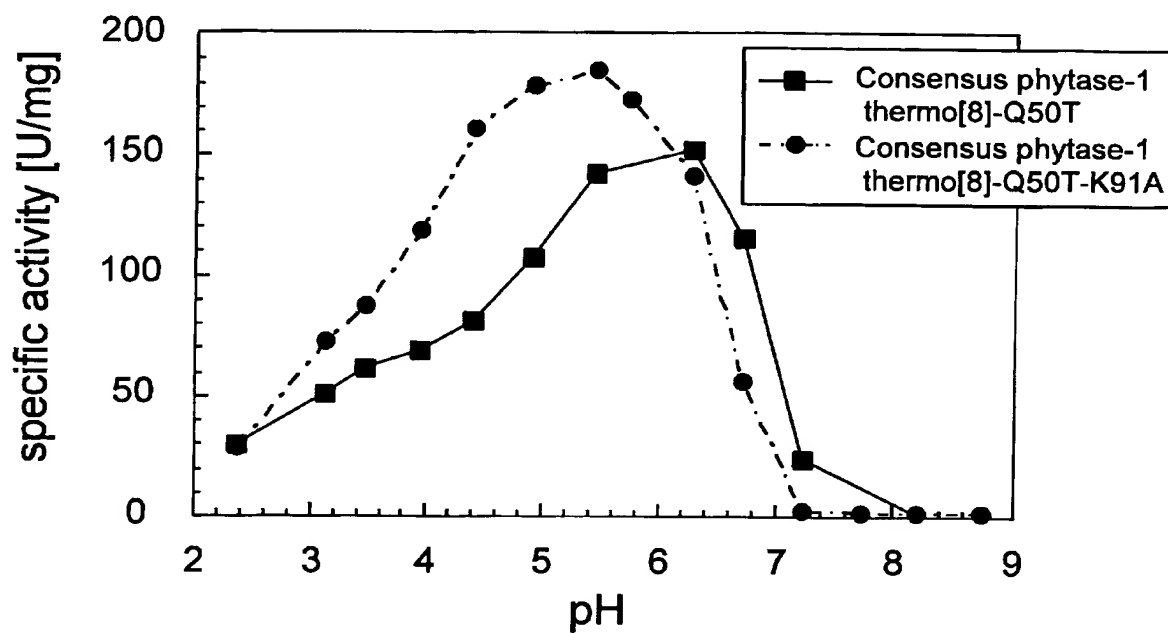




Figure 16

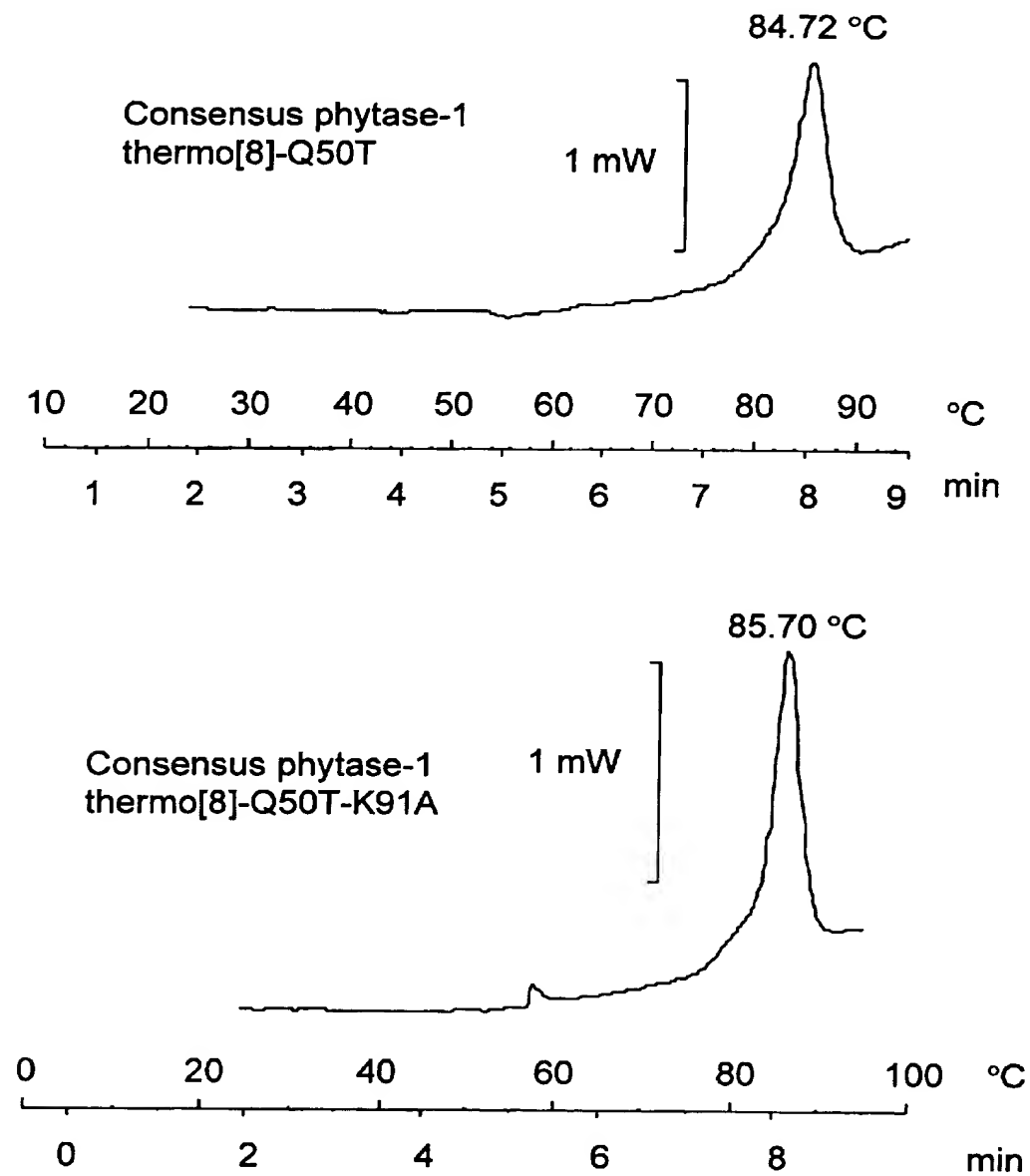


Figure 17

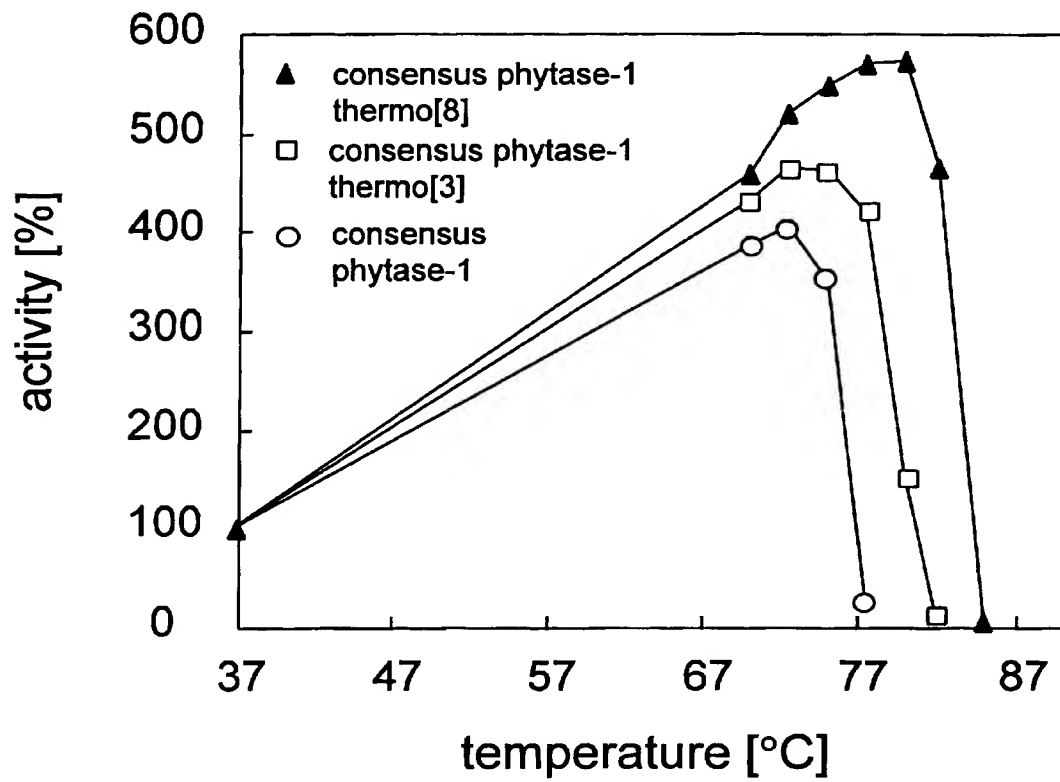
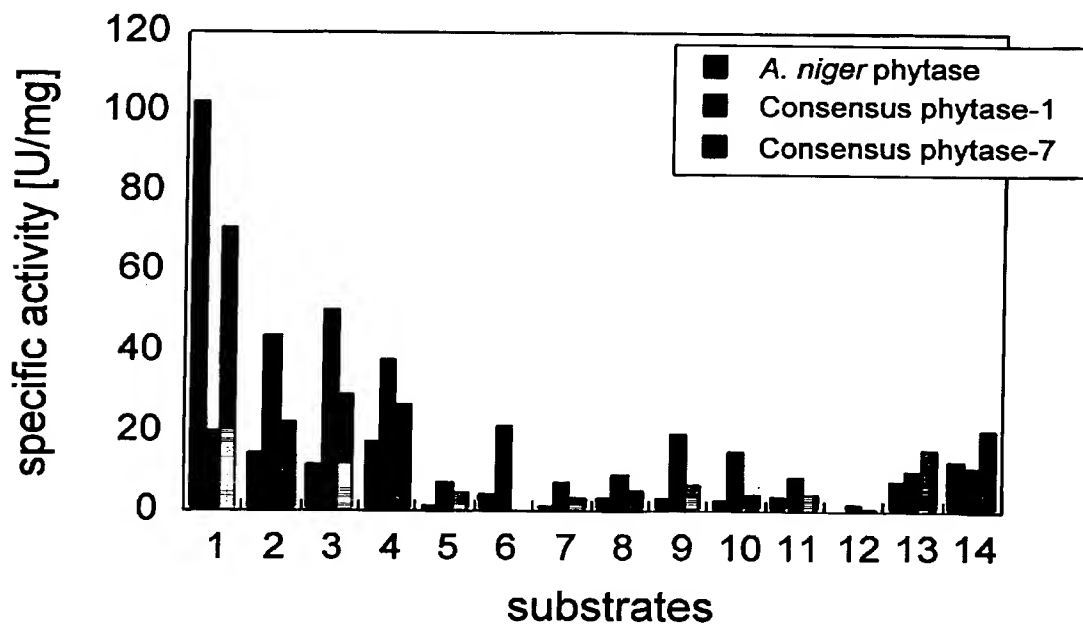
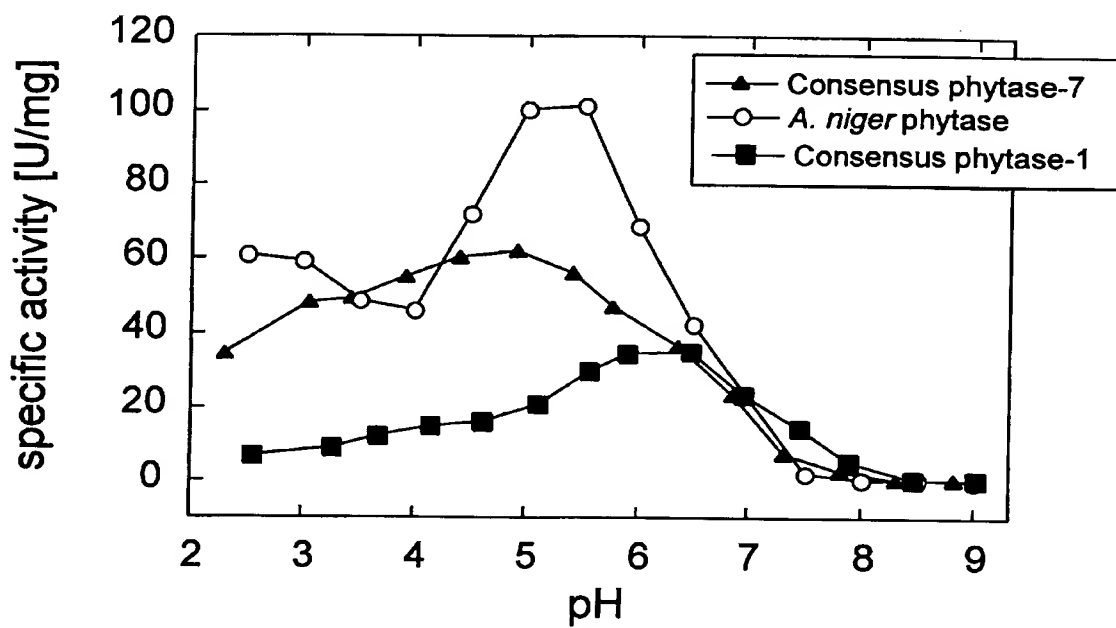


Figure 18



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Figure 19

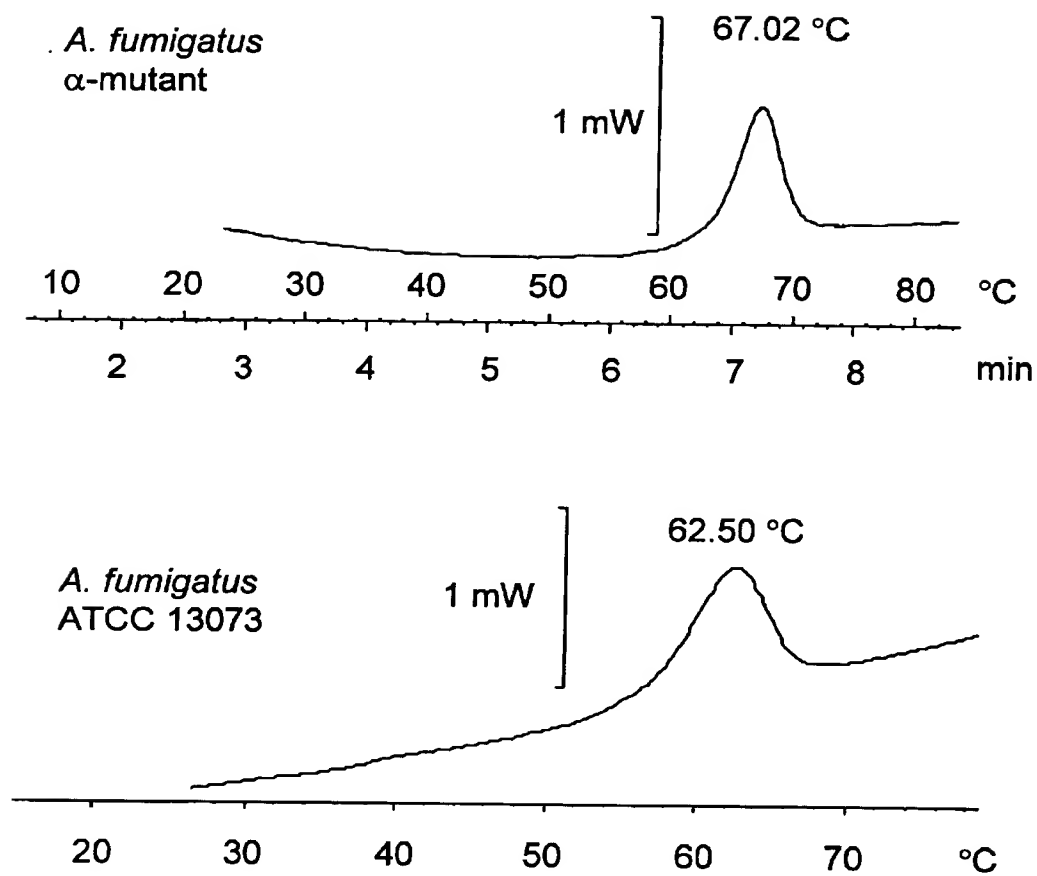
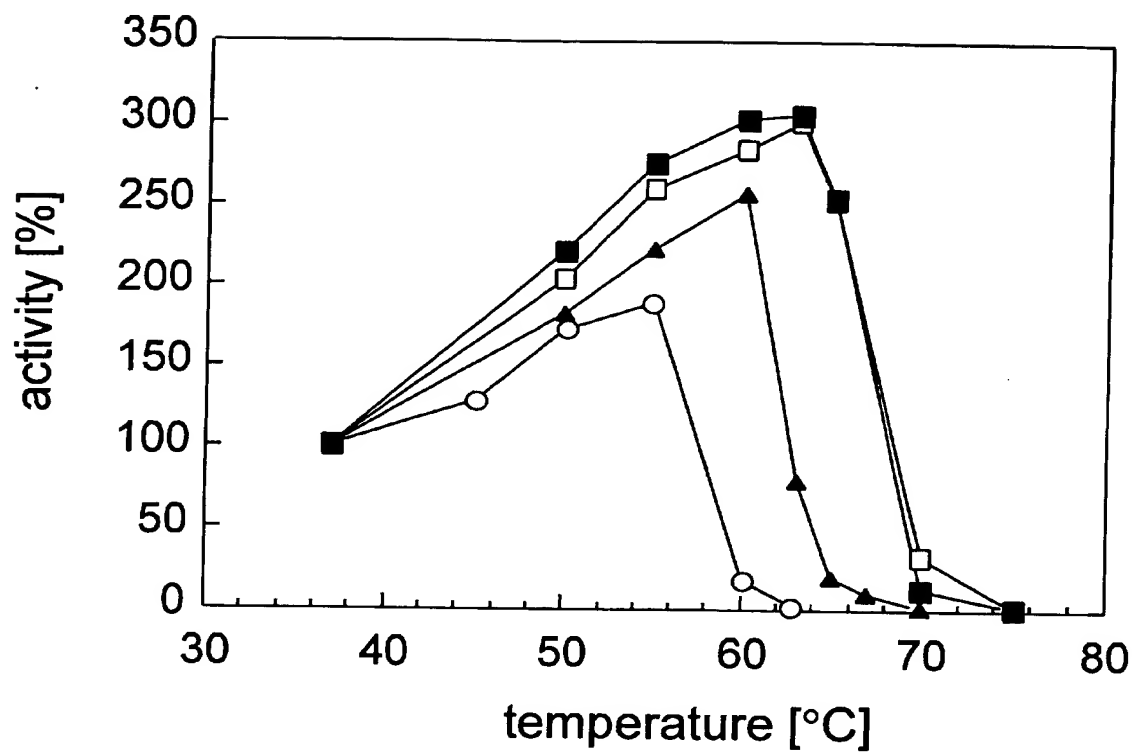


Figure 20



Modtaget PD  
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Figure 21

1 MGVFVLLSI ATLFGSTSGT ALGPRGNSHS CDTVDDGGYQC FPEISSNWSP  
51 YSPYFSLADE SAISPDVPKG CRVTFVQVLQ RHGAREFPTSG AATRISALIE  
101 AIQKNATAFK GKYAFLKTYN YTLGADDLVP FGANQSSQAG IKFYRRYKAL  
5 151 ARKIVPFIRA SGSDRVIDSA TNWIEGFQSA KLADPGANPH QASPVINVII  
201 PEGAGYNNTL DHGLCTAFEE SELGDDVEAN FTAVFAPPIR ARLEAHLPGV  
251 NLTDDEVVNL MDMCPFDIVA RTSDATELSP FCDLFTHDEW IQDYDLGDL  
301 KYYGTGAGNP LGPAQGVGVFV NELIARLTHS PVQDHTSTNH TLDSNPATFP  
351 LNATLYADFS HDNTMVAIFF ALGLYNGTKP LSTTSVESIE ETDGYSASWL  
10 401 VPFSARMYVE MMQCEAEKEP LVRVLVNDRV VPLHGCGVDK LGRCKRDDFV  
451 EGLSFARSGG NWEECFA

Abstract

This invention relates to a new improved consensus phytase by introduction of  
 5 additional phytase sequences into the sequence alignment and the method of  
 the introduction process. Furthermore, the invention relates to the transfer of  
 stabilizing amino acid exchanges found by the new method into homologous  
 proteins. Furthermore, the invention relates to the replacement of a whole  
 active site of a phytase. It also relates to the corresponding DNA sequences  
 10 and its generation, methods to produce such phytases and the use thereof.

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